
1 Cardiac Effects of (?)-Epigallocatechin on Isolated Rat Hearts

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3 *Received: 6 December 2017 Accepted: 31 December 2017 Published: 15 January 2018*

4

5 **Abstract**

6 (?)- Epigallocatechin is a flavonoid found in many plants, especially in tea. The consumption
7 of flavonoid- rich foods tends to reduce the risk of cardiovascular diseases and this has been
8 attributed to nonspecific activities such as antioxidant, anti- atherosclerotic and
9 anti-inflammatory properties. However, little is known about direct actions of (?)-
10 epigallocatechin on cardiac muscle. The aim of the present investigation was to evaluate the
11 effects of (?)- epigallocatechin on electrical and contractile activities of isolated rat hearts.
12 Surface electrogram and force of contraction were recorded in isolated rat hearts in control
13 and in increasing concentrations of (?)- epigallocatechin from 0.001 to 3 ?M.
14 (?)-Epigallocatechin tended to prolong the QRS interval, but this effect is significant only at
15 the highest concentration studied (3 ?M). QTc was not significantly affected by the flavonoid.
16 The effects of this flavonoid on RR interval were mild and statistically significant since 0.03
17 ?M. (?)- Epigallocatechin produced a negative inotropic effect in isolated rat hearts with an
18 IC₅₀ of 0.03 ?M. This flavonoid has direct actions on rat cardiac muscle.

19

20 *Index terms—*

21 **1 Introduction**

22 catechins are one group of natural polyphenols found in many plants, especially in green tea (leaves of *Camellia*
23 *sinensis*) (1)(2)(3). The four main catechin derivatives mainly find in green tea include the isomers epicatechin,
24 (-)-epicatechingallate (ECG), (-)-epigallocatechin (EGC), and (-)-epigallocatechin gallate (EGCG) (3). EGC is a
25 flavan-3-ol containing a benzopyran-3,5,7-triol linked to a 3,4,5-hydroxyphenyl moiety. Thus, EGC is considered
26 to be a flavonoid lipid molecule (4) (Figure 1).

27 The health benefits associated with the consumption of green tea are due to the activity of EGCG and EGC
28 which are both present at higher amounts (5). EGC has many beneficial cardiovascular properties. However,
29 most of these effects are nonspecific, such as antioxidant (1-2, 6-7), antiinflammatory (1,5,7), and antiatherogenic
30 activities (8).

31 Another remarkable property attributed to tea catechins is the cholesterol-lowering action, involving the
32 upregulation of the LDL receptor, the reduction of cholesterol absorption, and the modulation of both synthetic
33 and metabolic pathways (see for review 9).

34 **2 Further**

35 investigations of the cellular mechanisms are needed to investigate the cardiovascular effects of this flavonoid.
36 Other flavonoids such as naringenin, quercetin, and genistein have direct actions on rat cardiac and vascular
37 smooth muscles (10). The present work evaluated the possible direct effects of EGC on electrical and contractile
38 activities of rat isolated rat hearts.

8 RESULTS AND DISCUSSION

39 3 II.

40 4 Materials and Methods

41 5 a) Animals

42 Male adult (7-8 weeks) Wistar rats were brought from the National Center for Laboratory Animal Reproduction
43 (CENPALAB; La Habana). Before the experiments, animals were for seven days adapted to laboratory conditions
44 (controlled temperature $25 \pm 2^\circ\text{C}$, relative humidity $60 \pm 10\%$, and 12 h light/dark cycles). Tap water and
45 standard diet for rodents supplied by CENPALAB were freely provided. All procedures fulfilled with the European
46 Commission for the use and care of laboratory animals. The Committee for Animal Care in Research of the Center
47 (No. 08-2012, folio 73, book 01, 2012) approved the present study.

48 6 b) Isolated hearts

49 As previously reported (11), under pentobarbital anesthesia rat hearts were removed and placed in cold Tyrode
50 (see below). Hearts were carefully dissected, mounted on a Langendorff column and perfused at constant flow (10
51 mL/min) with a Tyrode solution of the following composition (mmol/L): 140 NaCl, 2.5 KCl, 0.5 MgCl₂, 2 CaCl₂,
52 10 Tris-hydroxymethyl amino methane, 10 Glucose (pH =7.4, gassed with O₂; T = 35°C). On the ventricular
53 epicardium was placed a bipolar platinum recording electrode to record the surface electrocardiogram. Another
54 bipolar platinum electrode was placed near the atrioventricular ring and was connected to an electronic stimulator.
55 To record the force of contraction (FC), the cardiac apex was fixed to a force-displacement transducer with a
56 surgical 6-0 silk thread. Surface electrocardiogram and FC values were recorded at the heart rate and a fixed
57 stimulus rate (500ms RR interval).

58 7 c) ECG and chemicals

59 Stock solutions of ECG were prepared in ethanol, and diluted in the bathing solution on the day of the experiment.
60 All chemicals were from Sigma Aldrich. Means and standard errors of means expressed the results. Student's
61 t-test evaluated the statistical significance for paired samples, previously checked that the data complied with
62 the premise of normality. Differences were considered statistically significant for $p < 0.05$. The graphics and the
63 statistical processing were done using the software OriginPro 8 SRO v8.0724 (MA, USA).

64 8 Results and Discussion

65 The corrected QT (QTc) interval of the surface electrocardiogram (QTc = QT/?RR) was not significantly affected
66 by EGC at concentrations from 0.001 to 3 μM (Table 1).

67 These results should be possible because this flavonoid could exert multiple actions on different ionic channels,
68 resulting in an apparent absence of effects on QT interval of the cardiac surface electrogram. As a fat, catechins
69 modulate several ionic channels (12)(13)(14)(15).

70 EGC showed a tendency to increase QRS interval of the surface electrocardiogram, but only at the highest
71 concentration studied (3 μM) this increase was statistically significant ($p < 0.05$) (Table 1). EGCG, catechin
72 structurally related to EGC, at 30 μM prolonged QRS interval in isolated spontaneously beating guinea pig hearts
73 (15). The QRS wave is dependent on sodium channel activity, Kang et al., 2010 showed that EGCG inhibited
74 the cloned human cardiac sodium channel Nav1.5 in a dose-dependent manner with $45.7 \pm 6.9\%$ inhibition at
75 100 μM (15). EGCG reduced the amplitude of voltage-gated sodium channel current in a concentration-depend
76 manner in the range of 0.1 -400 μM in rat hippocampal CA1 neurons (13).

77 On the other hand, EGC prolonged the RR interval of surface electrocardiogram and this increase was
78 statistically significant ($p < 0.05$) since 0.03 μM (Table 1).

79 EGCG at 30 μM did not affect heart rate of guinea pig hearts (15). Green tea extract used with dietary
80 supplements did not alter heart rate (16). Other study concluded that *Camellia sinensis* has effect on heart rate,
81 it decreases the heart rate in normotensive female individuals and increases the heart rate in the normotensive
82 male individuals (17). In the present study in the concentration range from 0.001 to 3 μM , EGC significantly
83 decreased the force of contraction (FC) in isolated rat hearts (Figure 2); concentrations as low as 0.001 μM of
84 EGC decreased FC by $28.4 \pm 8.7\%$. Since EGC slightly changed RR interval, hearts were paced at 500-ms
85 stimulus interval (over the spontaneous RR interval under control condition; 531.05 ± 18.9 ms) to avoid any
86 frequencydependent change in FC. Experimental data were fitted to a Hill function (Figure 2), and the estimated
87 IC50 for inhibition of contraction was 0.03 ± 7.8 μM for EGC. The action of EGC on FC was not reversible upon
88 washout with the normal Tyrode solution. Although further studies are needed to see if EGC has any direct
89 effect on calcium channels, the decrease of force of cardiac contraction by EGC should be at least partly due to
90 an inhibition of calcium channels.

91 The L-type calcium channel was inhibited by 20.8% at 30 μM by EGCG, reached a maximum of $37.1 \pm 4.2\%$
92 at a concentration of 100 μM (15). Tadano et al., 2010 reported that EGC had no significant effects on cardiac
93 myofilament Ca²⁺-sensitivity. However ECG and EGCG were found to decrease Ca²⁺ sensitivity, they were
94 Ca²⁺ desensitizers acting through binding to cardiac troponin C (18).

95 At concentrations within the same range at which similar flavonoid EGCG have vasorelaxant effects related to
96 the inhibition of Ca²⁺ influx in smooth muscle cells (19), in the present results, EGC concentrationdependently
97 relaxed with almost equal effectiveness the contraction of rat hearts.

98 On the strength of these results, the physiological relevance of the decrease of force of cardiac contraction
99 by EGC can be asserted by considering the data available on the in vivo level of the related catechin EGCG
100 ([EGCG] = 0.3-7.5 μ M in the blood of green tea consumers (20).

101 Three-month supplementation with green tea capsules decreased systolic (SBP) and diastolic blood pressure
102 (DBP) by four mmHg in obese hypertensive (21) but not obese subjects (22). A recent metaanalysis which
103 included eleven trials concluded that short-term consumption (>6 months) of black tea could decrease SBP and
104 DBP by 1-2 mmHg and green tea by three mmHg (23).

105 IV.

106 9 Conclusions

107 The present study revealed that EGC has direct cardiac effects. The results presented here con firm the role of
108 tea catechin EGC, as a precursor for the development of novel drugs for the treatment of cardiovascular disorders.

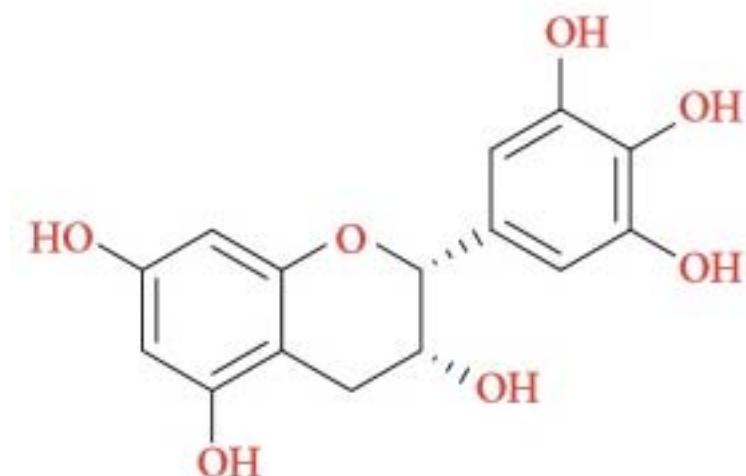
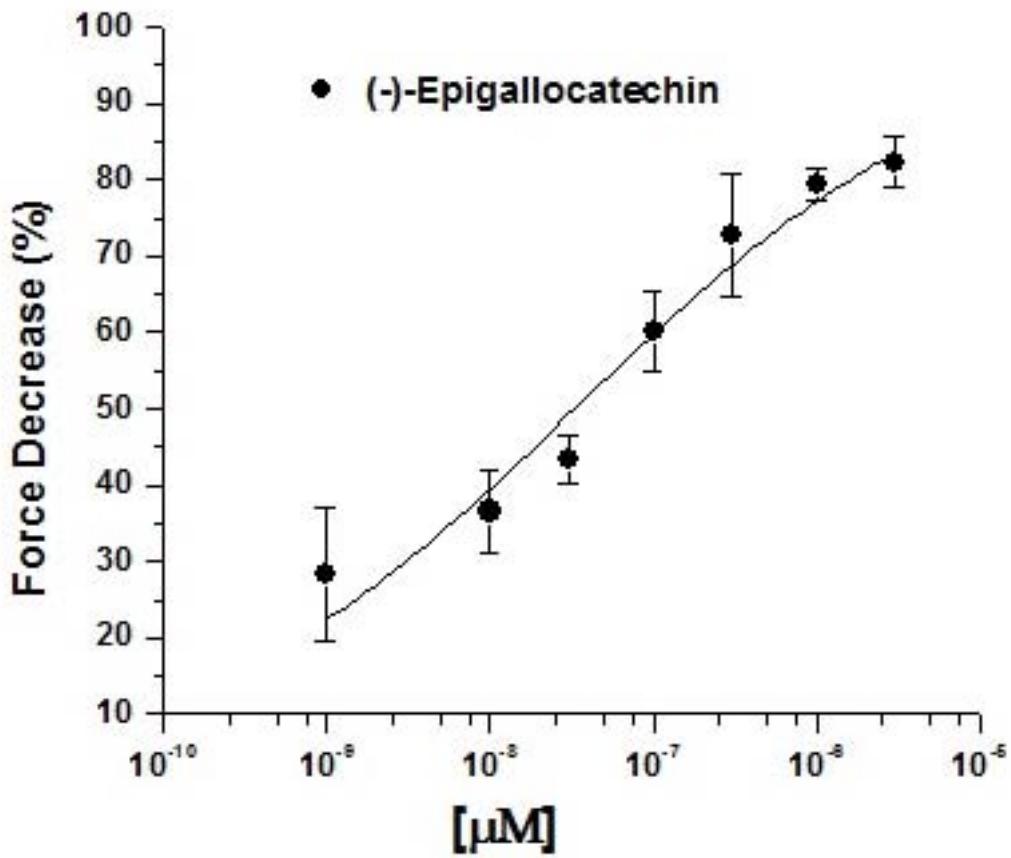


Figure 1: Figure 1 :

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109 1



2

Figure 2: Figure 2 :

1

	QTc (mseg)	p	QRS (mseg)	p	RR (mseg)	p
Control	88.55 ± 7.2		11.80 ± 0.7		531.05 ± 18.9	
EGC 0.001 ?M	84.20 ± 4.7	0.71	12.50 ± 0.1	0.36	541.48 ± 20.2	0.72
EGC 0.003 ?M	98.01 ± 7.1	0.46	12.65 ± 0.2	0.28	552.20 ± 20.1	0.47
EGC 0.01 ?M	84.20 ± 11.2	0.74	12.85 ± 0.3	0.22	605.63 ± 41.4	0.15
EGC 0.03 ?M	98.70 ± 0.3	0.39	13.20 ± 0.3	0.12	639.13 ± 37.5 *	0.04
EGC 0.1 ?M	90.02 ± 10.0	0.91	13.30 ± 0.2	0.09	669.50 ± 30.1 *	0.008
EGC 0.3 ?M	86.50 ± 5.5	0.86	13.40 ± 0.3	0.08	676.50 ± 33.4 *	0.009
EGC 1 ?M	87.40 ± 6.7	0.91	13.60 ± 0.3	0.05	682.78 ± 33.4 *	0.008
EGC 3 ?M	95.10 ± 13.1	0.65	13.78 ± 0.3 *	0.04	678.05 ± 34.8 *	0.009

p < 0.05 vs.
Control

Figure 3: Table 1 :

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9 CONCLUSIONS

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