

CrossRef DOI of original article:

1 Antihyperlipidemic Property of a Dietary Supplement of Moringa 2 Oleifera Leaves and Pleurotus Ostreatus in Wistar Rats Stressed 3 by Combination of Ethanol-Paracetamol

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7 *Received: 1 January 1970 Accepted: 1 January 1970 Published: 1 January 1970*

9 Abstract

10 High amounts of triglycerides and cholesterol in the blood result in the metabolic condition
11 known as hyperlipidemia. There is currently no specific therapy to reduce the effects of this
12 disorder. In underdeveloped nations, metabolic diseases are treated using Moringa oleifera and
13 Pleurotus ostreatus. Both the nutritional and therapeutic benefits of these two plants are
14 frequently utilized. Purpose: This study aims to investigate the antihyperlipidemic property of
15 dietary supplement of Moringa oleifera leaves and Pleurotus ostreatus in wistar rats.
16 Materials and methods: A variety of mushroom species were produced in the Mushroom
17 Biotechnology Laboratory, and M. oleifera was developed in the university's botanical garden
18 in Dakar, Senegal.

20 **Index terms**— moringa oleifera, pleurotus ostreatus, dietary supplement, antihyperlipidemic, oxidative
21 stress.

22 1 Introduction

23 Alcoholism and other serious health issues are brought on by excessive alcohol usage, including alcoholic liver
24 damage (ALD). Alcoholism has been linked to several illnesses, and it is currently one of the most challenging
25 health issues with substantial medical, social, and economic repercussions. ??Pari and Karthikesan, 2001;.
26 Alcohol abuse leads to significant illnesses such as hyperglycemia, cirrhosis, cardiovascular disease, pancreatic
27 inflammation, and alcoholic fatty liver. (Ponnappa et al., 2000). Oxidative stress is one of the elements that
28 are crucial in numerous pathways of alcohol-induced harm. The creation of ROS in our bodies is abnormally
29 increased by our unhealthy eating habits and our way of life (smoking, drinking, obesity, and strenuous activity).
30 When organisms experience oxidative stress brought on by free radical damage, antioxidants aid in coping.
31 Antioxidant defenses come from two different sources: the diet, which includes fruits and vegetables, which are
32 rich in vitamins C and E, carotenoids, ubiquinone, polyphenols, and lipoic acid. The other is endogenous and
33 is made up of proteins, enzymes, or tiny molecules such as glutathione, uric acid, superoxide dismutase, and
34 glutathione peroxidase (ferritin, transferrin, etc.). Additionally, some elements that are significant cofactors
35 include selenium, copper, and zinc. ??Pincemail et al., 2009).

36 We were particularly interested in the plant Moringa oleifera and the edible fungus Pleurotus ostreatus because
37 they contain significant antioxidant content. The plant Moringa oleifera Lam. (Moringaceae), also known as
38 Nebeday in Senegal, is of Indian ancestry and is now common throughout Asia and Africa. The leaves are
39 utilized in traditional African medicine and are commonly consumed as a legume. They are an excellent source
40 of protein (19-35% dry matter) ??Kane et al., 2017; Makkar et al., (1996); Abou-Elezz et al., ??2012) and are
41 rich in metabolizable energy (2273-2978 kcal/kg DM) (Makkar et al., (1996); Olugbemi et al., 2010). They A
42 11.2% dry Matter). The South African ecotype of the plant has been observed to contain 19.3% crude protein

43 (Moyo., 2011). Traditional Chinese medicine uses *M. oleifera* leaves to treat diabetes, headaches, fever, and
44 malnourishment (Ndong et al., 2007; ??errorho., 1994).

45 Preview studies have shown the health and nutritional interest of edible mushrooms (Zhang et al., 2016;Alam
46 et al., 2008;Pornariya and Kanok, 2009). *P. ostreatus* has been demonstrated for its antitumor effects, antioxidant
47 properties, antihyperlipidemic effects and antidiabetic effects, (Zhang et al., 2016;Abrams et al., 2011;Alam et
48 al., 2008;Elmastas et al., 2007;Jayakumar et al., 2007;Jayakumar et al., 2006). A daily intake of 15 g of dried
49 oyster mushrooms would have an anti-hyperlipidemic effect on the subjects, it would also cover up to 50% of
50 the recommended daily intakes of macronutrients and minerals, according to research on *Pleurotus ostreatus*
51 nutritional value and antihyperlipidemic effects on HIV-positive individuals taking ARVs (Abrams et al., 2011,
52 Alam et al., 2008; ??anzi et al., 2001; ??lam et al., 2009; ??anzi et al., 1999 ?? Kane et al., 2017).

53 Given the rich nutrient, phytochemical, and organoleptic potential of *M.oleifera* and *P.ostreatus*, we designed
54 this study to determine the antihyperlipidemic effect of *Moringa oleifera* leaves and *Pleurotus ostreatus* in Wistar
55 rats stressed by a combination of Ethanolparacetamol. In this paper, we will code the dietary supplement by
56 FMP16.

57 2 II.

58 3 Materials and Methods

59 4 a) Plant Material and Preparation of the mixture of Leaves 60 from *M. oleifera* and *P. ostreatus*

61 The fresh leaves of *M. oleifera* were harvested at the botanical garden of the University Cheikh Anta Diop (UCAD)
62 of Dakar, Senegal and identified at the botanical department (UCAD).The leaves were cleaned immediately after
63 harvest, cut into small pieces, and dried in the shade for two weeks. The dried material was ground into a powder
64 using a manual homogenizer. *P. ostreatus* were obtained by cultivation at the biotechnological laboratory of the
65 University Cheikh Anta Diop of Dakar. The *Moringa oleifera* and *Pleurotus ostreatus* powders were combined in
66 a 2:1 ratio to create the dietary supplement. The mixture was created following ??ane et al, instructions (2017).
67 The combination was dissolved in 0.01% starch paste before being fed to the rats.

68 5 b) Animals and grouping

69 Wistar rats strain to weigh 150 to 200 g were obtained from the Animal House of the National Institute of Youth
70 and Sports in Yaounde. They were placed in plastic cages under standard laboratory conditions (temperature
71 20 to 30°C, relative air humidity 45 to 55%, and 12/12h light/dark cycle). The rats were fed with a basal diet
72 and water ad libitum. The feed was a standard rat chow composed of carbohydrates (52%), protein (22%), fat
73 (6.5%), water (12%), ash (6%), and fiber (4.5%). Every two days for 21 days, between 10:00 and 11:00 am, before
74 the mixture administration of *Moringa oleifera* and *Pleurotus ostreatus* in proportion 2:1, made as reported by
75 Kane et al., the amount of food and water ingested by each group of rats as well as body weights were recorded
76 (2017). The experiments were performed during the day (09am-03pm).

77 6 c) Experimental design

78 Thirty (30) adult male and female Wistar rats weighing 150 to 200 g were separated into five groups of six after
79 two weeks of acclimatization:

80 -Group 1 (TG): a stress-free control group that consumed only their regular diet of water, food, and vehicle
81 (starch paste) once daily for 21 days, -Group 2 (TP): a control group that received paracetamol 12 hours after
82 ethanol administration, was supplied in five sequential doses of 2 g. kg⁻¹ using an orogastric tube to stress the
83 group. For 21 days, they consumed the standard diet of water and food at their leisure in addition to the vehicle
84 starch paste, -Group 3 (D1P): a group that received 500 mg/kg of FMP16 and was stressed by ethanol in five
85 sequential doses of 2 g. kg⁻¹, administered through an orogastric tube; then received paracetamol 12 h after the
86 last dose of ethanol. They received the standard diet (water and food ab-libitum) and the vehicle starch paste
87 once a day for 21 days, -Group 4 (D2P): a group that received 1000 mg/kg of FMP16 and was stressed by ethanol
88 in five sequential doses of 2 g. kg⁻¹, administered through an orogastric tube; then received paracetamol 12
89 h after the last amount of ethanol. They received the normal diet (water and food ab-libitum) and the vehicle
90 starch paste once a day for 21 days, -Group 5 (D3P): a group that received 1500 mg/kg of FMP16 and was
91 stressed by ethanol in five sequential doses of 2 g. kg⁻¹, administered through an orogastric tube; then received
92 paracetamol 12 h after the last dose of ethanol. They received the standard diet (water and food ab-libitum) and
93 the vehicle starch paste once a day for 21 days.The rats were given full access to food and water, and were on
94 12-hour light cycle each day (dark 12h-12h light). They were force-fed FMP16 using a gastroesophageal catheter
95 and weighed every day. They fasted for the entire day before the animal sacrifice.

96 On the 23rd day, the rats were given a night of rest before being slaughtered (while sedated with ether) by
97 having their jugular veins cut. Organs such the liver, kidneys, brain, and testicles were collected along with
98 blood. The liver, which was exclusively used in this processing day. It was then rinsed with ice-cold saline (0.9%
99 NaCl) to eliminate any remaining blood.

7 d) Determination of the biochemicals parameters in liver

-Preparation of liver supernates Prior to biochemicals analysis, each liver sample was homogenized using a Potterproctor placed on ice and 10% homogenate was prepared using the KCL buffer solution (1.15%). The homogenates were centrifuged at 3000 rpm for 30 min at 4 ° C to collect the supernatant used for analysis. The supernatant of each sample was aliquoted in 1. 5

8 e) Statistical Analysis

IBM SPSS Statistics 20 software was used for statistical analysis and data processing. P-values less than 0.05 were regarded as significant in the statistical analysis, which was conducted using one-way analysis of variance (ANOVA) and Bonferroni's post-test for multiple comparisons. The results are presented as the mean and standard deviation (SD).

9 III.

10 Results and Discussion

Results have shown that no significant difference was observed in final body weights (155-173g) (Table1). Body weight gain ranged between 0.8 and 19 g for the four treatment groups. A decrease of 10% in the weight of D3N group was observed. These results corroborate those of ??lam et al., (2011 ??lam et al., (, 2009)) who found that a diet enriched with *Pleurotus ostreatus* decreases the body weight of animals. However, Bobek et al., ??1998) have shown that it does not affect the weight as well as Schneider et al., (2011) who worked on humans with a daily dose of 30 g of dried oyster mushrooms, found that this does not affect anthropometric data. Bénissan et al., 2012, showed that the daily intake of 30 g of *Moringa oleifera* leaves improves nutritional recovery in children suffering from malnutrition. Hanaa et al., ??2014), showed that a dose of 600 mg/kg of *Moringa oleifera* lowers the body mass index in obese subjects. Furthermore, the mixture of these species at a high dose of 1500 mg / kg, would explain the weight loss. This result was in contrast with those of Osman et al. (2012), who reported up to 14% changes in body weight of rats given *M.oleifera* extract for 21 days, attributing these changes to the rich nutrient quality of the extract.

Results also have shown no significant difference in the amount of protein in the liver (figure1). Regarding lipid peroxidation (figure ??), results show no significant difference in the concentration of peroxidized lipids between the groups except between the unstressed control group (TG) and the 1500 mg/kg dose group where the concentration was 34% higher. These results are not in agreement with those of Mladenovic et al, (2013); Patere et al, (2011); Johnsen et al, ??2007). The effect of FMP16 on oxidative stress enzymes such as catalase and cellular glutathione was also studied. Our results showed an increase in catalase activity of 87%, 85%, 90%, 82% respectively for the TG, D1P, D2P and D3P groups compared to the TP group (intoxicated and untreated). Also, catalase activity of the 1000 mg/kg dose (D2P) was 35% and 43% higher respectively compared to the 500 and 1500 mg/kg doses (figure ?? and 4). These results corroborate those of ??amou et). The markers that are used to determine toxicity are usually transaminases (ALAT and ASAT), whose high concentration in the extracellular medium is synonymous with an alteration of the cells. In this study, our results showed that ALAT and ASAT activities were decreased in the FMP16 groups (Table 2). Thus, ALAT activity decreased by 29% in the 1000 mg/kg dose group compared to the intoxicated control. ASAT activity was higher in the intoxicated group (TP) by 28% and 26% compared to the 1000 and 1500 mg/kg doses. Compared to the results of the studies by Adedapo et al, (2009) and ??lam et al, (2011), who instead found that a dose of 1600 mg/kg of *Moringa oleifera* leaves rather increased ALAT and ASAT activity; and on the other hand that a diet supplemented with 5% *Pleurotus* rather decreased transaminase activities, we could think that this explains the fact that FMP16 rather tends to regulate their activities due to the antagonistic effect that these two species have.

Furthermore, results showed that the administration of FMP16 did not cause any significant difference in albumin and testosterone levels (Table 2). Regarding creatinine, FMP16 administration decreased creatinine levels in the treated groups (D1P, D2P, D3P) compared to the untreated and stressed group (TP) (Table 2). These results corroborate those of Sirag, (2009), Adedapo et al,(2009), Kane et al, 2022 who showed the protective effect of *Pleurotus ostreatus* and *Moringa oleifera* on kidney damage.

Our results on lipid metabolism in rats revealed a significant decrease in total cholesterol in the 500, 1000 and 1500 mg/kg dose groups (figure ??). There was a 28%, 39%, 30% and 38% difference in TP, D1P, D2P and D3P compared to TG. In addition, a difference of 15% and 14% of D1P and D3P compared to TP. However, there was no significant difference in HDL cholesterol levels (Table 2). The results of the Triglycerides levels (Table 2) show a difference of 47% and 41% of the 500 and 1000 mg/kg dose compared to the TG control group. There was also a 28% decrease in Triglycerides levels at the 500 mg/kg dose compared to the dose 1000 mg/kg. In most of the studies on the effects of *Pleurotus ostreatus* and *Moringa oleifera*, they found a decrease in the concentration of LDL cholesterol which is more related to cardiovascular diseases (Bobek and Galbavy, 1999; ??obek et al., 1998; ??ossain et al., 2003). These results are also in agreement with those of Alam et al, (2009), Schneider et al, (2011), Chumark et al, (2008), Kane et al, (2022). Our results (figure ??) and those of previous studies suggest that FMP16 would be an excellent cholesterol-lowering agent that could be recommended for the prevention and treatment of cardiovascular diseases. IV.

11 Conclusion

159
160 A dietary supplement of *Moringa oleifera* leaves and *Pleurotus ostreatus* in wistar rats shows that the powders of
161 *M. oleifera* leaves and *P. ostreatus* mixture have an antihyperlipidemic effect as it significantly lowers total and
162 LDL cholesterol levels in rats stressed by combination of ethanol and paracetamol. The dose 1000 mg/kg is most
appropriate for chemically stressed animals. FMP16 would have no effect on albumin and testosterone levels. ¹

1

| GROUPES | Starting Body weight (g) | Final Body weight (g) | P-value* |
|---------|--------------------------|-----------------------|----------|
| TG | 154 ± 3,34 | 173,67 ± 9,16 a | 0,02 |
| TP | 154,33±3,44 | 177,33±4,84 | 0,01 |
| D1P | 154±2,53 | 185,67±2,86 | 0,01 |
| D2P | 153,20±2,66 | 168±4,14 | 0,6 |
| D3P | 153±3,03 | 157±7,14 | 0,13 |

*ANOVA test; TG: control group; TP: stressed group ethanol+paracetamol; D1P: stressed and treated group 500 mg/kg; D2P: stressed and treated group 1000 mg/kg; D3P: stressed and treated group 1500 mg/kg; a, d, e: mean statistical significance to p < 0,05 (test de Bonferoni)

Figure 1: Table 1 :

2

| GROUPES | ALAT (U/I) | ASAT (U/I) | triglycerides activity | | | Triglyc. (mg/dl) |
|---------|------------|--------------|------------------------|-----------------|---------------|------------------|
| | | | ALBUMINE (g/dl) | TESTOS. (ng/dl) | HDL-C (mg/dl) | |
| TG | 32,74±7,09 | 160,92±30,02 | 1,53±0,20 | 0,31±0,06 | 39,07±2,18 | 52,03±0,68 |
| TP | 48,48±3,32 | 196,35±33,40 | 1,61±0,37 | 0,47±0,12 | 38,88±1,88 | 78,14±12,69 |
| D1P | 43,16±8,80 | 209,16±25,74 | 1,80±0,10 | 0,53±0,08 | 36,30±2,89 | 71,02±18,92 |
| D2P | 34,31±4,55 | 141,86±11,19 | 1,60±0,20 | 0,44±0,04 | 35,08±2,53 | 98,18±7,64 |
| D3P | 43,66±2,13 | 145,86±22,20 | 1,75±0,18 | 0,43±0,03 | 34,17±5,47 | 87,83±21,97 |

Figure 2: Table 2 :

163

¹The values are expressed as mean ± SD. TG: Control group rats with food and water ad libitum, TP: stressed rats without treatment, D1P: dose of 500 mg/kg, D2P: dose of 1000 mg/kg, D3P : dose of 1500 mg/kg.

Year 2022
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Volume XXII Issue II Version I
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Medical Research
Global Journal of
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[Note: The values are expressed as mean \pm SD. TG: Control group rats with food and water ad libitum, TP: stressed rats without treatment, D1P: dose of 500 mg/kg, D2P: dose of 1000 mg/kg, D3P: dose of 1500 mg/kg. a, b ,c, d statistically different mean compared to TP, D1P, D2P and D3P at $p < 0.05$ (LSD test) e, f mean statistically different from D1P and D3P at $p < 0.05$ (LSD test)]

Figure 3:

164 .1 Acknowledgements

165 The authors would like to thank the Biotechnology Laboratory of the University Cheikh Anta Diop of Dakar,
166 Senegal, and the Faculty of Science, the FODRUS-LAPHER-Biotech of the University of Yaoundé I, Cameroon.
167 The project was part of a scholarship provided to Fatou Corcka KANE through Africa for Innovations, Mobility,
168 Exchange, Globalization and Quality (AFIMEGQ) program sponsored by the European Commission's EACEA
169 program.

170 .2 Conflict of Interests

171 The authors declare that there is no conflict of interests regarding the publication of this paper.

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11 CONCLUSION

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