

Chronic Inflammation Induced Immunosuppression in Tumor Microenvironment of Oral Cancer

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Abstract

Oral Cancer is a wound that do not heal is a complex disease consists of heterogeneous tissue in their tumor microenvironment. Oral cancer accounts eighth most Common Cancer worldwide. Chronic inflammatory mediators released from immune cells in tumor microenvironment of oral cancer such as macrophages, T lymphocytes, dendritic cells ,Natural killer cells release cytokines, Chemokine?s and growth factors helps in generation of myeloid derived suppressor cells. Myeloid derived suppressor cells are derived from myeloid progenitor cells of bone marrow secretes inflammatory mediators iNOS, arginase-1, PGE2,IL-10 and IL-4 suppresses adaptive and innate immunity by interacting with macrophages ,T-cells, Natural killer cells and dendritic cells favours pro-tumoral activity by activating transcriptional factors (NF-KB,STAT -3,HIF) further progress in to oral cancer. Myeloid derived suppressor cells reduces T cell activation and function by Arginase-1, iNOS, peroxy nitrate over expression and cysteine depletion. This article describes mainly about immune cells in tumor microenvironment especially macrophages, T lymphocytes, dendritic cells, Natural killer cells their interactions with myeloid derived suppressor cells.

Index terms— myeloid derived suppressor cells, chronic inflammation, oral cancer, granulocytic monocytic colony stimulating factor, natural killer cells, transform

1 I. Introduction

Inflammation is the body response to any type of injury, in which various mediators are released in surrounding environment. Recent debated topic is inflammation associated onco -promotion in tumor microenvironment. Inflammatory mediators in oraltumor micro-environment consists of mediators of inflamamation are Neutrophils, lymphocytes, macrophages, Natural killer cells, Dendritic cells secreting cytokines. Which can induce Immunomodulation by myeloid derived suppressor cells (MDSC) results in Oral tumor promotion, progression, and metastasis (1). editing, immuneprocessing and immuneevasion. Immunoevasion is one of the hallmark of tumor in order to progress. Immunoevasion mechanism involves the production of cytokines, which are immunosuppressive, T cell apoptosis or loss of HLA class1 and costimulatory molecules. In Immunoediting high immunogenicity tumorseliminate tumor by NK cells, macrophages,T cells. Reduced tumor cell variant immunogenicity favour tumor progression by immunosuppression or resistant to immune attack. Immuno processing stage genetic instability and heterogeneity of cancer cells favour promotion of tumor which, are poorly recognized by immune system or immunosuppression.

Immunescape stage altered by expression of MHC1 and 11 and costimulatory molecules, antigen processing dysregulation antigen processing, expression of low levels tumor antigen, other mechanisms of immunosuppression are T cell tolerance to tumor antigen and immunosuppressive cytokines IL-10,TGF-Beta or T regulatory cells (Treg). (48) Oral cancer is an eighth most common cancer in the worldwide. Every year nearly 300,400 new cases have been reporting worldwide and costs 145,400 lives a year. Squamous cell carcinoma involves 90% of

4 III. ROLE OF MDSC IN IMMUNOSUPPRESSION

43 head and neck region especially from mucosal epithelium linked to various adverse habits such as smoking form
44 of tobacco, smokeless tobacco, alcohol drinking and also human papilloma virus.

45 Advance oral cancer locally, management has been a challenging issue involving multidisciplinary approach
46 of surgery, chemotherapy and radiotherapy. Despite recent improvement in management of oral cancer still the
47 prognosis is grave with five year survival rate nearly 50%.

48 Early stage of inflammation neutrophils are predominant leucocyte and first cell to migrate are regulated
49 by macrophages and mast cells in tissue. As inflammation proceeds various types of leucocytes majority of
50 them are lymphocytes gets activated and recruited to the inflammatory site by a signalling network involving
51 chemokines, cytokines, growth factors for defense against infection. Shifting of antimicrobial tissue damage
52 to tissue repair occurs mediated by PGE₂, TGF-Beta and reactive oxygen and nitrogen intermediates having
53 dual role in both aggravating and suppressing inflammation. Resolution of inflammation requires macrophages,
54 dendritic cells and phagocytes by apoptosis and phagocytosis, which promote an anti-Immune cells has an
55 important role in preventing or promoting cancer through immune surveillance of tumor by mechanism of
56 immune-inflammatory response. If inflammation is dysregulated, aggravating to chronic inflammatory cellular
57 response causing immunosuppression, tissue and DNA damage by cytokines, growth factors, reactive oxygen and
58 nitrogen species released from macrophages and lymphocytes (1,2).

59 2 II.

60 3 Factors Affecting Inflammation Induced Immunosuppression 61 in Tumor Microenvironment of Oral Cancer

62 Oral tumor microenvironment consists of various heterogeneous inflammatory mediators such as neutrophils,
63 natural killer cells, T and B lymphocytes, mast cells, and antigen presenting cells(APC) such as macrophages,
64 Dendritic cells and other distinct cell types including fibroblasts, Carcinoma associated fibroblast, smooth
65 muscle cells, myo-fibroblast, endothelial cells and their precursors, pericytes. Recent data have demonstrated a
66 role of these individual components, in particular carcinoma associated fibroblasts, macrophages and endothelial
67 cells, in promoting tumor growth and progression (1-2). The tumor stroma has an indispensable role in acquiring
68 hallmark capabilities. The stroma provides support with growth factors (GM-CSF, G-CSF, M-CSF; VEGF; TGF),
69 cytokines (IL-1, IL-4, IL-5, IL-6, IL-10, IL-13, TNF-Alpha, Interferon -Gamma), chemokines (CCL2, CCL4, CCL5,
70 CXCL1, CXCL12 and CXCL8) along with COX2 which, secrete prostaglandin E₂, promotes the generation of
71 Myeloid derived suppressor cells.

72 4 III. Role of mdsc in Immunosuppression

73 These are immature heterogeneous myeloid cells that fail to terminally differentiate in to granulocytes, dendritic
74 cells or macrophages on chronic inflammatory conditions and exhibit immunosuppressive function by multiple
75 mechanism. Their broadly distinct phenotypical characteristics, Among human MDSCs, the two subsets can be
76 distinguished as Granulocytic and Monocytic(3). which, is responsible for immuno-modulatory activity in tumor
77 microenvironment by evading active immune system by various factors by potent inhibitors of both antigenic
78 -specific and non-specific T-cell activation. These factors are arginase, nitric oxide, Reactive oxygen species
79 (ROS), PGE₂, Cystein, peroxy nitrate. An important mutagenic factor frequently abundant in an inflammatory
80 microenvironment is ROS (eg. Oxygen ions and peroxides) results from oxidative stress induced by phagocytic
81 cells. ROS are highly reactive, unstable molecules that damage DNA increases the cell mutation rate, thus
82 favouring the appearance of clones with oncogenic properties. Potential key mechanism of MDSC -induced CD8+
83 T-cell immunosuppression in tumor bearing hosts by increased NADPH oxidase, NOX 2 activity (4-5). Nitric
84 oxide is produced by MDSC by utilising L-arginine as substrate for nitric oxide synthase (6,7). Which, suppresses
85 T-cell activation, adhesion, proliferation and migration (8)(9)(10)(11)(12)(13). It also suppresses T cell function,
86 particularly CD8+ T cells by blocking the activation of signalling molecules in T cells, including JAK1 (Janus
87 activated kinase 1), STAT5, ERK and Akt (8,11). It has also been shown to inhibit MHC class 2 expression
88 and promote CD8 T-cell apoptosis (14,15). Other important moderator synthesized by MDSC is Arginase.
89 L-Arginine is a conditionally essential amino acid and metabolized by arginases and nitric oxide synthases to
90 produce either L-ornithine and urea (16,17,18). L-arginine is an amino acid required for T cell function and
91 proliferation. L-arginine deprivation has been reported to induce T-cell dysfunction and suppression of T-cell
92 function (19,20,21). These mechanisms seem to contribute to the protumoral function of MDSC (22). MDSC
93 are copious producers of peroxy nitrate and increased levels are associated with tumor progression by inhibiting
94 antigen specific, cytotoxic T-cell responses (23). Cysteine is an essential amino acid required for T-cell activation,
95 differentiation and proliferation (24). MDSC mediated cysteine depletion, block activation of T-cell from the
96 local microenvironment results in the inhibition of T-cell activation and function (25). PGE₂ is an eicosanoid
97 synthesized by COX2 produced and secreted by MDSC, mediated over expression of arginase, Correlated with their
98 pro-inflammatory and immunosuppressive properties, further inhibiting the activity of CD8+ T cells. MDSCs
99 immunosuppressive function, activation and proliferation is activated by IFN-gamma, TLR ligands, IL-13, IL-
100 4, and TGF -beta, which trigger STAT3 and NF-kb signalling pathways (26,27,28). These various factors are
101 produced during the course of inflammation following cellular stresses, in response to hormones, growth factors,

102 endotoxin and inflammatory cytokines or by growing tumors which induces angiogenesis, apoptosis, chronic
103 inflammation and immunosuppression (28,29).

104 5 IV. Interaction of mdsc with other Immuncells

105 MDSCs communication network between macrophages and DCs that promotes and maintains an immunosup-
106 pressive microenvironment. This communication is mainly mediated by inflammatory mediators IL-1beta, IL-
107 6, IL-10, PGE-2, and TGF -beta (30,31). The activating NK receptors inhibited by IDO (Indoleamine 23-
108 dioxygenase) andPGE2 are counteracted by NKG2A an inhibitory receptor utilized by both T and NK cells
109 (32). An early response of damaged tissue is production of IL-8 by the epithelial cell itself, which together with
110 macrophages and mast cells secrete TNF-alfa and histamine allows neutrophil extravasion to injure site initiating
111 inflammation. Chemokines secreted by endothelial cells and macrophages brings inflammatory and immune cells
112 to the site of inflammation (33). Among inflammatory factors promoting proliferation are TGFbeta, fibroblast
113 growth factor, epithelial growth factor, TGF-beta synthesized by mast cells, macrophages and lymphocytes as an
114 inactive precursor in inflammatory microenvironment activated by proteases. TGF-beta promotes mesenchymal
115 Cell proliferation and immunomodulation by promoting N2 neutrophils and M2 macrophages, facilitates tumor
116 invasion and metastasis (34,35). LPS is a known activator of macrophage cross talk with MDSC in the presence of
117 LPS. Later LPS binds to LPS binding protein. Which helps in transfer ofLPS to the membrane bound receptor
118 CD14 through TLR4signalling pathway. TLR4 signalling pathway gets activated by CD14 binds with TLR4
119 further downstream activation of NF-kb driving MDSC production of IL-10 resulting in immunosuppression and
120 immune evasion by promoting M2 polarization of macrophages (36). Alternatively activated macrophages(M2
121 type) are an important source of both Fibroblast growth factors, and Endothelial growth factors activated
122 by cytokines such as IL-4, IL-5, IL-6, IL-9, IL-13, IL-17 and TGF-beta acts as a immunosuppressor towards
123 Treg (Regulatory T cell) cells maintain immunosuppressive microenvironment (37,38). Tumor stromacan also
124 suppress immune effector function. Extra cellular accumulation of lactate, adenosine, VEGF under hypoxic
125 condition activated by hypoxia inducible transcriptional factor (HIF) further induces angiogenesis. Cross talk
126 between MDSC and dendritic cells in presence of cytokines such as IL-4, GM-CSF and PGE2 results in decrease
127 in production of mature dendritic cells , blocking T-cell production of IFN-gammaand dendritic cells production
128 of Proinflammatory cytokine IL-23driving the proliferation and inflammatory function of Th17 cells. Which
129 suppresses both adaptive and innate immunity, due to immunosuppressive network, the immature dendritic cell
130 fail to activate to become mature dendritic cell on antigenic presentation. So, the activation ofCD4+ and CD8+ T
131 cells don't take place. All together co-operate to inhibit Dendritic cell antigen-processing, presenting activity and
132 dendritic cell tolerance (39)(40)(41)(42)(43)(44)(45)(46)(47).All these factors contribute to pro-tumoral activity,
133 tumor progression, invasion and metastasis. Inflammation is considered to be a' Seventh hallmark' of cancer (4).

134 Myeloid derived suppressor cells are immature myeloid cells of myeloid progenitor cells upon chronic
135 inflammation. They are of two types Monocystic-MDSC and Granulocytic-MDSC. Myeloid derived suppressor
136 cells induce immunosuppression by various mechanisms suppresses both innate and adaptive immunity, it
137 also possess plasticity and the type of MDSC in tumor microenvironment determines the immunosuppression.
138 Complex interactions between MDSC and immune cells and their role in immunosuppression need to be studied.
139 Understanding of MDSC biology, chronic inflammatory mediators, which helps in MDSC recruitment, generation,
140 activation and their role in immunosuppression must be revealed for therapeutic strategy and its role in tumor
141 prognosis.

142 6 V. Conclusion

143 Thorough understanding of immune cells of Oral tumor microenvironment , role of immune cells such as
144 Macrophages, T lymphocytes and natural killer cells which, drive towards tumorigenesis. Role of Inflammatory
145 cells and their mediators such as cytokines, their interactions with myeloid derived suppressor cells are major
146 immunosuppressor and immune evasion cells. Phonotypical and functional role of myeloid derived suppressor
147 cells in oral tumor microenvironment linking between inflammation and oral cancer. Hence, modulating targeted
148 or combined immune cells in oral tumor microenvironment, could possibly hold a future therapeutic opportunity
149 with better survival rate and less possible complications.

150 7 Abbreviations

151 1



Figure 1:

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- 152 [Gout ()] 'A potent suppressor of lymphoma growth by inhibition of the x(c)-cystine transporter: anew action
153 for an old drug'. P W Gout . *oncogene* 2011. 15 p. .
- 154 [Nagaraj et al.] *Altered recognition of antigen is a mechanism of*, S Nagaraj , K Gupta , V Pisarev , L Kinarsky
155 , S Sherman , L Kang , D L Herber , J Schneck , D I Gabrilovich .
- 156 [Nagaraj ()] 'Altered recognition of antigen is a mechanism of CD8+ T cell tolerance in cancer'. S Nagaraj . *Nat*
157 *Med* 2007. 13 p. .
- 158 [Rodriguez ()] 'Arginase -1 production in the tumor microenvironment by mature myeloid cells inhibits T-cell
159 receptor expression and antigenspecific T-cell responses'. P C Rodriguez . *Cancer Res* 2004. 64 p. .
- 160 [Wu ()] 'Arginine metabolism: nitric oxide and beyond'. G Wu . *Biochem J* 1998. 336 p. .
- 161 [Rodriguez and Ochoa ()] 'Arginine regulation by myeloid derived suppressor cells and tolerance in cancer:
162 mechanisms and therapeutic perspectives'. P C Rodriguez , A C Ochoa . *Immunol Rev* 2008. 222 p. .
- 163 [Rodriguez ()] 'Arginine regulation by myeloid derived suppressor cells and tolerance in cancer: mechanisms and
164 therapeutic perspectives'. P C Rodriguez . *Immunol Rev* 2008. 222 p. .
- 165 [Medot-Pirenne et al. ()] 'Augmentation of an antitumor CTL response In vivo by inhibition of suppressor
166 macrophage nitric oxide'. M Medot-Pirenne , M J Heilman , M Saxena , P E Mcdermott , C D Mills .
167 *J Immunol* 1999. 163 p. .
- 168 [Medot-Pirenne ()] 'Augmentation of an antitumor CTL response in vivo by inhibition of suppressor macrophage
169 nitric oxide'. M Medot-Pirenne . *J Immunol* 1999. 163 p. .
- 170 [Colotta et al. ()] 'Cancer -related inflammation, the seventh hallmark of cancer; links to genetic instability'. F
171 Colotta , P Allavena , A Sica , C Garlanda , MantovaniA . *Carcinogenesis* 2009. 30 p. .
- 172 [CD8+ T cell tolerance in cancer Nat Med ()] 'CD8+ T cell tolerance in cancer'. *Nat Med* 2007. 13 p. .
- 173 [Ostrand -Rosenberg et al. ()] 'Cross-talk between myeloid -derived suppressor cells (MDSC), macrophages, and
174 dendritic cells enhances tumor -induced immune suppression'. S Ostrand -Rosenberg , P Sinha , Beury Dw ,
175 V K Clements . *Seminars in cancer biology* 2012. 22 (4) p. .
- 176 [Ostrand -Rosenberg ()] 'Cross-talk between myeloid -derived suppressor cells (MDSC), macrophages, and
177 dendritic cells enhances tumor -induced immune suppression'. S Ostrand -Rosenberg . *Seminars in cancer*
178 *biology* 2012. 22 p. .
- 179 [Parker et al. ()] 'Derived Suppressor Cells: Critical Cells Driving Immune Suppression in the Tumor Microen-
180 vironment'. K H Parker , D W Beury , S Ostrand-Rosenberg , Myeloid . *Advances in Cancer Research* 2015.
181 12 p. .
- 182 [Edgington-Mitchell and Parker ()] 'Disparate functions of myeloid-derived suppressor cells in cancer metastasis'.
183 Laura E Edgington-Mitchell , Belinda S Parker . *Cancer Forum* 2014. 38 p. .
- 184 [Mancino et al. ()] 'Divergent effects of hypoxia on dendritic cell functions'. A Mancino , T Schioppa , P Larghi
185 . *Blood* 2008. 112 p. .
- 186 [Mancino ()] 'Divergent effects of hypoxia on dendritic cell functions'. A Mancino . *Blood* 2008. 112 p. .
- 187 [Sosroseno et al. ()] 'Effect of exogenous nitric oxide on murine splenic immune response induced by Aggregati-
188 bacter actinomycetemcomitans lipopolysaccharide'. W Sosroseno , P S Bird , G J Seymour . *Anaerobe* 2009.
189 15 p. .
- 190 [Sosroseno ()] 'Effect of exogenous nitric oxide on murine splenic immune response induced by Aggregatibacter-
191 actinomycetemcomitans lipopolysaccharide'. W Sosroseno . *Anaerobe* 2009. 15 p. .
- 192 [Harda et al. ()] 'Essential involvement of interleukin -8 (IL-8) in acute inflammation'. A Harda , N Sekido , T
193 Akahoshi , T Wada , N Mukaida , K Matsushima . *Journal of leukocyte biology* 1994. 56 p. .
- 194 [Harda ()] 'Essential involvement of interleukin -8 (IL-8) in acute inflammation'. A Harda . *Journal of leukocyte*
195 *biology* 1994. 56 p. .
- 196 [Expect the unexpected The journal of Clinical investigation ()] 'Expect the unexpected'. *The journal of Clinical*
197 *investigation* 2015. 7 p. .
- 198 [Pietras K, Ostman ()] 'Hallmarks of cancer; Interactions with the tumorstroma'. A Pietras K, Ostman . *Exp*
199 *cell Res* 2010. 316 p. .
- 200 [Yang et al. ()] 'HIF-dependent induction of adenosine receptor A2b skews human dendritic cells to a Th2 -
201 stimulating phenotype under hypoxia'. M Yang , C Ma , S Liu . *Immunology and cell biology* 2010. 88 p.
202 .
- 203 [Yang ()] 'HIF-dependent induction of adenosine receptor A2b skews human dendritic cells to a Th2 -stimulating
204 phenotype under hypoxia'. M Yang . *Immunology and cell biology* 2010. 88 p. .
- 205 [Elia et al. ()] 'Human dendritic cells differentiated in hypoxia down modulate antigen uptake and change their
206 chemokine expression profile'. A R Elia , P Cappello , M Puppo . *Journal of leukocyte biology* 2010. 84 p. .

7 ABBREVIATIONS

- 207 [Elia ()] ‘Human dendritic cells differentiated in hypoxia down modulate antigen uptake and change their
208 chemokine expression profile’. A R Elia . *Journal of leukocyte biology* 2010. 84 p. .
- 209 [Lo Manaco et al. ()] ‘Human Leukocyte antigen E contributes to protect tumor cells from lysis by natural killer
210 cells’. E Lo Manaco , E Tremante , C Cerboni , E Melluci , L Sibilio , A Zingoni . *Neoplasia* 2011. 13 p. .
- 211 [Lo Manaco ()] ‘Human Leukocyte antigen E contributes to protect tumor cells from lysis by natural killer cells’.
212 E Lo Manaco . *Neoplasia* 2011. 13 p. .
- 213 [Langrish ()] ‘IL-23 drives a pathogenic T cell population that induces autoimmune inflammation’. C L Langrish
214 . *J Exp Med* 2005. 201 p. .
- 215 [Langowski ()] ‘IL-23 promotes tumour incidence and growth’. J L Langowski . *Nature* 2006. 442 p. .
- 216 [Teng et al. ()] ‘IL-23 suppresses innate immune response independently of IL-17A during carcinogenesis and
217 metastasis’. M W Teng , D M Andrews , N Mclaughlin , Von Scheidt , B Ngiow , S F Moller , A . *Proc Natl*
218 *Acad Sci* 2010. 107 p. .
- 219 [Teng ()] ‘IL-23 suppresses innate immune response independently of IL-17A during carcinogenesis and metas-
220 tasis’. M W Teng . *Proc Natl Acad Sci* 2010. 107 p. .
- 221 [Bronte et al. ()] ‘IL-4-induced arginase 1 suppresses alloreactive T cells in tumor-bearing mice’. V Bronte , P
222 Serafini , De Santo , C Marigo , I Tosello , V Mazzoni , A . *J Immunol* 2003. 170 p. .
- 223 [Bronte ()] ‘IL-4-induced arginase 1 suppresses alloreactive T cells in tumor-bearing mice’. V Bronte . *J Immunol*
224 2003. 170 p. .
- 225 [Rivoltini et al. ()] ‘Immunity to cancer: attack and escape in T lymphocyte-tumor cell interaction’. L Rivoltini
226 , M Carrabba , V Huber , C Castelli , L Novellino , P Dalerba . *Immunol Rev* 2002. 188 p. .
- 227 [Rivoltini ()] ‘Immunity to cancer: attack and escape in T lymphocyte-tumor cell interaction’. L Rivoltini .
228 *Immunol Rev* 2013. 188 p. .
- 229 [Harari and Liao ()] ‘Inhibition of MHC II gene transcription by nitric oxide and antioxidants’. O Harari , J K
230 Liao . *Curr Pharm Des* 2004. 10 p. .
- 231 [Harari ()] ‘Inhibition of MHC II gene transcription by nitric oxide and antioxidants’. O Harari . *Curr Pharm*
232 *Des* 2012. 10 p. .
- 233 [Tindall et al. ()] ‘Interleukin -6 promoter variants, prostate cancer risk and survival’. E A Tindall , G Severi ,
234 N Hoang . *The prostate* 2012. 72 p. .
- 235 [Tindall ()] *Interleukin -6 promoter variants, prostate cancer risk and survival. The prostate*, E A Tindall . 2012.
236 72 p. .
- 237 [Wang et al. ()] ‘Interleukin -6(IL-6) signaling regulates anchorage independent growth, proliferation, adhesion
238 and invasion in human ovarian cancer cells’. Y Wang , L Li , X Guo . *Cytokine* 2012. 59 p. .
- 239 [Wang ()] ‘Interleukin -6(IL-6) signaling regulates anchorage independent growth, proliferation, adhesion and
240 invasion in human ovarian cancer cells’. Y Wang . *Cytokine* 2012. 59 p. .
- 241 [Jiang et al. ()] ‘Lipopolysaccharide induces physical proximity between CD14 and toll-like receptor 4 (TLR4)
242 prior to nuclear translocation of NF-kappa B’. Q Jiang , S Akashi , K Miyake , H R Petty . *J Immunol* 2000.
243 165 p. .
- 244 [Jiang ()] ‘Lipopolysaccharide induces physical proximity between CD14 and toll-like receptor 4 (TLR4) prior
245 to nuclear translocation of NF-kappa B’. Q Jiang . *J Immunol* 2000. 165 p. .
- 246 [Bingisser et al. ()] ‘Macrophage-derived nitric oxide regulates T cell activation via reversible disruption of the
247 Jak3/ STAT5 signaling pathway’. R M Bingisser , P A Tilbrook , P G Holt , Kees U R . *J Immunol* 1998.
248 160 p. .
- 249 [Bingisser ()] ‘Macrophagederivednitric oxide regulates T cell activation via reversible disruption of the
250 Jak3/STAT5signaling pathway’. R M Bingisser . *J Immunol* 1998. 160 p. .
- 251 [Gabrilovich ()] ‘Mechanisms and functional significance of tumour-induced dendriticcell defects’. D Gabrilovich
252 . *Nat Rev Immunol* 2014. 4 p. .
- 253 [Xu et al. ()] ‘Mesenchymal stem cells play a potential role in regulating the establishment and maintenance of
254 epithelial -mesenchymal transition in MCF7 human breast cancer cells by paracrine and induced autocrine
255 TGF-beta’. Q Xu , L Wang , H Li . *International journal of oncology* 2012. 41 p. .
- 256 [Xu ()] ‘Mesenchymal stem cells play a potential role in regulating the establishment and maintenance of
257 epithelial -mesenchymal transition in MCF7 human breast cancer cells by paracrine and induced autocrine
258 TGF-beta’. Q Xu . *International journal of oncology* 2012. 41 p. .
- 259 [Dilek et al. ()] ‘Myeloid -derived suppressor cells: mechanisms of action and recent advances in their role in
260 transplant tolerance’. N Dilek , R Vuillefroy D Silly , G Blancho , B Vanhove . *Frontiers in immunology* 2012.
261 3 p. .

-
- 262 [Dilek ()] ‘Myeloid -derived suppressor cells: mechanisms of action and recent advances in their role in transplant
263 tolerance’. N Dilek . *Frontiers in immunology* 2014. 3 p. .
- 264 [Douglas and Gabrilovich] *Myeloid derived suppressor cells in the tumor microenvironment*, M Douglas , Dmitry
265 I Gabrilovich .
- 266 [Keskinov and Shurin ()] ‘Myeloid regulatory cells in tumor spreading and metastasis’. A A Keskinov , M R
267 Shurin . *Immunobiology* 2015. 220 (2) p. .
- 268 [Keskinov and Shurin (2015)] ‘Myeloid regulatory cells in tumor spreading and metastasis’. A A Keskinov , M R
269 Shurin . *Immunobiology* 2015 Feb 28. 220 (2) p. .
- 270 [Mazzoni ()] ‘Myeloid suppressor lines inhibit T cell responses by an NO-dependent mechanism’. A Mazzoni . *J*
271 *Immunol* 2002. 168 p. .
- 272 [Mazzoni et al. ()] ‘Myeloid suppressor lines inhibit T cell responses by an NOdependent mechanism’. A Mazzoni
273 , V Bronte , A Visintin , J H Spitzer , E Apolloni , P Serafini , P Zanovello , D M Segal . *J Immunol* 2002.
274 168 p. .
- 275 [Medina-Echeverz et al. ()] ‘Myeloid-derived cells are key targets of tumor ‘immunotherapy’. J Medina-Echeverz
276 , F Aranda , P Berraondo . *Oncoimmunology* 2014. 3 (4) p. .
- 277 [Peranzoni et al. ()] ‘Myeloid-derived suppressor cell heterogeneity and subset definition’. E Peranzoni , S Zilio ,
278 I Marigo , L Dolcetti , P Zanovello , S Mandruzzato , V Bronte . *Curr OpinImmunol* 2010. 22 p. .
- 279 [Raffaghello and Bianchi ()] ‘Myeloid-Derived Suppressor Cells and Tumor Growth. In Interaction of Immune
280 and Cancer Cells’. L Raffaghello , G Bianchi . *Journal of cancer* 2014. p. .
- 281 [Poschke ()] ‘Myeloid-derived suppressor cells impair the quality of dendritic cell vaccines’. I Poschke . *Cancer*
282 *Immunol Immunother* 2011. 8 p. .
- 283 [Poschke et al. ()] ‘Myeloid-derived suppressor cells impair the quality of dendritic cell vaccines’. I Poschke , Y
284 Mao , L Adamson , F Salazar-Onfray , G Masucci , R Kiessling . *Cancer Immunol Immunother* 2011. 8 p. .
- 285 [Srivastava et al. ()] ‘Myeloid-derived suppressor cells inhibit T-cell activation by depleting cystine and cysteine’.
286 M K Srivastava , P Sinha , V K Clements , P Rodriguez , S Ostrand-Rosenberg . *Cancer Res* 2010. 70 p. .
- 287 [Srivastava ()] ‘Myeloid-derived suppressor cells inhibit T-cell activation by depletingcystine and cysteine’. M K
288 Srivastava . *Cancer Res* 2010. 70 p. .
- 289 [Diaz-Montero et al. ()] ‘Myeloidderived suppressor cells in cancer: therapeutic, predictive, and prognostic
290 implications’. C M Diaz-Montero , J Finke , A J Montero . *Seminars in oncology* 2014. 41 p. .
- 291 [Ostrand -Rosenberg and Sinha ()] ‘Myeloidderived suppressor cells: linking inflammation and cancer’. S Os-
292 trand -Rosenberg , P Sinha . *Journal of immunology* 2009. 188 (8) p. .
- 293 [Ostrand -Rosenberg ()] ‘Myeloidderived suppressor cells: linking inflammation and cancer’. S Ostrand -
294 Rosenberg . *Journal of immunology* 2009. 188 p. .
- 295 [Mittal ()] ‘New insights into cancer immunoediting and its three component phaseselimination, equilibrium and
296 escape’. D Mittal . *Curr opin immunol* 2014. 27 p. .
- 297 [Bogdan ()] ‘Nitric oxide and the immune response’. C Bogdan . *Nat Immunol* 2001. 2 p. .
- 298 [Bogdan ()] ‘Nitric oxide and the immune response’. C Bogdan . *Nat Immunol* 2011. 2 p. .
- 299 [Bauer et al. ()] ‘Nitric oxide inhibits the secretion of T-helper 1-and T-helper 2-associated cytokines in activated
300 human T cells’. H Bauer , T Jung , D Tsikas , D O Stichtenoth , J C Frolich , C Neumann . *Immunology*
301 1997. 90 p. .
- 302 [Lejeune et al. ()] ‘Nitric oxide involvement in tumorinduced immunosuppression’. P Lejeune , P Lagadec , N
303 Onier , D Pinard , H Ohshima , J F Jeannin . *J Immunol* 1994. 152 p. .
- 304 [Bobe ()] ‘Nitric oxide mediation of active immunosuppression associated with graft-versus hostreaction’. P Bobe
305 . *Blood* 1999. 94 p. .
- 306 [Sato et al. ()] ‘Nitric oxide plays a critical role in suppression of T-cell proliferation by mesenchymal stem cells’.
307 K Sato , K Ozaki , I Oh , A Meguro , K Hatanaka , T Nagai , K Muroi , K Ozawa . *Blood* 2007. 109 p. .
- 308 [Sato ()] ‘Nitric oxide plays a critical role in suppression of T-cell proliferation by mesenchymal stem cells’. K
309 Sato . *Blood* 2007. 109 p. .
- 310 [Lejeune ()] ‘Nitricoxide involvement in tumor-induced immunosuppression’. P Lejeune . *J Immunol* 1994. 152
311 p. .
- 312 [Fiaschi and Chiarugi ()] ‘Oxidative stress, tumor microenvironment, and metabolic reprogramming: a diabolic
313 liaison’. T Fiaschi , P Chiarugi . *International Journal of Cell Biology* 2012. 20 p. .
- 314 [Jiang et al. ()] ‘Phenotypes, accumulation, and functions of myeloid-derived suppressor cells and associated
315 treatment strategies in cancer patients’. J Jiang , W Guo , X Liang . *Human immunology* 2014. 75 (11) p. .

7 ABBREVIATIONS

- 316 [Eruslanov ()] ‘Pivotal advance ; Tumormediated induction of myeloid -derived suppressor cells and M2-polarized
317 macrophages by altering intracellular PGE2 catabolism in myeloid cells’. E Eruslanov . *J. Leukol. Biol* 2010.
318 88 p. .
- 319 [Eruslanov et al. ()] ‘Pivotal advance; Tumor -mediated induction of myeloid -derived suppressor cells and M2-
320 polarized macrophages by altering intracellular PGE2 catabolism in myeloid cells’. E Eruslanov , I Daurkin ,
321 J Ortiz , J Vieweg , S Kusmartsev . *J. Leukol. Biol* 2010. 88 p. .
- 322 [Legler et al. ()] ‘Prostaglandin E2 at new glance; Novel insights in functional diversity offer therapeutic chances’.
323 D F Legler , M Bruckner , E Vetz-Von Allmen , P Krause . *International. Journal of Biochem cell Biol* 2010.
324 42 p. .
- 325 [Legler ()] ‘Prostaglandin E2 at new glance; Novel insights in functional diversity offer therapeutic chances’. D
326 F Legler . *International. Journal of Biochem cell Biol* 2010. 42 p. .
- 327 [Morris and Jr ()] ‘Regulation of enzymes of the urea cycle and arginine metabolism’. S M Morris , Jr . *Annu*
328 *Rev Nutr* 2002. 22 p. .
- 329 [Morris ()] ‘Regulation of enzymes of the urea cycle and arginine metabolism’. S M Morris . *Annu Rev Nutr* 2002.
330 22 p. .
- 331 [Bronte ()] ‘Regulation of immune responses by L-arginine metabolism’. V Bronte . *Nat Rev Immunol* 2005. 5 p.
332 .
- 333 [Condamine et al. ()] *Regulation of tumor metastasis by myeloid-derived suppressor cells. Annual review of*
334 *medicine*, T Condamine , I Ramachandran , J I Youn , D I Gabrilovich . 2015. 66 p. .
- 335 [Zhao et al. ()] ‘Subsets of myeloid-derived suppressor cells in hepatocellular carcinoma express chemokines and
336 chemokine receptors differentially’. W Zhao , Y Xu , J Xu , D Wu , B Zhao , Z Yin , X Wang . *International*
337 *immunopharmacology* 2015. 26 (2) p. .
- 338 [Wang ()] ‘Transforming growth factor beta -induced epithelial -mesenchymal transition increases cancer stem
339 -like cells in the PANC-1 cell line’. H Wang . *Oncology letters* 2012. 3 p. .
- 340 [Wang et al. ()] ‘Transforming growth factor beta -induced epithelial -mesenchymal transition increases cancer
341 stem -like cells in the PANC-1 cell line’. H Wang , J Wu , Y Zhang . *Oncology letters* 2012. 3 p. .
- 342 [Mckenzie et al. ()] ‘Understanding the IL-23-IL-17 immune pathway’. B S Mckenzie , R A Kastelein , D J Cua
343 . *Trends Immunol* 2006. 27 p. .
- 344 [Mckenzie ()] ‘Understanding the IL-23-IL-17 immune pathway’. B S Mckenzie . *Trends Immunol* 2006. 27 p. .