Artificial Intelligence formulated this projection for compatibility purposes from the original article published at Global Journals. However, this technology is currently in beta. *Therefore, kindly ignore odd layouts, missed formulae, text, tables, or figures.*

Miscarriage and Oxidative Stress of Iron Supplementation during Pregnancy

Ahmed M. Issa¹
A ¹ Jabir inb Hayyan Medical University
Received: 12 December 2015 Accepted: 4 January 2016 Published: 15 January 2016

7 Abstract

⁸ Background: Iron supplementations to the pregnant women in Iraq mostly considered as

⁹ obligatory routine health care additive treatment without real assessing to the iron status in

¹⁰ these people, therefore the acceptable range of iron must be estimated and the relationship

¹¹ between iron status and miscarriage should be explored. Objective: In miscarriage and

¹² pregnant women that supplemented with iron tablets the levels of oxidative stress should be

assessed and compared to the iron status to show the type and the strength of correlation

- ¹⁴ between them.Patients and methods: 96 pregnant women were participate in this study and
- ¹⁵ categorized into. First group: Pregnant iron supplemented group (PIS group) includes 35
- ¹⁶ pregnant women (36.46
- 17

18 Index terms— iron; oxidative stress; miscarriage; free radicals.

Objective: In miscarriage and pregnant women that supplemented with iron tablets the levels of oxidative stress should be assessed and compared to the iron status to show the type and the strength of correlation between them.

22 Patients and methods: 96 pregnant women were participate in this study and categorized into.

First group: Pregnant iron supplemented group (PIS group) includes 35 pregnant women (36.46%) during their 1 st trimester or middle of 2 nd trimester of pregnancy ??18-36 year) and they were received an oral iron supplementation as 250 mg of ferrous gluconate tablet, three times a day.

Second group: Pregnant iron non-supplemented group (PINS group) includes 31 pregnant women (32.29%) during their 1 st trimester or middle of 2 nd trimester of pregnancy ??19-37 year) and they were non supplemented with iron and they considered as control group.

Third group: Miscarriage iron supplemented group (MIS group) includes 30 miscarriage pregnant women(31.3%) during their 1 st trimester or middle of 2 nd trimester ??19-40 year) taken an oral iron supplementation as 250 mg of ferrous gluconate tablet, three times a day.

Results: The study at first assess the iron status by measuring Hb, serum ferritin and serum iron in the three 33 groups of women and shows that there is a significant differences p < 0.005 between oxidative stress parameters for the three stress parameters of the three stress parameters are stress of the three stress parameters are stress parameters.

of the miscarriage women and the pregnant women that taken oral supplements of iron tablets. The MDA and PC were higher in miscarriage in comparison to pregnant. The decrement in GSH level in miscarriage was also

significant (P value< 0.025). The correlations between serum iron concentration and oxidative stress parameters

37 MDA, PC and GSH were plotted using the regression line analysis where the r 2 values were obtained respectively

r 2 = 0.63, r 2 = 0.55 and r 2 = 0.70. It is obvious that there is a high correlation between the level of iron in the circulation and the oxidative stress of the reactive oxygen species that generated by the catalytic effect of this transition metal.

41 **1** Conclusion:

⁴² The high level of iron that obligatory supplemented to the pregnant women in Iraq should be restricted and early

43 careful investigation should be performed to assess the status of iron before any iron supplements prescribed.
44 The spontaneous abortion or miscarriage is highly

45 2 I. introduction

ron is an essential metal for hemoglobin biosynthesis of erythrocytes, oxidation-reduction reactions, and cellular 46 proliferation, whereas excess iron accumulation causes organ dysfunction through the production of reactive 47 oxygen species (ROS) and other mechanisms. The total amount of body iron is approximately 3-4 g in subject 48 with 70 kg, only 1-2 mg of iron is absorbed daily from the intestinal tract and circulated in the blood (1). Although 49 50 the body loses iron e.g through menstruation, this way does not considered as a systematic excretion route of 51 iron especially during pregnancy, and therefore the cumulative effect from supplementation can be dangerous; a continuous load exceeding 1-2 mg/day can eventually result in iron overload leading to organ failure and many 52 other complications (2,3). The benefits of improved maternal iron status by iron supplementation could be offset 53 by increased maternal and/or infant infection risk, due to increased availability of iron to host pathogens which 54 cause prenatal infections and miscarriage (4). 55

Miscarriage, also known as spontaneous abortion and pregnancy loss, is the natural death of an embryo or fetus before it is able to survive independently. Some use the cutoff of 20 weeks of gestation after which fetal death is known as a stillbirth. The most common symptoms of a miscarriage is vaginal bleeding with or without pain. Sadness, anxiety, and guilt may occur (5). Tissue or clot like material may also come out of the vagina (6). Ferrous iron is a principle pro-oxidant in human by being the most abundant transitional metal in human. It can initiate and potentiate the generation of free radicals through Fenton chemistry in which Fe +2 is oxidized to Fe +3 in existence of hydrogen peroxide and highly reactive hydroxyl free radical OH* is produced (7).

63 It is well known that the absorption of dietary iron occurs in the intestinal duodenum (8). This amount of absorbed dietary iron is enough to compensate for the estimated 1 to 2 mg of unregulated lose of iron through 64 sweat, dermal turnover, and incidental amounts excreted in urine (9,10). The need for iron varies markedly 65 during each trimester of pregnancy. Iron requirements decrease during the first trimester because menstruation 66 stops (11). During the second trimester, iron requirements begin to increase and continue to do so throughout 67 the remainder of pregnancy (12). As pregnancy progresses, iron requirements for fetal growth rise steadily in 68 69 proportion to the weight of the fetus, with most of the iron accumulating during the third trimester. The average 70 iron content of a fetus weighing >3 kg is ?270 mg (13). Pregnancy, mostly because of the mitochondrial-rich 71 (mitochondrial mass increase with gestational age), placenta is a conduction that favors oxidative stress transition 72 metals, especially iron, which is particularly abundant in placenta, are important in the production of free radicals . At the same time placenta is highly vascular and exposed to high maternal oxygen partial pressure (14). When 73 exposed to excess intake of iron, it is vulnerable to oxidative damage secondary to the continuous presence of 74 75 a relatively some excess of iron intake (15). This imbalance between pro-oxidants and antioxidants can lead to 76 a number of reproductive diseases such as endometriosis, polycystic ovary syndrome (PCOS), and unexplained infertility. Pregnancy complications such as spontaneous abortion, recurrent pregnancy loss, and preeclampsia, 77 78 can also develop in response to oxidative stress (16). Data from the United States National Health and Nutrition 79 Examination Survey (NHANES) in 1999-2006 for 1171 pregnant women showed that pregnant women in the 80 first trimester had the highest mean total body iron compared with that of pregnant women in the second or third trimesters (17,18). It has been reported by many investigators that the prevalence of fetal and maternal 81 82 complications is higher in women with excess iron complications or ?-thalassemia than in the general population (19)(20)(21)(22).83 In the presence of excess iron the highly reactive OH* (hydroxyl radical) species will be generated (23). In 84 1894, Fenton described the oxidizing potential of hydrogen peroxide when mixed with ferrous salts (24). Fe +2 +85 H 2 O 2 ? Fe + 3 + OH * + OH -86

Forty years later, in 1934, Haber and Weiss identified the hydroxyl radical as the oxidizing species in this reactions: $(25).O2^*$ -+ H 2 O 2 ? O 2 + OH -+ OH *

89 Overall summation of Fenton reaction in two steps gives the Haber-Weiss reaction ??-

90 — Fe +2 + H 2 O 2 ? Fe +3 + OH * + OH - Fe +3 + O * 2-? Fe +2 + O 2 - O *2-+ H 2 O 2 ? 91 O2 + OH -+ OH *

This reaction is often called the iron-catalyzed Haber-Weiss reaction, or sometimes the superoxidedriven Fenton reaction (25).

There are many different markers that can be used to demonstrate the presence of oxidative stress. In the current study reduced glutathione GSH, protein carbonyl group PC and malondialdehyde MDA all were used to monitors the oxidative stress because the free radicals are very unstable due to their high reactivity and cannot be practically determined at least by our humble laboratory (26). Protein carbonyl (PC) measurement is the most widely accepted possibility to evaluate the oxidative damage to the protein fraction as the content of carbonyl groups increases in free radical induced reactions (27,28).

The usage of (PC) groups as biomarkers of oxidative stress has some advantages in comparison with the measurement of other oxidation products because of the relative early formation and the relative stability of carbonylated proteins (29). Malondialdehyde is an aldehyde (3 carbon molecules with two aldehyde groups) (30). It is considered to be the terminal compound and the most important marker for monitoring lipid peroxidation and oxidative damage induced by ROS (31). About MDA Stewart AJ et al in 2005 mentioned that this naturally occurring end product of ROS is a marker of oxidative stress and used as a biomarker to determine oxidative stress level in organisms (32).

107 Glutathione is an essential nutrient for humans and, since it can be synthesized in the body from the amino

acids L-cysteine, L-glutamic acid, and glycine, it does not have to be present as a supplement in the diet (33).
The strong evidence that glutathione depletion causes cell death comes from the studies by Li and colleagues (34).
They emphasized that a decrease in GSH triggers the activation of neuronal 12-lipoxygenase pathway which leads
to the production of peroxides. However the direct depletion of cytoplasmic GSH resulted in increased generation
of ROS (35).

Blood hemoglobin, serum iron and ferritin parameters were used to assess the iron status in the body. After considering many indicators, a World Health Organization (WHO) and Centers for Disease Control (CDC) and technical consultation on the assessment of iron status at the population level concluded that serum iron, Hb and ferritin were the most efficient combination of indicators for monitoring changes in the iron status of a population as a consequence of iron supplementation (36).

The iron as a routine administered treatment, given to most pregnant women in Iraq, need to be studied 118 intensively to explore its effects on oxidation status in those subjects and which range would be acceptable and 119 safe for iron administration during pregnancy to prevent more pregnancy complications or miscarriage problems. 120 This was a 12 months cross-sectional study, conducted in alzahra teaching hospital in Al-Najaf and some other 121 private clinics, under supervision of specialist physicians, during 2014-2015. The study was designed in accordance 122 with the Ethics Committee approval before the start of the practical part and blood collection. The 96 pregnant 123 124 women were participate in this study and categorized into First group: Pregnant iron supplemented group (PIS 125 group) includes 35 pregnant women (36.46%) during their 1 st trimester or middle of 2 nd trimester of pregnancy 126 (18-36 year) and they were received an oral iron supplementation as 250 mg of ferrous gluconate tablet, three times a day. 127

¹²⁸ 3 Volume XVI Issue II Version I

Second group: Pregnant iron non-supplemented group (PINS group) includes 31 pregnant women (32.29%) during their 1 st trimester or middle of 2 nd trimester of pregnancy (19-37 yer) and they were non supplemented with iron and they considered as control group.

Third group: Miscarriage iron supplemented group (MIS group) includes 30 miscarriage pregnant women (31.3%) during their 1 st trimester or middle of 2 nd trimester (19-40 year)taken an oral iron supplementation as 250 mg of ferrous gluconate tablet, three times a day. The blood samples were drawn from iron supplemented group after 28 days from the time of starting the iron supplementation. The age of the pregnant women (supplemented and non supplemented group) was 18-40 years. The exclusion criteria contains diabetes mellitus, gestational hypertension, Rheumatoid, liver disease and hemolytic anemia which may interfere with our measurements.

Hemoglobin: Blood hemoglobin Concentration was determined by the Hb -Meter method. In the presence
of alkaline potassium ferricyanide, hemoglobin is oxidized to methaemoglobin, which then react with potassium
cyanide to form cyanmethaemoglobin complex which absorbe the wave length of 540 nm .The absorbance is
directly related to total hemoglobin concentration (37).

Serum Iron: After dissociation of iron-transferrin bound in acidic medium, ascorbic acid reduced Fe 3+ iron into Fe 2+ iron, Fe 2+ iron then form a colored complex with 3-(2pyridyl)-5,-6-difuryl-1,-2,-4-triazine-disulfonate (ferene). The absorbance thus measured at 600nm (580-620) which is directly proportional to the amount of iron

¹⁴⁶ in the specimen. Thiourea is added in the reagent to prevent copper interference (38).

Serum ferritin: The quantitative determination of circulating ferritin concentration in human sera was done
by the micro plate immunoenzymometric assay. The kit was purchased from Monobid inc, USA (39).

Serum malondialdehyde (MDA): level of serum malondialdehyde was determined spectrophotometrically by a modified procedure described by Guidet B. and Shah S.V. (40). The test is based on the reaction of MDA with thiobarbituric acid (TBA); forming MDA-TBA2 product that absorbs strongly at 532 nm.

Serum protein carbonyl (PC): Serum protein carbonyl (P.C) groups dimerizes with 2,4-dinitrophenylhydrazine
(DNPH), which leads to the formation of a stable dinitrophenyl hydrazone (DNP) product, that can be detected
by enzyme-linked immunosorbent assay (ELISA), We followed the method described by Levine et al in 1990 (41).
Serum reduced glutathione (GSH): 5,5-Dithiobis(2nitrobenzoic acid) (DTNB) is a disulfide chromogen that is

readily reduced by sulfahydryl group of GSH to an intensely yellow compound. The absorbance of the reduced thromogen is measured spectrophotometrically at 412 nm, which is directly proportion to the GSH concentration (42).

Statistical analysis: The results were expressed as Mean \pm SD. Student's t-test was used to examine the correlation of oxidative stress parameters (MDA, PC and GSH). Significant variation was considered when Pvalue was less than 0.05. Linear regression analysis was used to explore the correlation between the values of the oxidative stress parameters and serum iron in miscarriage women.

¹⁶³ 4 III. Results

In table 1 serum iron concentration and ferritin level both were raise in PIS group women if compared to the control group (PINS group) with p value < 0.005. The level of malondialdehyde MDA and the serum protein carbonyl PC were significantly enhanced p< 0.005 in pregnant women that taken iron supplements(PIS GROUP)

in comparison to the age matched control group that didn't take any iron supplementation during pregnancy. The

reduced glutathione GSH show a significant decrement P < 0.005 in pregnant women that taken iron regularly unlike those didn't take iron supplementation during pregnancy.

It is obvious from table 2 that serum iron concentration and ferritin level both were raise in MISG women if compared to the PIS GROUP women with p value < 0.025. The level of malondialdehyde MDA and the serum protein carbonyl PC were significantly enhanced p< 0.005 in miscarriage women that taken iron supplements (MISG) in comparison to the age matched pregnant women that taken a regular oral doses of iron supplementation during pregnancy (PIS GROUP). The reduced glutathione GSH show a significant decrement P< 0.025 in miscarriage women that taken iron regularly unlike the high level of GSH in pregnant group (PIS GROUP).

Table (1) : Blood hemoglobin conc., serum iron, ferritin, malondialdehyde, protein carbonyl groups and reduced glutathione in iron supplemented and non supplemented pregnant women. Table (??) : Blood hemoglobin conc., serum iron, ferritin, malondialdehyde, protein carbonyl group and reduced glutathione in pregnant iron supplemented and miscarriage iron supplemented women.

In figure 1 below the regression analysis was used to show the linear relationship between serum iron 180 concentration as a dependent variable and the MDA as the independent variable, however r = 0.63 and r = 2181 = 0.80. The correlation was positive and significant as shown from the plotted regression line and its equation. 182 In figure 2 the regression analysis was used to show the linear relationship between serum iron concentration as 183 184 a dependent variable and protein carbonyl groups as the independent variable, however r = 0.74 and r = 0.55185 The correlation was positive and progressive as shown in fig. 2 from the regression line and its equation. In 186 figure 3 the regression analysis was used to show the linear relationship between serum iron concentration as a dependent variable and serum reduced glutathione as the independent variable, however r = -0.84 and r = 2 = 0.70. 187 The correlation was negative and descendent as shown below in the regression line and its equation. 188

¹⁸⁹ 5 Volume XVI Issue II Version I

¹⁹⁰ 6 IV. Discussion

In the current study the oxidative stress parameters were considerably employed to explore the role of iron supplementation in pregnant and miscarriage women and to realize the consequences of oxidative stress load complications in those individuals. Pregnancy is a physiological state accompanied by higher energy demand and an increase in oxygen requirement, with various compensate enzyme adaptation (43). The changes occur with advancing pregnancy to met the increase demands for proper body function of mother to fulfill the requirement of fetus (44,45). The increase in oxygen demand is met by increase ventilation. Such conditions are responsible for raised oxidative stress in normal pregnancy (46).

The results in table 1 revel that there is a general decrement in oxidative stress in the pregnant women that not taken any supplementation of iron when compared with those supplemented with iron tablets and this fact may be related to the physiological changes that associated to the process of gestation itself. However Adiga U. et al in 2009 reported that the Pregnancy is a state, which is more prone for oxidative stress. Various studies reported that there is a development of a strong defense mechanisms against free radical damage, as the pregnancy progresses (47).

The enhancement of serum iron and ferritin levels in PIS group in table 1 is attributed to the supplements of iron tablets that taken orally. Hillman and Henderson in 1969 reported that with oral iron supplementation, patients were able to achieve serum iron values between 12.5 ?M and 27.0 ?M, and red blood cell production was able to increase to four or five times normal (48).

In table 1 the levels of MDA and PC in PIS group were significantly (p< 0.005) higher than that of PINS group this result were in consistent with the outcomes of many studies for example Minic oka in 2005 (49), noticed that during the periods of intravenous iron therapy the amount of iron promotes ironmediated formation of free radical species which could result in lipid peroxidation which give raise to MDA and oxidation of plasma proteins. He concluded an elevation in serum P.C groups after intravenous iron supplementation, through a study on evaluation of oxidative stress after intravenous iron supplementation.

The same upturn was noticed in table 2, the protein carbonyl groups (PC) in sera of miscarriage women (MIS 214 group) and serum MDA both were significantly (p < 0.005) higher than that of pregnant women in (PIS group) 215 The attribution involve the catalyzing role of iron as in Fenton reaction. Fenton reaction lead to generation of 216 highly reactive hydroxyl radicals which in turn would attack preferentially the proteins as a target molecules. 217 This reaction will result in the enhancement of protein carbonyl content (50,51). Iron serves as a catalyst for 218 protein oxidation and the formation of reactive oxygen species that in turn cause side chain modification of 219 220 many amino acid and ultimately to carbonyl groups formation (52). The higher serum iron concentration were in 221 coordinate with higher oxidative stress and raise in serum MDA concentration. This can be further demonstrated 222 by using the linear regression analysis as in figure 1 concluded that ,there is a positive correlation between MDA 223 level and serum iron status in young women (53).

Figure 2 shows a significant correlation between serum iron and PC groups in the circulation of the miscarriage women where r = 0.74 and $r^2 = 0.55$. This finding was also supported by M.J.Davies et al who mentioned that the protein carbonyls are formed early during oxidative stress conditions and are not a result of one specific oxidant, thus they can be called a marker of overall protein oxidation (54).

In table 1 GSH decreased in PIS group when compared to PINS group and this may be related to the differences

in serum iron concentrations in the two groups. In the same trend the results in table 2 reveal that the serum 229 GSH in miscarriage group was lower than that of pregnant women by ? 3.4% and the correlation between serum 230 iron and serum GSH was shown in figure 3 which display a significant negative regression linearity where r = -0.84231 and $r^2 = 0.70$. This decrement may be in coincidence with the speculations of Rajdl D. et al who documented 232 that the lower GSH levels may be due to the increased turnover of GSH for preventing oxidative damage in 233 iron supplemented group (55). The generation of free radicals is dependent on the presence of various transition 234 metal ions and the most important transition metal in vivo is believed to be the iron (56). Paik et al in 1999, 235 demonstrated that the higher level of ferritin have lower levels of GSH (negative correlation), and this conclusion 236 was obtained through a study on enhanced oxidative stress in hemodialysis patient receiving intravenous iron 237 therapy (57). Mustafa et al in 2010 detected markedly higher levels of MDA and significantly lower GSH levels 238 in the maternal blood of pregnant women (58). 239

Finally the oxidative stress that induced by iron supplementation in miscarriage as shown in table2 is often referred to by many other investigators. Many scientists indicated that Oxidative stress has also been implicated in early miscarriage. Jauniaux et al (59) suggested that early pregnancy loss may results from premature oxygenation of the early embryonic environment. By using an O 2 probe in women before first-trimester termination, they observed a steep rise in placental pO 2 between 8 and 12 wk of gestation.

Thus, it is well established that maternal metabolic disorders such as diabetes, which are associated with an increased generation of oxygen free radicals, are known to be associated with a higher incidence of miscarriages and fetal structural defects (60), indicating that the mammalian conceptus can be irreversibly damaged by oxidative stress (61).

Ruder EH in 2008 reported that in vitro fertilization is also affected by excessive ROS in embryo culture media, and the routine practice of incubating embryos at low oxygen tension can prevent embryo arrest and enhance the chances of successful fertilization (62).

²⁵² 7 V. Conclusion

Miscarriage in many pregnant women especially in the early first weeks of gestation affected significantly by 253 the iron that frequently and routinely prescribed to these women in Iraq by most physicians in the form of 254 different types of tablets and formulas. The oxidative stress that induced by the upturn of this transition metal 255 (Fe) in the circulation may be the real causative factor in the spontaneous abortion and many other pregnancy 256 complications. The iron supplements should be given to those who actually suffering from iron deficiency after 257 accurate diagnosis by the suitable biochemical tools. In addition to that the administration of excess iron should 258 be followed by supplying the necessary amount of antioxidant to neutralize the probable oxidation potential of 259 iron during pregnancy. 260

²⁶¹ 8 Volume XVI Issue II Version I

262

1

 $^{^{1}}$ $^{\odot}$ 2016 Global Journals Inc. (US) Miscarriage and Oxidative Stress of Iron Supplementation during Pregnancy



Figure 1:

Parameters	PINS GROUP NO = 31 (Control group)		PIS GROUP NO = 35		P <
	Mean ± SD	Range	Mean ± SD	Range	
Hb (g/dl)	11.50 ± 2.40	7.1-13.5	11.2 ± 1.70	7.3 –13.1	N.S
S.iron (µM)	16.50 ± 3.58	10.40- 24.02	21.03 ± 3.72	11.17-26.4	0.00
S.ferritin (ng /ml)	35.53 ± 15.25	14.75-65.99	54.11 ± 17.18	28.67-79.5	0.00
MDA (μM)	9.50 ± 2.43	5.16-13.96	13.43 ± 3.22	7.51-18.30	0.00
P.C nmol / mg . protein	2.62±0.72	4.41 - 1.60	3.41 ± 0.87	1.31-5.10	0.00
GSH (μM)	173.6 ± 20.34	135 - 225	154.4 ± 13.35	124-187	0.00

Figure 2: Figure 1 :

Parameters	PIS GROUP NO = 35		MIS GROUP NO = 30		P <
	Mean ± SD	Range	Mean ± SD	Range	
Hb (g/dl)	11.60 ± 1.70	7.30 –13.60	10.40 ± 1.76	7.60 -13.71	0.025
S.iron (μM)	21.03 ± 3.72	11.17-26.40	23.24 ± 3.41	14.17-28.30	0.025
S .ferritin (ng /ml)	54.11 ± 17.18	28.67-79.5	57.78 ± 14.57	30.57-82.20	0.025
MDA (µM)	13.43 ± 3.22	7.51-18.30	16.59 ± 3.28	8.10 -20.73	0.005
P.C nmol / mg prot	3.41 ± 0.87	1.31-5.10	4.96 ± 0.98	1.65 – 6.25	0.005
GSH (µM)	154.4 ± 13.35	124-187	149.3 ± 11.62	125.3-160.7	0.025

Figure 3: Figure 2 :

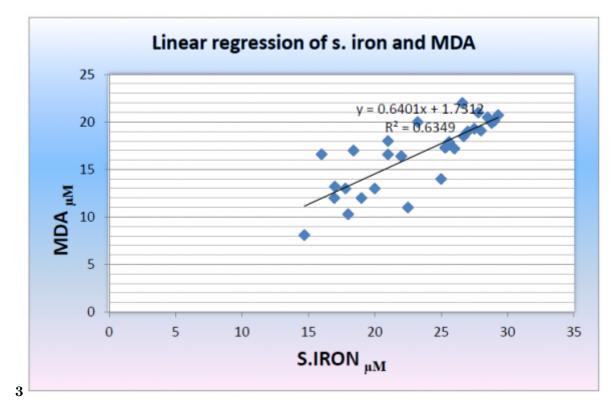


Figure 4: Figure 3 :

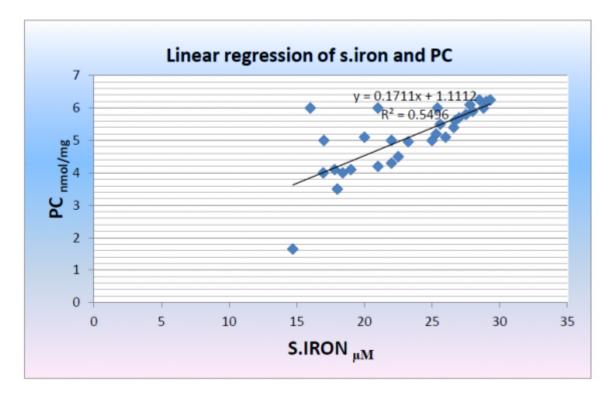


Figure 5:

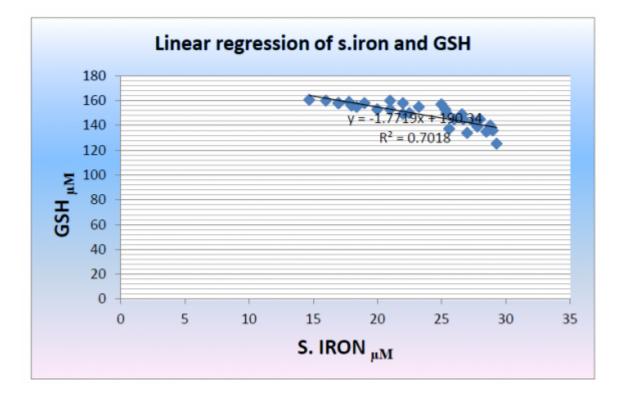


Figure 6:

- 263 [Dalle-Donne] , I Dalle-Donne .
- 264 [Dalle-Donne] , I Dalle-Donne .
- 265 [Rossi] , R Rossi .
- ²⁶⁶ [Guidet and Shah ()], B Guidet, S V Shah. Am J Physiol 1989. 257 (26) p. 440.
- 267 [Starkee-Reed and Oliver ()], P E Starkee-Reed, C N Oliver. Arch. Biochem. Biophys 1989. 275 p. .
- 268 [Fisher and Stadtman ()], M T Fisher, E R Stadtman. J. Biol. Chem 1992. 267 p. .
- [Mei et al. (2011)], Z Mei, M E Cogswell, A C Looker, C M Pfeiffer, S E Cusick, D A Lacher, L M
 Grummer-Strawn. Am J Clin Nutr 2011 Jun. 93 (6) p. .
- [Perry et al. ()] 'A metabolic basis for Alzheimer disease'. G Perry , A Nunomura , A K Raina . Neurochem Res
 2003. 28 p. .
- 273 [Rio et al. ()] 'A review of recent studies on malondialdehyde as toxic molecule and biological marker of oxidative

stress'. Del Rio , D Stewart , A J Pellegrini , N . Nutr Metab Cardiovasc Dis 2005. 4 p. .

- [Li et al. ()] 'A role for 12-lipoxygenase in nerve cell death caused by glutathione depletion'. Y Li , P Maher , D
 Schubert . Neuron 1997. 19 p. .
- [Assessment of iron status in US pregnant women from the National Health and Nutrition Examination Survey (NHANES)]
 Assessment of iron status in US pregnant women from the National Health and Nutrition Examination
 Survey (NHANES), p. .
- [Mustafa et al. ()] 'Association of glutathione S transferase M1 and T1 gene polymorphisms and oxidative stress
 markers in preterm labor'. M D Mustafa , R Pathak , T Ahmed , R S Ahmed , A K Tripathi , K Guleria , B
 D Banerjee . *Clin Biochem* 2010. 43 p. .
- [West ()] Best in Taylor's Physiological Basis of Medical Practice, John B West . 1991. p. 894. (12th ed)
- [Kohgo et al. ()] 'Body iron metabolism and pathophysiology of iron overload'. Yutaka Kohgo , Katsuya Ikuta ,
 Takaaki Ohtake . Int J Hematol 2008. 88 p. .
- [Gitto et al. ()] 'Causes of oxidative stress in the perinatal period'. E Gitto , R J Reiter , M Karbownik . *Biol. Neonate* 2002. 81 (3) p. .
- [Lund et al. ()] 'Chonic exposure to high levels of dietary iron fortification increases lipid peroxidation in the
 mucosa of the rat large intestine'. E K Lund , S J Fairweather-Tait , S G Warf . J Nutr 2001. 131 p. .

[Makarem (ed.) ()] Clinical chemistry -principles and techniques .second, A Makarem . Ed.RF. Henry, D.C.
 Cannon, j.w. winkelman (ed.) 1974. Hagerstown (MD: Harper and Row. p. .

- 292 [Tietz (ed.) ()] Clinical Guide to laboratory, N W Tietz . test. 3 rd Ed.N.W. (ed.) 1995.
- [Hillman and Henderson ()] 'Control of marrow production by the level of iron supply'. R S Hillman , P A
 Henderson . J Clin Invest 1969. 48 p. .
- [Sarah M King ()] daily supplementation with iron increase lipid peroxidation in young women with iron store
 Experimental biology and medicine, Carmen M Sarah M King , Donangelo . 2008. 233 p. .
- [Davies et al. ()] 'Dean Stable markers of oxidant damage to proteins and their application in the study of human
 disease'. M J Davies , S Fu , H Wang , RT . *Biol. Med* 1999. 27 p. .
- [Levine and Oliver ()] 'Determination of carbonyl content in oxidatively modified proteins'. R L Levine , Garland
 D Oliver , CN . Meth Enzymol1990. 186 p. .
- 301 [Andrews ()] 'Disorders of iron metabolism'. N C Andrews . N Engl J Med 1999. 341 p. .
- 302 [Imlay and Linn ()] 'DNA damage and oxygen radical toxicity'. A J Imlay , S Linn . Science 1988. 240 p. .
- [Thomas et al. ()] 'Elevated Iron Indices in Patients with Diabetes'. M C Thomas , R J Maclsaac , C Tsalamandris
 Diabet Med 2004. 21 (7) p. .
- [Toumba et al. ()] 'Endocrine complications in patients with thalassaemia major'. M Toumba , A Sergis , C
 Kanaris , N Skordis . *Pediatr Endocrinol Rev* 2007. 5 (2) p. .
- [Lim et al. ()] 'et al Enhanced oxidative stress in hemodialysis patient receiving intravenousiron therapy'. Paik Seong Lim , Yau-Huei Wei , Yu York Leng . nephrol Dial Transplant 1999. 14 p. .
- [Minic -Oka ()] Evaluation of oxidative stress after intravenous iron supplementation. renal failure, Minic -Oka
 . 2005. 27 p. .
- [Wardman and Candeias ()] 'Fenton centennial symposium'. P Wardman , P L Candeias . Radiat Res 1996. 145
 p. .
- 313 [White et al. ()] 'Fluorescence-based microtiter plate assay for glutamate-cysteine ligase activity'. C C White, H
- Viernes, C M Krejsa, D Botta, T J Kavanagh. 10.1016/S00032697(03)00143-X.PMID12814619. Analytical
 Biochemistry 2003. 318 (2) p. .

8 VOLUME XVI ISSUE II VERSION I

- [Halliwell and Gutteridge ()] 'Free radicals, antioxidants and human disease: Where are we now?'. B Halliwell ,
 C M Gutteridge . J Lab Clin Med 1992. 119 p. .
- 318 [David ()] 'Genetic emochromatosis'. V David . Ann. Boil.clin 1997.
- [Wuèllner et al. ()] 'Glutathione depletion and neuronal cell death: the role of reactive oxygen intermediates and
 mitochondrial function'. U Wuèllner , J Seyfried , P Groscurth , S Beinroth , S Winter , M Gleichmann , M
 Heneka , P Loèschmann , J B Schulz , M Weller , T Klockgether . Brain Res 1999. 826 p. .
- [Halliwell and Gutteridge ()] B Halliwell , Jmc Gutteridge . Free Radicals in Biology and Medicine, (Oxford)
 1989. Clarendon Press. 333 p. 56. (2nd ed)
- [Casanueva and Viteri ()] 'Iron and oxidative stress in pregnancy'. E Casanueva , F E Viteri . J Nutr 2003. 133
 p. .
- Beard ()] 'Iron biology in immune function, muscle metabolism and neural functioning'. J L Beard . JNutr 2001.
 131 p. .
- 328 [Iron in Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic Silicon. Vanadium and Zinc ()] 'Iron in
- Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic'. Silicon. Vanadium and Zinc 2001. National
 Academy Press. p. .
- [Hallberg and Rossander-Hultén ()] 'Iron requirements in menstruating women'. L Hallberg , L Rossander-Hultén
 Am J Clin Nutr 1991. 54 p. .
- [Valerio and Luis ()] 'Mammalian Iron Metabolism'. ValerioJr , G Luis . Toxicology Mechanisms and Methods
 2007. 17 (9) p. .
- [Rajdl et al. ()] 'Markers of oxidative stress in diabetic mothers and their infants during delivery'. D Rajdl , J
 Racek , A Steiherova . *Physiol Res* 2005. 54 (4) p. .
- [Cighetti et al. ()] 'Mechanism is action of malondialdehyde and 4-hydroxyl non enal on Xanthine oxidoreduc tase'. G Cighetti , L Bortone , S Sala . Arch Biochem Biophys 2001. 389 (2) p. .
- Jauniaux et al. ()] 'Onset of maternal arterial blood flow and placental oxidative stress: a possible factor in human early pregnancy failure'. E Jauniaux , A L Watson , J Hempstock , Y-P Bao , J N Skepper , G J Burton . Am J Pathol 2000. 157 p. .
- [Brissot and Deugnier ()] Oxford Textbook of Clinical Hepatology, P Brissot , Y Deugnier . 1999. Oxford: Oxford
 University Press. p. .
- [Ruder et al. ()] 'Oxidative stress and antioxidants: exposure and impact on female fertility'. E H Ruder , T J
 Hartman , J Blumberg , M B Goldman . *Reprod Biomed Online* 2008. 14 p. . (Hum Reprod Update)
- Savona-Ventura and Grech ()] 'Pregnancy complications in homozygous thalassaemia patients'. C Savona Ventura , E Grech . J Obstet Gynaecol 1991. 11 p. .
- [Tuck et al. ()] 'Pregnancy management and outcomes in women with thalassaemia major'. S M Tuck , C E
 Jensen , B Wonke , A Yardumian . J Pediatr Endocrinol Metab 1998. 11 p. .
- [Giustarini ()] 'Protein carbonyl groups as biomarkers of oxidative stress'. D Giustarini . Clin Chim Acta 2003a.
 329 p. .
- [Colombo ()] 'Protein carbonylation in human diseases'. Giustarini D Colombo , REt . Trends Mol Med 2003b.
 9 p. .
- [Who/Cdc ()] 'Report of a joint World Health Organization/Centers for Disease Control and Prevention technical
 consultation on the assessment of iron status at the population level'. Who/Cdc . World Health Organization
 and Centers for Disease Control and Prevention 2005. 2005. p. . (Centers for Disease Control and Prevention)
- [Mashaal ()] Study of serum ferritin and other hematological parameter in pregnancy, Al-Toub Mashaal . 2006.
 108 p. .
- [Burtis and Ashwood ()] 'Text Book of Clinical Chemistry'. C A Burtis , E R Ashwood . *Philadelphia WB SAUNDERS* 1999. p. . (3rd ed.)
- [Cunningham et al. ()] 'Thalassemia Clinical Research Network. Complications of ?-thalassemia major in North
 America'. M J Cunningham , E A Macklin , E J Neufeld , A R Cohen . *Blood* 2004. 104 (1) p. .
- [Ashok et al. (2012 v10)] The effects of oxidative stress on female reproduction: a review biomedical central, A
 Ashok , A Anamar , Mellado , J Beena , Premkumar . 2012 v10. p. .
- [Kose et al. ()] 'The evaluation of lipid peroxidation and adensine Deaminase activity in patient with Behcehs
 disease'. K Kose , C Yazici , O Assioglu . Clin Bio Chem 2001. 34 (2) p. .
- [Amirkhizi et al. ()] 'the relation of body iron stores and oxidative stress markers related toatherosclerosis in women of reproductive ARYA Atherosclerosis'. Farshad Amirkhizi , Fereydoun Siassi , Sara Minaie . *Journal*
- 371 2008. 4 (4) p. .

- [Burton et al. ()] 'The role of alterations in arachidonic acid metabolism and nitric oxide homeostasis in rat models of diabetes during early pregnancy'. G J Burton , J Hempstock , E ; Jauniaux , A Jawerbaum ,
- GonzalesE . *Curr Pharm Des* 2003. 2005. 11 p. . (Oxygen, early embryonic metabolism and free 61)
- [Buss et al. ()] 'The role of iron chelating in the cancer therapy'. J L Buss , F M Torti , S V Toriti . *curr Med Chem* 2003. 10 p. .
- [Lao et al. ()] Third trimester iron status and pregnancy outcome in non-anemic women. pregnancy unfavourably
 affected by maternal iron excess Human Reproduction, K.-F Lao , L Y Tam , Chan . 2000.
- [Adiga and Adiga ()] 'Total Antioxidant Activity in Normal Pregnancy'. U Adiga , Mns Adiga . Online J Health
 Allied Scs 2009. 8 (2) p. 8.
- [Pernoll et al. ()] 'Ventilation during rest and exercise in pregnancy & Post partum Respir'. M L Pernoll , J
 Metcalfe , P A Kovacl . *Physiol* 1975. 295 p. 25.
- [What are the symptoms of pregnancy loss/miscarriage? (2015)] What are the symptoms of pregnancy
 loss/miscarriage?, http://www.nichd.nih.gov/.2013-07-15 14 March 2015.
- 385 [What is pregnancy loss/miscarriage? (2015)] What is pregnancy loss/miscarriage?, http://www.nichd.
- nih.gov/.2013-07-15 14 March 2015. (Jump up to: a b)