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Role of Heme-Oxygenase-1 in Attenuated Cardioprotective Effect of Ischemic Preconditioning in Hyperlipidemic Myocardium

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7 Abstract

Nitric Oxide (NO) is responsible for cardioprotective effect of ischemic preconditioning (IPC). 8 Heme Oxygenase-1 (HO-1) facilitates release of NO by disrupting caveolin-eNOS complex. Both, the expression/activity of HO-1 and the IPC mediated cardioprotection are decreased 10 significantly in hyperlipidemia. In this study the role of HO-1 in attenuation of IPC- induced 11 cardioprotective effect in hyperlipidemic rat was investigated. Hyperlipidemia was induced by 12 feeding high fat diet to Wistar rats. Isolated Langendorff heart preparation model was used. 13 Cardioprotective effect was assessed by myocardial infarct size measurement and release of 14 Lactate Dehydrogenase (LDH), Creatine Kinase (CK-MB) in coronary effluent. Nitrite 15 estimation was done to indirectly infer the level of cardiac NO production. In hyperlipidemic 16 rat, IPC-induced cardioprotection and release of NO were significantly decreased. Perfusion 17 with sodium nitrite (NO precursor) and pre-treatment with daidzein (DDZ) (caveolin 18 inhibitor) and hemin (HO-1 inducer), alone or in combination significantly restored the 19 attenuated cardioprotective effect of IPC in hyperlipidemic rats. Administration of zinc 20 protoporphyrin (ZnPP), HO-1 inhibitor, significantly abolished the observed cardioprotection 21 in hemin pre-treated hyperlipidemic rat. The significant restoration of the attenuated 22 cardioprotective effect of IPC following induction of HO-1 by hemin in hyperlipidemia was 23 observed. The results indicated that attenuation of IPC-induced cardioprotective effect may 24 be due to the decrease in HO-1 induced NO release in hyperlipidemic rat heart. 25

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27 Index terms— heme oxygenase-1, ischemic preconditioning, hyperlipidemic rat heart, hemin, diadzein.

²⁸ 1 I. Introduction

oronary artery disease is a leading cause of morbidity and mortality worldwide [1,2]. Inadequate blood flow to the myocardium leads to ischemia and, early reperfusion is necessary for the viability of myocardium [3]. Reperfusion after a prolonged period of ischemia is not without risk, it damages the myocardium, which is known as ischemiareperfusion injury [4,5]. Ischemic preconditioning (IPC), is a powerful endogenous cardioprotective phenomenon in which short intermittent cycles of sublethal ischemia, followed by reperfusion before the subsequent prolonged ischemic insult, improves the tolerance against ischemia-reperfusion-induced injury [6,7]. IPC mediated cardioprotection has been documented in various species including human beings [7,8].

³⁶ IPC produces cardioprotection by stimulating the generation of various endogenous ligands which bind ³⁷ to their respective G-protein coupled receptors [9,10] and initiate a signalling cascade i.e., activation of PI-³⁸ 3K/Akt [11], phosphorylation of eNOS, generation of NO and by opening of mito K ATP channel [12,13]. The ³⁹ cardioprotective effect of IPC is attenuated in in hyperlipidemic myocardium and it may be due to decreased HO-⁴⁰ 1 [14,15], impairment of K ATP channel [16] impairment of PI-3K/AKT pathway [17,18] and altered activation ⁴¹ of JAK/STAT and MAPK, GSK-3? [19,20]. Hence, the mechanism involved in attenuation of cardioprotective

42 effect of IPC in hyperlipidemic myocardium, remain to be elucidated.

43 Caveolae are the specialised membrane domains which serve as organizing centres for cellular signal 44 transduction [21]. Various signalling molecules like src-like kinases, tyrosine kinase, members of Ras-MAPK 45 cascade and eNOS [22] are localized within caveolae. Caveolin is also a well known negative regulator of eNOS 46 and these results in decreased availability of NO [23,24] which is responsible for cardioprotective effect of IPC 47 [13]. It has been reported that expression of caveolin is upregulated in hyperlipidemic myocardium [25].

Heme-oxygenase is the rate-limiting enzyme in the biochemical pathway responsible for catabolism of heme into ferrous (Fe ++) ion, carbon monoxide, and biliverdin, the latter being subsequently converted into bilirubin by biliverdin reductase [26]. HO-1 is localized in the membrane caveolae and the inner leaflet of the plasma membrane where it is interacts with caveolin [27]. In transgenic mice, the overexpression of Hemeoxygenase-1, conversely regulates the expression of caveolin [25]. Moreover, HO-1 facilitates release of NO by disrupting association of caveolin with eNOS [25]. It has been reported that a decrease in the cardiospecific expression of HO-1 exacerbates the ischemia reperfusion-induced injury [26], while upregulation of HO-1 produces cardioprotection against ischemiareperfusion induced injury [27]. Transgenic mice expressing cardiac-specific HO-1 are resistant, while the heart of HO-1 knock-out mice is more susceptible to ischemia-reperfusion-induced injury [28]. In hyperlipidemia, the expression and activity of HO-1 is reduced [29] whereas the increase in HO-1 in hyperlipidemic rats is associated with activated eNOS [30]. Therefore, the present study was designed to investigate the role of

rats is associated with activated eNOS [30]. Therefore, the present study was designed to investigate the ro
 Heme-Oxygenase-1 in attenuated cardioprotective effect of IPC in hyperlipidemic rat hearts.

⁶⁰ 2 II. Materials and Methods

Daidzein (0.2mg/Kg/s.c) (Enzo Life Sciences International, Inc., USA) was dissolved in 10% Dimethyl Sulphoxide 61 (DMSO) and then injected to the animals for 7 days after 8 weeks of high fat diet administration. Hemin 62 (4mg/kg/i.p.) (Himedia Laboratories Pvt. Ltd., Mumbai) was dissolved in 0.2M NaOH and was injected 63 18 h before isolation of heart. Zinc Protoporphyrin (50µg/kg/i.p.) (Enzo Life Sciences International, Inc., 64 USA) was dissolved in DMSO and injected 6 hr before hemin treatment [31]. TTC Stain, Tris-chloride buffer, 65 sulphanilamide, phosphoric acid and sodium nitrite was purchased from CDH Pvt. Ltd., New Delhi. N-(1-66 Naphthyl) ethylenediamine dihydrochloride was purchased from Himedia Laboratories Pvt. Ltd., Mumbai. The 67 LDH enzymatic estimation kit and CK-MB enzymatic estimation kit was purchased from Coral Clinical Systems, 68 Goa, India. All other reagents used in this study were of analytical grade and always freshly prepared before use. 69

$_{70}$ 3 a) Animals

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 $_{\rm 71}$ $\,$ Age matched young male Wistar rats, weighing 180-250 g housed in animal house and provided 12 h light and

72 12 h dark cycle were used. They were fed on standard chow diet (Ashirwad Industries Ltd., Ropar, India)

- and provided water ad libitum. The experimental protocol was approved by the Institutional Animal Ethics
 Committee in accordance with the National (CPCSEA) Guidelines on the Use of Laboratory animals. All efforts
- were made to minimize animal suffering and reduce the number of animals used.

⁷⁶ 4 b) Induction of experimental hyperlipidaemia

Male Wistar rats (180-250) were employed in the present study. Experimental hyperlipidemia was induced by high fat diet (corn starch 44.74 g, casein 14 g, sucrose 10 g, butter 20 g, fibre 5 g, mineral mix 3.5 g, vitamin mix 1 g, choline 0.25 g, terbutylhydroquinone 0.0008 g, cholesterol 1 g, cholic acid 0.5 g) for 8 weeks. Serum cholesterol and triglyceride level was estimated spectrophotometrically at 505 nm by PEG and GPO / PAP method ??Trinder, 1969; ??ucolo, 1973 ?? Fossati, 1982) using enzymatic kits (Coral Clinical Systems, Goa, India). Serum cholesterol level 800-1000 mg/dl and serum triglyceride level 200-300 mg/dl were considered to be hyperlipidemic.

$_{84}$ 5 c) Isolated rat heart preparation

Rats were administered heparin (500 IU/L, i,p) 20 min, prior to sacrificing the animal by cervical dislocation. 85 Heart was rapidly excised and immediately mounted on Langendorff's apparatus [30]. Isolated heart was 86 retrogradely perfused at constant pressure of 80 mmHg with Kreb's-Henseleit buffer (NaCl 118 mM; KCl 4.7 87 mM; CaCl 2 2.5 mM; MgSO 4 .7H 2 0 1.2 mM; KH 2 PO 4 1.2 mM; C 6 H 12 O 6 11 mM), pH 7.4, maintained 88 at 37? bubbled with 95% O 2 and 5% CO 2. Flow rate was maintained at 7-9 ml/min. using Hoffman's screw. 89 The heart was enclosed in double wall jacket, the temperature of which was maintained by circulating water 90 heated at 37?. Ischemic preconditioning was produced by closing the inflow of K-H solution for 5 min followed 91 by 5 min of reperfusion. Four such episodes were employed. Global ischemia was produced for 30 min. followed 92 by 120 min. of reperfusion. Coronary effluent was collected before ischemia, immediately, 5 min. and 30 min. 93 after reperfusion for estimation of LDH, CK-MB and nitrite release [32]. 94

⁹⁵ 6 d) Assessment of myocardial injury

The assessment of myocardial infarct size was done by using triphenyltetrazolium chloride (TTC) staining method.
 The heart was removed from the Langendorff's apparatus. Both the atria and root of aorta were excised and

ventricles were kept overnight at -4? temperature. Frozen ventricles were sliced into uniform sections of about

1-2 mm thickness. The slices were incubated in 1% w/v triphenyltetrazolium chloride stain (TTC stain) at 37? in 99 0.2M Tris-chloride buffer for 30 min. The normal myocardium was stained brick red while the infarcted portion 100 remained unstained. Infarct size was measured by the volume method [33]. LDH and CK-MB were estimated 101 by using commercially available kits. Values of LDH and CK-MB were expressed in international units per litre 102

(IU/L).103

e) Nitrite estimation 7 104

Nitrite is stable nitrogen intermediate formed from the spontaneous degradation of NO. Unlike NO, nitrite can 105 be measured easily and nitrite concentrations can be used to infer levels of NO production. Nitrite release in 106 coronary effluent was measured. Greiss reagent 0.5 ml (1:1 solution of 1% sulphanilamide in 5% phosphoric acid 107 and 0.1% N-(1-Naphthyl) ethylenediamine dihydrochloride in water) was added to 0.5 ml of coronary effluent. 108 The optical density at 550 nm was measured using spectrophotometer (UV-1700 Spectrophotometer, Shimadzu, 109 Japan). Nitrite concentration was calculated by comparison with spectrophotometer reading of standard solution 110 of sodium nitrite prepared in K-H buffer [32]. Diagrammatic representation of experimental protocol is shown. 111 In all groups, isolated rat heart was perfused with K-H (Krebs-Hensleit) solution and allowed for 10 min of 112 stabilization. Isolated rat heart preparation was stabilized for 10 min and then perfused continuously with K-H 113 solution for 190 min. 114

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g) Data analysis and statistical procedures 9 116

All values were expressed as mean \pm standard deviation (S.D). Statistical analysis was performed using Graphpad 117 Prism Software (5.0). The data obtained from the various groups were statistically analysed using student t-test, 118 one-way analysis of variance (ANOVA), two way analysis of variance (ANOVA) followed by Tukey's multiple 119 comparison test. P? 0.05 was considered to be statistically significant. 120

III. Results 10121

a) Effect of high fat diet on body weight 11 122

The high fat diet was fed for 8 weeks to the rats and a significant increase in body weight was observed as 123 compared to basal value (Fig. ??). Values are expressed as mean \pm S.D. 124

c) Effect of ischemic preconditioning and pharmacological 12125 interventions on myocardial injury (Infarct size, LDH and 126 CK-MB) 127

Global ischemia for 30 min followed by 120 min of reperfusion significantly increased the myocardial injury as 128 compared to sham control. Four episodes of IPC significantly decreased I/R-induced increase in myocardial 129 injury in normal rat heart. However, ischemic preconditioning failed to decrease the myocardial injury 130

13interventions on the nitrite release in coronary effluent 131

Global ischemia for 30 min followed by 120 min of reperfusion significantly decreased the nitrite release. Four 132 episodes of IPC significantly restored the I/R induced decrease in nitrite release in normal rat heart. However, 133 ischemic preconditioning failed to increase the nitrite release in hyperlipidemic rat heart. Moreover, IPC induced 134 increase of nitrite release was significantly restored in sodium nitrite perfused hyperlipidemic rat heart. Pre-135 treatment with daidzein and hemin, alone or in combination also restored it. Furthermore, administration of 136 ZnPP significantly abolished the restored cardioprotective effect of hemin in hyperlipidemic rat (Fig. 6). 137

IV. Discussion 14 138

This study was designed to investigate the role of HO-1 in attenuated cardioprotective effect of IPC in 139 hyperlipidemic rat hearts. After a prolonged period of ischemia, reperfusion produces further damage to 140 myocardium which is known as ischemia reperfusion injury. The ischemic preconditioning induced by four episodes 141 142 of 5 min global ischemia and 5 min reperfusion was reported to produce cardioprotective effect in isolated rat heart preparation [34]. Our findings were in agreement with these phenomenon's. The cardioprotective effect 143 144 of IPC had been reported to be significantly attenuated in hyperlipidemia. Our results were in accordance with 145 these published studies [35].

Perfusion of sodium nitrite (NO donor) produces cardioprotection in isolated heart from normal rat, subjected 146 to global ischemia [36]. In our study, perfusion of sodium nitrite in isolated hyperlipidemic rat heart followed by 147 IPC, significantly restored the attenuated effect of IPC in diabetic myocardium. 148

Release of nitric oxide during the ischemic preconditioning was reported to produce cardioprotection against 149 ischemia-reperfusion induced injury [12]. In our study, IPC significantly increased the release of NO (measured 150

in coronary effluent), as compared to ischemia reperfusion control group. However, this IPC mediated increase 151 in release of nitric oxide was significantly decreased in hyperlipidemic rat heart. Sodium nitrite perfusion in 152 hyperlipidemic rat heart significantly restored the attenuated cardioprotective effect of ischemic preconditioning. 153 Thus, the reduced release of NO in hyperlipidemic rat heart may be responsible for attenuation of cardioprotection 154 mediated by IPC in hyperlipidemic rat. It was interesting to note that treatment with sodium nitrite did not 155 enhance the cardioprotective effect of IPC in normal rat heart. This indicated that once IPC mediated increased 156 generation of NO achieved the threshold for cardioprotection; addition of sodium nitrite was unable to further 157 increase the myocardial protection by IPC. Caveolae are 50-100 nm invaginated plasma membrane domains which 158 serve as organizing centers of signal transduction [37]. Caveolins are proteins that form the structure of caveolar 159 membrane, act as signalosomes for GPCR and other molecules such as NOS and Src-like kinases [38]. Increased 160 expression of caveolin, leads to the, decreased phosphorylation of endothelial nitric oxide synthase and consequent 161 decreased generation of nitric oxide. Further, it has been reported that expression of caveolin is upregulated in 162 hyperlipidemic myocardium [39]. Thus, it may results in increased formation of Caveolin-eNOS complex, which 163 decreases the availability of nitric oxide. It has been reported that NO is responsible for cardioprotective effect 164 of ischemic preconditioning [40]. 165

¹⁶⁶ Upregulation of caveolin in diabetic rat heart may inhibit the activity of eNOS by making its complex which ¹⁶⁷ leads to a decrease in the release of NO [41]. Administration of daidzein increases the generation of nitric ¹⁶⁸ oxide by inhibiting the caveolin-eNOS complex and subsequent activation of the eNOS [42]. In our study, one ¹⁶⁹ week of pretreatment of hyperlipidemic rat with daidzein, a caveolin inhibitor [42], significantly restored the ¹⁷⁰ cardioprotective effect of ischemic preconditioning in hyperlipidemic rat heart, noted in terms of decrease in ¹⁷¹ infarct size and release of LDH, CKMB, and also increase in the release of NO. Our findings were in agreement ¹⁷² with reports from other laboratories [20].

Heme-Oxygenase-1 is localized in the membrane caveolae of the plasma membrane where it is interacts with caveolin [27]. It has been reported that a decrease in the cardiospecific expression of HO-1 exacerbates while an upregulation of HO-1 produces cardioprotection against ischemia-reperfusion injury [43]. HO-1 facilitates release of NO by disrupting complex of caveolin and eNOS [43]. The expression of HO-1 is diminished into hyperlipidemic myocardium. In our study, pretreatment with hemin, a heme-oxygenase-1 inducer, restored the decrease in release of nitric oxide and significantly restore the attenuated cardioprotective effect of ischemic preconditioning in hyperlipidemic rat heart.

Thus it was speculated that the attenuated cardioprotective effect of IPC in hyperlipidemic rat heart may be due to inhibition of eNOS by enhancing the binding of eNOS with caveolin, which leads to decrease in the release of nitric oxide. Also, administration of ZnPP, an inhibitor of HO-1, significantly blocked the observed cardioprotection and increase in release of NO in hearts of hemin pretreated hyperlipidemic rats. Furthermore, the restoration of the attenuated cardioprotective effect of IPC in hyperlipidemic rat heart by combination of daidzein and hemin was not greater than that observed when the drugs were administered alone. This suggested that these two drugs may be acting via the same mechanism i.e., NO pathway.

On the basis of above discussion it was clear that activation of heme-oxygenase-1 enzyme, by a specific inducer i.e. hemin, restored the cardioprotective effect of ischemic preconditioning in hyperlipidemic rat heart, by disrupting the caveolin-eNOS complex and there by enhancing the release of NO. Further, pretreatment with ZnPP, a specific heme-oxygenase-1 inhibitor, significantly blocked the restoration of cardioprotective effect of ischemic preconditioning in hemin pretreated hyperlipidemic rat heart. Therefore, it was concluded that attenuation of cardioprotective effect of ischemic preconditioning in hyperlipidemic rat heart, was due to impairment of HO-1 induced release of nitric oxide.

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Figure 3: Fig. 3 : Fig. 4 : Fig. 5 :

10' S	40′ p	30'1	120′ R
Figure 4:			
6 10' 5 5'1	5' R 5' 5' R 5'	5'R 5'I 5'R 30'I	120' R
		Figure 5: Fig. 6 :	

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