



GLOBAL JOURNAL OF MEDICAL RESEARCH: K
INTERDISCIPLINARY
Volume 17 Issue 4 Version 1.0 Year 2017
Type: Double Blind Peer Reviewed International Research Journal
Publisher: Global Journals Inc. (USA)
Online ISSN: 2249-4618 & Print ISSN: 0975-5888

Prevalence of Antimicrobial Resistance among Gram-Negative Isolates in an Adult Intensive Care Unit at a Tertiary Care Center in Saudi Arabia (2010-2014)

By Roaa R Amer, Bayan T Alzomaili, Rawan M AlTuwaijri, Rana S AlZahrani,
Samaher H AlHarbi, Alaa AlThubaiti & Sameera M Al Johani

King Saud Bin Abdulaziz University for Health Sciences

Abstract- Introduction: Infections caused by multidrug resistant (MDR) organisms can result in significant increases in morbidity and mortality. This risk is amplified in critically ill patients usually residing in intensive care units (ICU).

Methods: A retrospective cross-sectional study was conducted to explore the progression of antimicrobial resistance of Gram negative bacteria (GNB) in a tertiary care hospital in Riyadh, Saudi Arabia. All organisms were isolated from the adult ICU of King Abdulaziz Medical City between 2010 to 2014. Organisms were identified to the species level. Antimicrobial susceptibility testing was performed using an automated system (The VITEK® 2 system, BioMérieux, France) and the antimicrobial susceptibility testing was confirmed by E-Test.

GJMR-K Classification: NLMC Code: WX 218



Strictly as per the compliance and regulations of:



Prevalence of Antimicrobial Resistance among Gram-Negative Isolates in an Adult Intensive Care Unit at a Tertiary Care Center in Saudi Arabia (2010-2014)

Roaa R Amer ^α, Bayan T Alzomaili ^σ, Rawan M AlTuwaijri ^ρ, Rana S AlZahrani ^ω, Samaher H AlHarbi [¥], Alaa AlThubaiti [§] & Sameera M Al Johani ^x

Abstract- Introduction: Infections caused by multidrug resistant (MDR) organisms can result in significant increases in morbidity and mortality. This risk is amplified in critically ill patients usually residing in intensive care units (ICU).

Methods: A retrospective cross-sectional study was conducted to explore the progression of antimicrobial resistance of Gram negative bacteria (GNB) in a tertiary care hospital in Riyadh, Saudi Arabia. All organisms were isolated from the adult ICU of King Abdulaziz Medical City between 2010 to 2014. Organisms were identified to the species level. Antimicrobial susceptibility testing was performed using an automated system (The VITEK® 2 system, BioMérieux, France) and the antimicrobial susceptibility testing was confirmed by E-Test.

Results: The total number of GNB was 7600. The most frequently isolated GNB were *Klebsiella pneumoniae*, *Acinetobacter baumannii*, and *Escherichia coli*. *Klebsiella* resistance was significantly increased to Cefepime and Ceftazidime. *Acinetobacter baumannii* demonstrated an increase in resistance towards Imipenem. The resistance pattern for *E.coli* seemed to be increasing to Cefazolin, Cefepime, and Ceftazidime.

Conclusion: A continuous surveillance program to observe the emergence of different bacterial resistance patterns is advised to establish unified guidelines across Saudi Arabia to reduce further progression in the emergence of MDR organisms.

I. INTRODUCTION

Antibiotic resistance is when bacteria develop the ability to resist the bactericidal or bacteriostatic effects of one or more antibiotic class (multidrug resistance (MDR)) (1). This resistance is most commonly noted in intensive care units (ICUs), which is due to the widespread use of antibiotics in these units compared to the other hospital departments (2). A study found that the incidence of ICU nosocomial infections worldwide was between 5%-30% (3). According to the national

healthcare safety network report in the United States (US); age, comorbid diseases, duration of hospitalization, length of ICU stay, immune status, and disease severity are all considered host risk factors for developing nosocomial infections in ICUs (4). In a study done on southern and eastern Mediterranean hospitals, overuse was one of the factors associated with increased antibiotic resistance (5). However, antibiotic resistance differs between ICUs in different countries due to various reasons including the different patterns of antibiotic use, the variation in infection control policies, and the effect of local resistance data in some countries directing the suitable antibiotic therapy which in turn leads to various outcomes on patients and healthcare systems (6). A previous study done in King Abdulaziz Medical City (KAMC), Riyadh Saudi Arabia from 2004-2009 including only Gram-negative bacteria (GNB) in the adult ICU, *Acinetobacter baumannii*, followed by *Pseudomonas aeruginosa*, *Escherichia coli* (*E.coli*), *Klebsiella pneumoniae*, *Stenotrophomonas maltophilia*, and *Enterobacter* were the most commonly isolated organisms (7). During the study period, the resistance of different common pathogens was increasing significantly. Globally, the efficacy of antibiotics against various ICU pathogens is decreasing over the past few years (7). Therefore, continuous surveillance studies should be conducted locally to observe the emergence of different bacterial resistance patterns, as there are clear differences between international and national data.

II. METHODOLOGY

A retrospective cross-sectional study was carried out of GNB from the adult ICU of King Abdulaziz Medical City (KAMC) between 2010 and 2014. The yearly antibiogram data obtained from the ICU department was used to seek the percentage of GNB resistance against specific antibiotics. The result of 7600 GNB isolates were interpreted according to the guidelines of the Clinical and Laboratory Standards Institute (CLSI). Gram-negative bacilli were identified to the species level and AST performed using an

Author ^α ^σ ^ρ ^ω [¥]: College of Medicine, King Saud Bin AbdulAziz University for Health Sciences, Riyadh, Saudi Arabia.
e-mail: roaa1414@hotmail.com

Author [§]: Department of Basic Medical Sciences, College of Medicine, King Saud bin Abdulaziz University for Health Sciences\King Abdullah International Medical Research Centre, Riyadh, Saudi Arabia.

Author ^x: Department of Pathology and Laboratory Medicine, King Abdulaziz Medical City, Riyadh, Saudi Arabia.

automated system (The VITEK® 2 system ,BioMerieux, France) and the antimicrobial susceptibility testing confirmed by E-Test (AB Biodisk). Only one isolate per patient per year was included in the analysis. The following antimicrobial agents were tested either by the breakpoint method (with the vitek 2 system) or by the ETEST method using the following antibiotics on (Muller Hinton Agar Plate): amikacin ampicillin ceftazidime ceftriaxone ciprofloxacin gentamicin imipenem trimethoprim- sulfamethoxazole Quality control was performed by testing these same antimicrobials on E.coli ATCC 25922, E coli ATCC 35218, P aeruginosa ATCC 27853, and Enterococcus faecalis ATCC 29212 to check the thymidine level on Muller Hinton Agar.

The proportion of susceptible isolates was calculated as the sum of susceptible organisms (neither intermediately susceptible nor resistant) relative to the total number of organisms tested. Multidrug resistance was defined as resistance to three or more antimicrobials (imipenem, ceftazidime, ciprofloxacin, piperacillin-tazobactam, and/or an aminoglycoside). The trend in the susceptibility rate over a 5-year period (between 2010-2014) was calculated and analyzed to identify a statistically significant increasing or decreasing trend using chi-square for linear trend analysis. Associations between categorical variables were tested using the chi-square test. The percent of change of antibiotic susceptibility was calculated as the difference between the later (e.g. 2014) and earlier (e.g. 2010) susceptibilities percentages divided by the earlier one. All P values were two-tailed. P value <0.05 was considered as significant. The data were analyzed using the Statistical Package for the Social Sciences, Version 20.0 (IBM Corporation, Armonk, NY, USA).

III. RESULTS

Throughout the study period (2010-2014), *Klebsiella* was the most commonly GNB in ICU (20.26%), and number of isolates in 2010 was 22.5% and 21.4% in 2014. *Klebsiella* resistance was significantly increased for Cefepime (81% to 89%; P-value= 0.001), and Ceftazidime (58% to 94%; P-value<.0001). In addition, *Klebsiella* resistance faced significant decrease in Ceftriaxone (67% to 43%; P-value<.0001), Carbapenems (meropenem 22% to 11%; P-value<.0001, and Imipenem 18% to 14%; P-value<.0001), Aminoglycosides (Amikacin 45% to 12%; P-value<.0001, and Gentamicin 50% to 27%; P-value<.0001), and Fluoroquinolone (Ciprofloxacin 70% to 38%; P-value<.0001).

Acinetobacter baumannii accounts for 17.97% of all GNB, and number of isolates were 17.04% in 2010 and 11.8% in 2014. *Acinetobacter baumannii* demonstrated increase in resistance toward Carbapenems (Imipenem 87% to 92%; P-value<.0001); however, resistance pattern seems to be decreasing in

Meropenem (97% to 92%; P-value= 0.473), Colistin (22% to 7%; P-value<.0001), and Amikacin (81% to 77%; P-value= 0.121).

E.coli was 9.6% of all GNB, and the number of isolates were 10.17% in 2010 and 9.32% in 2014. The resistance pattern seems to be increasing in beta-lactam antibiotics including Cefazolin (67% to 100%; P-value<.0001), Cefepime (48% to 100%; P-value<.0001), Ceftazidime (38% to 100%; P-value<.0001), and fluoroquinolone (Ciprofloxacin 65% to 70%; P-value= 0.271). On the other hand, *E. coli* resistance rate decreased for Piperacillin-tazobactam (36% to 27%; P-value= 0.276), and no resistance difference in imipenem and meropenem throughout the study period (0%; P-value=0.325).

Enterobacter isolates account for 4.5% of GNB, and number of isolates were 5.4% in 2010 and 4.3% in 2014. The resistance for some beta-lactam is increasing especially in Cefepime (47% to 69%; P-value=0.260), Ceftazidime (56% to 95%; P-value=0.002). Moreover, Carbapenems (meropenem 3% to 5%; P-value=0.670, and Imipenem 6% to 23%; P-value<.0001) showed slight increase in the resistance pattern against Enterobacter. Aminoglycosides (Amikacin 41% to 2%; P-value<.0001, and Gentamicin 31% to 8%; P-value<.0001), and fluoroquinolones (Ciprofloxacin 31% to 19%; P-value=0.016) showed decrease in resistance toward Enterobacter.

IV. DISCUSSION

Most of the hospital-acquired infections are related to invasive procedures and devices which are commonly seen in ICUs (8). The resistance pattern is most commonly noted in ICUs due to the widespread use of antibiotics in these units compared to the other hospital departments (2), and 70% of these infections were caused by GNB. (3). The increase in multidrug resistant organisms were shown to negatively affect the patient safety in which they can prolong the hospital stay, increase mortality rates, and health care costs (9).

This 5-year surveillance study is aimed to continue assessing the pattern of antibiotic resistance in GNB from adult ICU KAMC, Riyadh. As the annual antibiogram system were used in 2004 to 2009 to analyze the most common organisms and pattern of antibiotic resistance in our ICU. During the previous study period *Acinetobacter baumannii* revealed significant increase in resistance toward imipenem (45% to 90%), meropenem (67% to 90%), ciprofloxacin (78% to 90%), and amikacin (88% to 94%). *Pseudomonas aeruginosa* resistance markedly increased in 2007 specifically to carbapenems (34% to 74%), and ciprofloxacin (33% to 51%). *E.coli* showed significant increase in resistance to Cefuroxime (26% to 64%), ceftazidime (24% to 54%), cefotaxime (24% to 54%), cefepime (23% to 50%), and ampicillin (64% to 73%). S

marcescens showed increase in resistance toward cefotaxime (27% to 68%), ceftazidime (9% to 65%), and piperacillin-tazobactam (20% to 36%). Enterobacter resistance was markedly increased to ceftazidime (66% to 95%), cefotaxime (66% to 94%), and piperacillin-tazobactam (49% to 65%).

In our study (2010-2014) the most commonly isolated GNB were *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Escherichia coli*, and *Enterobacter*. In contrast, the previous surveillance (2004-2009), *Pseudomonas aeruginosa* and *Stenotrophomonas maltophilia* were considered as part of the most common GNB. Our data showed significant increase in resistance of *Klebsiella* toward beta-lactams antibiotics especially ceftazidime (58% to 94%), and significant decrease in resistance in meropenem (22% to 11%). Most of the isolated *Klebsiella* showed increased beta-lactamase activity, and the rate of Extended-spectrum beta-lactamases (ESBL) isolates increased from 12% in 2004 to 21.4% 2014. This increase might be due to implementation of new screening program in 2007. In the previous study, there was one case of carbapenem-resistant *klebsiella*. However, carbapenems are still considered very effective agent against *Klebsiella* and the resistance pattern seems to be decreasing during our study period (meropenem 22% to 11%, and Imipenem 18% to 14%). Despite that, carbapenemase resistant isolates should be taken into consideration due to their potential dissemination. The trend of the overall resistance pattern is illustrated in figure-1 and figure-2.

In addition, *Acinetobacter baumannii* resistance was significant toward imipenem (87% to 92%). For that, the resistance pattern seems to be progressing over the period of 2004-2014. Furthermore, meropenem showed a slight decrease in resistance (97% to 92%) that is not statistically significant. Colistin remains the most effective antibiotic against *Acinetobacter baumannii* and our study showed significant decrease in the resistance (22% to 7%). As the treatment options for carbapenem resistant *Acinetobacter baumannii* are limited and challenging, colistin might be used empirically in the setting of our ICUs. The trend of the overall resistance pattern is illustrated in figure 3 and figure 4.

Most of *E. coli* isolates exhibited ESBL activity, and resistance is significantly increased in all beta-lactams antibiotics especially ceftazidime (38% to 100%); while the previous surveillance study showed *E. coli* resistance to ceftazidime (24% to 54%). Piperacillin-tazobactam showed slight decrease in resistance (36% to 27%); however, this decrease is not statistically significant. All our ESBL-producing isolates were susceptible to carbapenems. There was no significant increase in the rate of *E. coli* ESBL from 2004 (9%) to 2014 (9.34%). The trend of the overall resistance pattern is illustrated in figure-5 and figure-6.

Enterobacter exhibited significant increase in resistance mostly toward ceftazidime (56% to 95%), and carbapenems showed unique increase in resistance to imipenem (6% to 23%). However, meropenem increase in resistance was not statistically significant. Aminoglycosides remain the most effective antibiotic against *Enterobacter* with amikacin being broadly active. The trend of the overall resistance pattern is illustrated in figure-7 and figure-8.

V. CONCLUSION

Our study concluded that Gram-negative bacterial resistance is still a major issue in KAMC, Riyadh adult. ICU. The most commonly isolated GNB were *Klebsiella pneumoniae* (20.26%), *Acinetobacter baumannii* (17.97%), *Escherichia coli* (9.6%), and *Enterobacter* (4.15%). Carbapenems is considered the most effective agent for *E. coli* and *Klebsiella* ESBL. Aminoglycosides is the most effective agent for *Enterobacter*, and Colistin is the drug of choice for most cases of *Acinetobacter baumannii*. This significant resistance observed in ICU is mostly due to the overuse of broad-spectrum antibiotics, prolonged patient stay, and variation in infection control policies. Thus, the importance of collaboration between the ICU, infection control, infectious disease departments is very essential to substantially decrease the resistance rates. Furthermore, establishment of local database of antibiogram across the whole kingdom of Saudi Arabia will aid in the improvement of treatment strategies and guidelines based on unit-specific data.

REFERENCES RÉFÉRENCES REFERENCIAS

1. Cdc.gov. About Antimicrobial Resistance| Antibiotic/Antimicrobial Resistance | CDC [Internet]. 2015 [cited 1 December 2015]. Available from: <http://www.cdc.gov/drugresistance/about.html>
2. Radji M, Fauziah S, Aribinuko N. Antibiotic sensitivity pattern of bacterial pathogens in the intensive care unit of Fatmawati Hospital, Indonesia. *Asian Pacific Journal of Tropical Biomedicine*. 2011; 1(1): 39-42.
3. Hospital-Acquired Infections Due to Gram-Negative Bacteria". *New England Journal of Medicine* 363. 15 (2010): 1482-1484. Web.
4. Doyle JS e. Epidemiology of infections acquired in intensive care units. - PubMed - NCBI [Internet]. Ncbi.nlm.nih.gov. 2015 [cited 1 December 2015]. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/2150604>
5. Borg MA e. Antibiotic consumption in southern and eastern Mediterranean hospitals: results from the ARMED project. - PubMed - NCBI [Internet]. Ncbi.nlm.nih.gov. 2015 [cited 1 December 2015]. Available from: <http://www.ncbi.nlm.nih.gov/pubmed/18593724>

6. Lee G, Reveles K, Attridge R, Lawson K, Mansi I, Lewis J et al. Outpatient antibiotic prescribing in the United States: 2000 to 2010. *BMC Medicine*. 2014; 12(1): 96.
7. Al Johani S, Akhter J, Balkhy H, El-Saed A, Younan M, Memish Z. Prevalence of antimicrobial resistance among gram-negative isolates in an adult intensive care unit at a tertiary care center in Saudi Arabia. *Annals of Saudi Medicine*. 2010; 0(0): 0.
8. Dasgupta, Sugata et al. "Nosocomial Infections In The Intensive Care Unit: Incidence, Risk Factors, Outcome And Associated Pathogens In A Public Tertiary Teaching Hospital Of Eastern India". *Indian Journal of Critical Care Medicine* 19.1 (2015): 14. Web.
9. Slama, Thomas G. "Gram-Negative Antibiotic Resistance: There Is A Price To Pay". *Critical Care* 12. Suppl 4 (2008): S4. Web.

APPENDIX

Appendix 1:

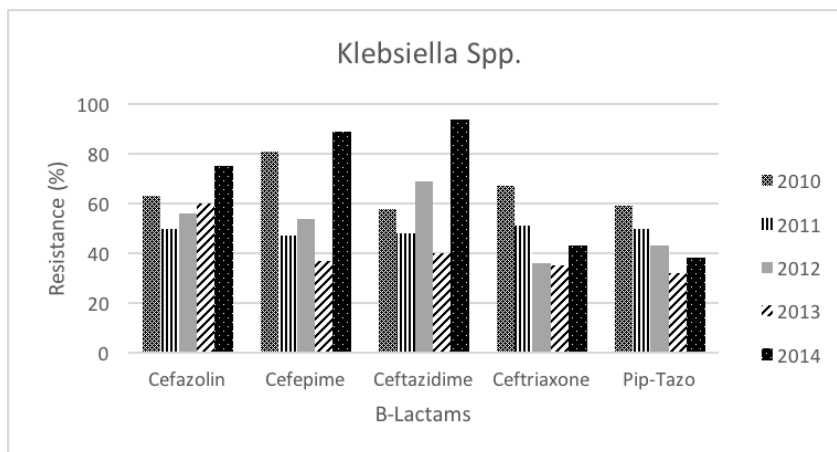


Figure 1: Beta-lactams Antibiotic Resistance for Klebsiella Spp. Isolates Tested Between 2010-2014.

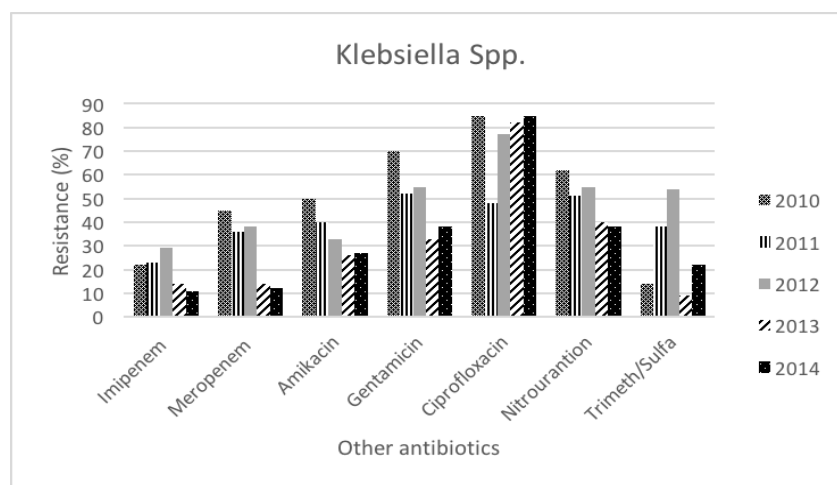


Figure 2: Carbapenems, Aminoglycosides, Fluoroquinolones, Clostin, Nitroulantion, Trimethoprim/Sulfamethaxole Antibiotic resistance for Klebsiella Spp. Isolates Tested Between 2010-2014.

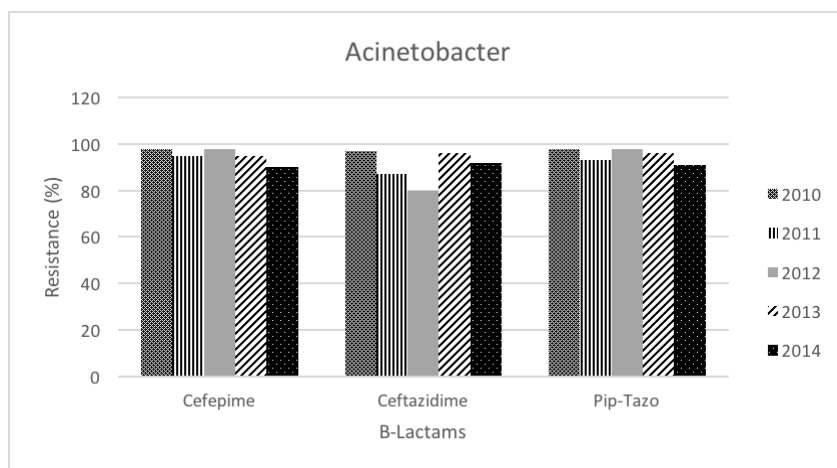


Figure 3: Beta-lactams Antibiotic Resistance for Acinetobacter Isolates Tested Between 2010-2014.

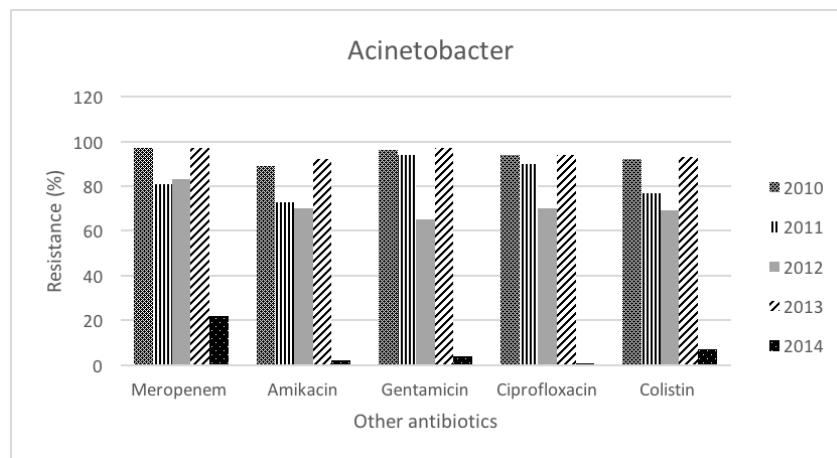


Figure 4: Carbapenems, Aminoglycosides, Fluoroquinolones, and Clostin Antibiotic resistance for Acinetobacter Isolates Tested Between 2010-2014.

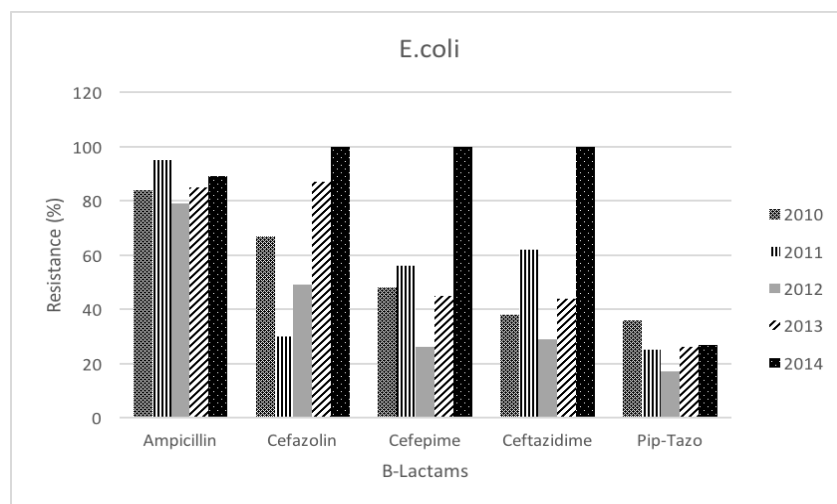


Figure 5: Beta-lactams Antibiotic Resistance for E.coli Isolates Tested Between 2010-2014.

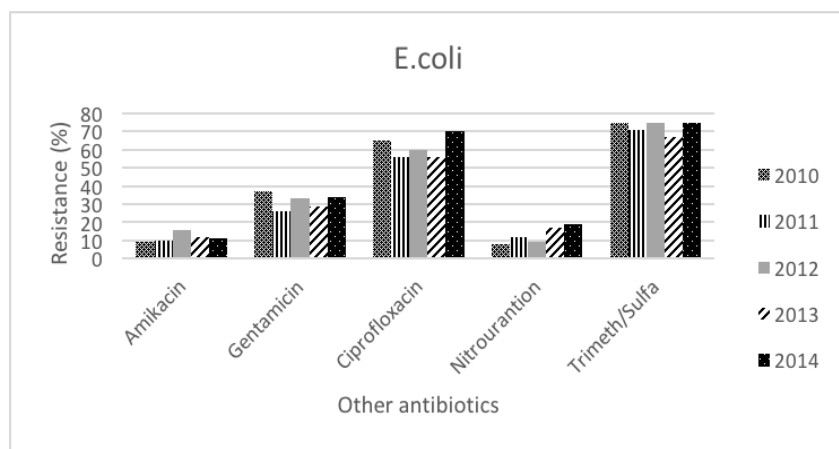


Figure 6: Aminoglycosides, Fluoroquinolones, Nitrofurantoin, and Trimethoprim/Sulfamethoxazole Antibiotic resistance for E. coli Isolates Tested Between 2010-2014.

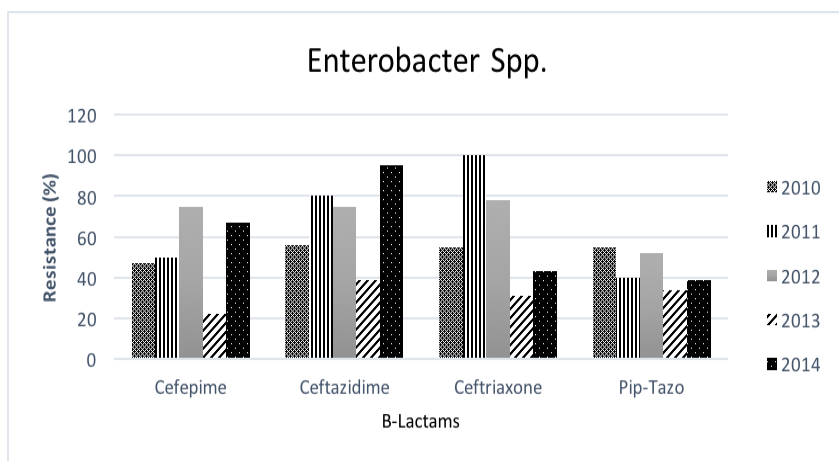


Figure 7: Beta-lactams Antibiotic Resistance for Enterobacter Spp. Isolates Tested Between 2010-2014.

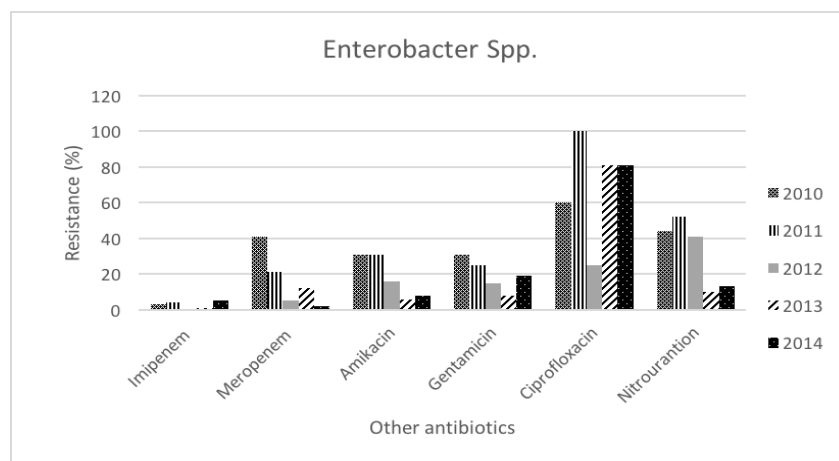


Figure 8: Carbapenems, Aminoglycosides, Fluoroquinolones, Nitrofurantoin, and Trimethoprim/Sulfamethoxazole Antibiotic resistance for Enterobacter Spp. Isolates Tested Between 2010-2014.

Appendix 2:

Table 1: Comparison between 2010 and 2014 antibiotic resistance of Klebsiella Spp.:

Antibiotic	Resistance (%) in 2010	Resistance (%) in 2014	P-value	Trend
Beta-Lactam Antibiotics:				
Cefazolin	67%	100%	<.0001	↑
Cefepime	48%	100%	<.0001	↑
Ceftazidime	38%	100%	<.0001	↑
Ceftriaxone	45%	59%	<.0001	↑
Pip-Tazo	36%	27%	<.0001	↑
Other Antibiotic Groups:				
Imipenem	18%	14%	<.0001	↓
Meropenem	22%	11%	<.0001	↓
Amikacin	45%	12%	<.0001	↓
Gentamicin	50%	27%	<.0001	↓
Ciprofloxacin	70%	38%	<.0001	↓
Nitrofurantoin	85%	85%	<.0001	—
Trimeth/Sulfa	62%	38%	<.0001	↓

Table 2: Comparison between 2010 and 2014 antibiotic resistance of Acinetobacter baumannii:

Antibiotic	Resistance (%) in 2010	Resistance (%) in 2014	P-value	Trend
Beta-Lactam Antibiotics:				
Cefepime	98%	90%	0.001	↓
Ceftazidime	97%	92%	0.298	↓
Pip-Tazo	98%	91%	0.026	↓
Other Antibiotic Groups:				
Imipenem	87%	92%	<.0001	↑
Meropenem	97%	92%	0.473	↓
Amikacin	81%	77%	0.121	↓
Gentamicin	81%	69%	0.010	↓
Ciprofloxacin	97%	93%	0.232	↓
Colistin	22%	7%	<.0001	↓

Table 3: Comparison between 2010 and 2014 antibiotic resistance of E.coli:

Antibiotic	Resistance (%) 2010	Resistance (%) 2014	P-Value	Trend
Beta- Lactams antibiotics				
Cefazolin	67	100	<0.0001	↑
Cefepime	48	100	<0.0001	↑
Ceftazidime	38	100	<0.0001	↑
Pip-Tazo	36	27	0.276	↓
Others antibiotics				
Amikacin	9	11	0.617	↑
Gentamicin	37	34	0.908	↓
Ciprofloxacin	65	70	0.271	↑
Nitroulantion	8	19	0.002	↑
Trimeth/Sulfa	75	75	0.809	—

Table 4: Comparison between 2010 and 2014 antibiotic resistance of Enterobacter:

Antibiotic	Resistance (%) 2010	Resistance (%) 2014	P-Value	Trend
Beta- lactams antibiotics				
Cefepime	47	67	0.260	↑
Ceftazidime	56	95	0.002	↑
Ceftriaxone	55	43	<0.0001	↓
Pip-Tazo	55	39	0.047	↓
Others antibiotics				
Amikacin	41	2	<0.0001	↓
Gentamicin	31	8	<0.0001	↓
Ciprofloxacin	31	19	0.016	↓
Nitroulantion	60	81	0.064	↑
Imipenem	0	23	<0.0001	↑
Meropenem	3	5	0.670	↑
Trimeth/Sulfa	44	13	<0.0001	↓