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Use of Dental Stem Cells Harvested from a Tooth or Follicle

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Abstract- Regenerative medicine is gaining popularity and inspires dentists to follow evidence-based literature for educating the parents about collecting, storing, sustainability and utility of dental stem cells about autologous regenerative therapies. Stem cells are present intrinsically in the apical papilla or dental pulp cells for treatment associated to regenerative endodontics.^{1, 2, 3}

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Abstract- Regenerative medicine is gaining popularity and inspires dentists to follow evidence-based literature for educating the parents about collecting, storing, sustainability and utility of dental stem cells about autologous regenerative therapies. Stem cells are present intrinsically in the apical papilla or dental pulp cells for treatment associated to regenerative endodontics.^{1,2,3}

INTRODUCTION

Stem cells are pluripotent cells which can divide and increase for a protracted period and differentiate into specialized cell types and tissues of a wide variety. Dental stem cells are a subset of adult mesenchymal stem cells which proliferate rapidly and differentiate into many cell lines.⁴

Adult stem cell therapy applies to medical presentations as metabolic disorders, bone marrow transplant for hematopoietic cancers and congenital immunodeficiency syndromes. It is undergoing clinical testing for diabetes, brain trauma/ spinal cord injuries and Parkinson's disease among other conditions.^{5,6}

The oral health applications of stem cells include dental and periodontal tissue regeneration, craniofacial structure repair (cleft lip/palate) and wound healing.⁷ They are derived from various sources such as preserved umbilical cord blood, pulp tissue of exfoliating primary teeth, fibroblasts from oral mucosa,⁸ periodontal ligaments, surgically removed third molars⁹ and gingival fibroblasts.^{2, 9, 10} They are secured and stored for conserving their proliferation and differentiation potential.^{11,12} Since these dental stem cells are not very stable and lead to in vivo tumours,² more studies are required to assess their safety and efficacy prior to initiating clinical trials in humans.^{11,12}

Modern regenerative endodontics is augmented by newer techniques including tissue engineering and regenerative medicine.¹³ Stem cell transplantation and cell homing strategies are applicable to pulp regeneration. Stem cells are isolated, expanded, seeded and transplanted into the scaffold. Cell homing achieve tissue repair/regeneration by replacing the necrotic, inflamed or injured pulpal tissue with a regenerated pulp-like tissue by recruiting endogenous cells via signaling molecules. Compared with stem cell transplantation, cell homing strategies do not need to isolate and manipulate stem cells in vitro.^{14, 15, 16}

An example of cell homing strategy is pulp revascularization of immature teeth. Histologically most of the tissues formed in such cases are unlike pulp like tissue and comprise of cementum, periodontal, and bone-like tissues.¹⁷ Additional studies are required to increase the success rate of pulp revascularization in immature teeth, facilitate the formation of pulp-like tissues and applying cell homing in mature teeth.

The first prerequisite for promoting pulpal regeneration include disinfection of the pulp space and dentinal walls via different root canal disinfectants including intracanal medication with antibiotics, ultrasonic-assisted irrigation, EndoVac apical negative-pressure system of irrigation and laser irradiation.¹⁸

Another precondition for pulp regeneration is the proper size of the apical foramen, especially in mature teeth with closed apex in adults. An apical foramen too small in size will affect both the migration of endogenous cells as well as neovascularization and re-innervation during regeneration. A minimum of 1.1 mm apical foramen is necessary to obtain revascularization.¹⁹

Morphologically the regenerated tissues should encompass connective tissues for new dentin formation at a controlled rate similar to healthy pulp, display cell density, and architecture as the natural tissue, and have innervation and vascularization.²⁰ Vascularization and innervation are vital pulpal characteristics hence the new regenerated blood vessels must connect with the periapical or bone marrow tissues around the teeth to receive a regular blood flow from circulation, nutrient supply and sense hot/cold stimulation and pain during infection.²¹

An area of concern for homing cell strategy is the source of stem. They are derived from dental pulp stem cells (DPSCs), apical papilla (SCAP), bone marrow stem cells (BMSCs), periodontal ligament cells migrating to the root canal by chemokines and others.²² To conclude; even though some experimental and clinical studies yield pulp-like vascularized tissue; the information about the function of this tissue is insufficient. Hence further studies are required to succeed in functional pulp regeneration.

REFERENCES RÉFÉRENCES REFERENCIAS

1. Conde MC, Chisini LA, Demarco FF, et al. Stem cell based pulp tissue engineering: Variables enrolled in translation from the bench to the bedside, a

- systematic review of literature. *Int Endod J* 2016; 49(6):543-50.
2. Yang J, Yuan G, Chen Z. Pulp regeneration: Current approaches and future challenges. *Front Physiol* 2016; 7:58.
3. Hynes K, Menichanin D, Bright R, et al. Induced pluripotent stem cells: A new frontier for stem cells in dentistry. *J Dent Res* 2015; 94(11):1508-15.
4. Govindasamy V, Ronald VS, Abdullah AN, et al. Differentiation of dental pulp stem cells into islet-like aggregates. *J Dent Res* 2011; 90(5):626-52.
5. Nourbakhsh N, Soleimani M, Taghipour Z, et al. Induced in vitro differentiation of neural-like cells from human exfoliated deciduous teeth-derived stem cells. *Int J Dev Biol* 2011; 55(2):189-95.
6. Kadar K, Kiraly M, Porcsalmy B, et al. Differentiation potential of stem cells from human dental origin – Promise for tissue engineering. *J Physiol Pharmacol* 2009; 60(suppl 7):167-75.
7. Nishino Y, Yamada Y, Ebisawa K, et al. Stem cells from human exfoliated deciduous teeth (SHED) enhance wound healing and the possibility of novel cell therapy. *Cytotherapy* 2011; 13(5):598-605.
8. Miyoshi K, Tsuji D, Kudoh K, et al. Generation of human induced pluripotent stem cells from oral mucosa. *J Biosci Bioeng* 2010; 110(3):345-50.
9. Wada N, Wang B, Lin NH, Laslett AL, et al. Induced pluripotent stem cell lines derived from human gingival fibroblasts and periodontal ligament fibroblasts. *J Periodontal Res* 2011; 46(4): 438-47.
10. Eslaminejad MB, Vahabi S, Shariati M, Nazarian H. In vitro growth and characterization of stem cells from human dental pulp of deciduous versus permanent teeth. *J Dent (Tehran)* 010; 7(4):185-95.
11. Perry BC, Zhou D, Wu X, et al. Collection, cryopreservation, and characterization of human dental pulp-derived mesenchymal stem cells for banking and clinical use. *Tissue Eng Part C Methods* 2008; 14(2):149-56.
12. Yildirim S, Zibandeh N, Genc D, Ozcan EM, Goker K, Akkoc T. The comparison of the immunologic properties of stem cells isolated from human exfoliated deciduous teeth, dental pulp, and dental follicles. *Stem Cells Int* 2016; 2016:4682875. Available at: "<https://www.hindawi.com/journals/sci/2016/4682875/>". Accessed June 9, 2017. (Archived by WebCite® at: "<http://www.webcitation.org/6tkJoR5zd>").
13. Nakashima, M., and Iohara, K. (2011). Regeneration of dental pulp by stem cells. *Adv. Dent. Res.* 23, 313–319. doi: 10.1177/0022034511405323.
14. Sun, H. H., Chen, B., Zhu, Q. L., Kong, H., Li, Q. H., Gao, L. N., et al. (2014). Investigation of dental pulp stem cells isolated from discarded human teeth extracted due to aggressive periodontitis. *Biomaterials* 35, 9459–9472. doi: 10.1016/j.biomaterials.2014.08.003.
15. Yang, J. W., Zhang, Y. F., Wan, C. Y., Sun, Z. Y., Nie, S., Jian, S. J., et al. (2015a). Autophagy in SDF-1 α -mediated DPSC migration and pulp regeneration. *Biomaterials* 44, 11–23. doi: 10.1016/j.biomaterials.2014.12.006.
16. Yang, J. W., Zhang, Y. F., Sun, Z. Y., Song, G. T., and Chen, Z. (2015b). Dental pulp tissue engineering with bFGF-incorporated silk fibroin scaffolds. *J. Biomater. Appl.* 30, 221–229. doi: 10.1177/0885328215577296.
17. Thibodeau, B., and Trope, M. (2007). Pulp revascularization of a necrotic infected immature permanent tooth: case report and review of the literature. *Pediatr. Dent.* 29, 47–50.
18. Fouad, A. F. (2011). The microbial challenge to pulp regeneration. *Adv. Dent. Res.* 23, 285–289. doi: 10.1177/0022034511405388.
19. Kling, M., Cvek, M., and Mejare, I. (1986). Rate and predictability of pulp revascularization in therapeutically reimplanted permanent incisors. *Endod. Dent. Traumatol.* 2, 83–89. doi: 10.1111/j.1600-9657.1986.tb00132.x.
20. Fawzy El-Sayed, K. M., Jakusz, K., Jochens, A., Dörfer, C., and Schwendicke, F. (2015). Stem Cell transplantation for pulpal regeneration: a systematic review. *Tissue Eng. Part B Rev.* 21, 451–460. doi: 10.1089/ten.teb.2014.0675.
21. Kökten, T., Bécavin, T., Keller, L., Weickert, J. L., Kuchler-Bopp, S., and Lesot, H. (2014). Immunomodulation stimulates the innervation of engineered tooth organ. *PLoS ONE* 9:e86011. doi: 10.1371/journal.pone.0086011.
22. Zhou, J., Shi, S., Shi, Y., Xie, H., Chen, L., He, Y., et al. (2011). Role of bone marrow-derived progenitor cells in the maintenance and regeneration of dental mesenchymal tissues. *J. Cell. Physiol.* 226, 2081–2090. doi: 10.1002/jcp.22538.
23. Shi, S., Bartold, P. M., Miura, M., Seo, B. M., Robey, P. G., and Gronthos, S. (2005). The efficacy of mesenchymal stem cells to regenerate and repair dental structures. *Orthod. Craniofac. Res.* 8, 191–199. doi: 10.1111/j.1601-6343.2005.00331.x