Neuromodulation of Mu-opioid Receptor (MOR-1) Gene (OPRM1) Alternatively-Spliced Variants Following Exposure to Morphine with Alma Fig (Ficus carica) Leaf Extract in Human Neuroblastoma (SH-SY5Y) Cells: Review & Pilot Study Carl B. Goodman Received: 6 December 2019 Accepted: 5 January 2020 Published: 15 January 2020

8 Abstract

The role of morphine in regulating the mu-opioid receptor (MOR-1) relative to pain is g well-established. Efforts are ongoing to elucidate the pharmacological significance of newly 10 identified MOR-1 alternative splice variants. Aberrant splicing events have been implicated in 11 a growing number of diseases, including cancer, but it is uncertain whether any 12 pharmacological benefit may be derived from the use of these variants. Chronic use of opioids 13 yields tolerance, withdrawal, and potentially fatal addiction. With current interests so high on 14 developing marijuana as a marketable drug, there is concern whether its introduction as a 15 mainstay may interfere with pain medications, such as opioids, for which there is a growing 16 concern of epidemic proportions. We, therefore, hypothesized that the introduction of 17 traditional herbal medicines while taking morphine would interfere with normal pain receptor 18 functions. We tested this hypothesis by chronically (48hrs) exposing human neuroblastoma 19 (SH-SY5Y) cells to a pain medication (morphine) followed by a natural herb, and measuring 20 its effect on the expression of MOR-1 alternatively-spliced variants. (RA)-differentiated 21 human neuroblastoma (SH-SY5Y) cells treated with morphine (10 ?M), fig leaf extract (3 22 2L/30 mL media), or both for 48 hours, were analyzed by quantitative real-time polymerase 23 chain reaction (qRT-PCR) using the Bio-Rad iCycler/MyiQ?. Of the seven fig (Ficus carica 24 L.) cultivars (Green Ishia, Brown Turkey, Mission, Alma, Giant Celeste, Nero, Hollier) 25 identified for this pilot study, Alma fig leaf extract was selected for combined therapy with 26 morphine. Statistically significant differential regulation of MOR-1 alternative splice variants 27 was widely observed in control, morphine, Alma fig leaf extract, and morphine/Alma fig 28 samples. The results of this pilot study confirm our hypothesis that MOR-1 splice variants are 29 differentially regulated following chronic exposure to morphine and Ficus carica. Further 30 examination of the relationship between morphine and herbs used in traditional medicine may 31 enhance our understanding of the mechanistic basis of morphine tolerance and may give clues 32 concerning the therapeutic benefit of using Ficus carica Author ??? : Basic Pharmaceutical 33 Sciences Division, Florida AM University, College of Pharmacy Pharmaceutical Sciences, 34 Tallahassee, FL, USA. e-mail: lightbournal@vahoo.com leaf extracts to counteract the effects 35 of opioids via targeted posttranscriptional isoforms of the mu-opioid receptor. (333 words) 36

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Index terms—Ficus carica; G-protein; mu-opioid receptor; opioid; morphine; tolerance; alternative splicing. Abstract-The role of morphine in regulating the mu-opioid receptor (MOR-1) relative to pain is wellestablished. Efforts are ongoing to elucidate the pharmacological significance of newly identified MOR-1

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alternative splice variants. Aberrant splicing events have been implicated in a growing number of diseases, 41 including cancer, but it is uncertain whether any pharmacological benefit may be derived from the use of these 42 variants. Chronic use of opioids yields tolerance, withdrawal, and potentially fatal addiction. With current 43 interests so high on developing marijuana as a marketable drug, there is concern whether its introduction 44 45 as a mainstay may interfere with pain medications, such as opioids, for which there is a growing concern of epidemic proportions. We, therefore, hypothesized that the introduction of traditional herbal medicines while 46 taking morphine would interfere with normal pain receptor functions. We tested this hypothesis by chronically 47 (48hrs) exposing human neuroblastoma (SH-SY5Y) cells to a pain medication (morphine) followed by a natural 48 herb, and measuring its effect on the expression of MOR-1 alternatively-spliced variants. (RA)-differentiated 49 human neuroblastoma (SH-SY5Y) cells treated with morphine ?? The results of this pilot study confirm our 50 hypothesis that MOR-1 splice variants are differentially regulated following chronic exposure to morphine 51 and Ficus carica. Further examination of the relationship between morphine and herbs used in traditional 52 medicine may enhance our understanding of the mechanistic basis of morphine tolerance and may give clues 53 concerning the therapeutic benefit of using Ficus carica I. Introduction he human nervous system is composed of 54 a complex and highly organized network of excitable tissues, neurons, and their receptors, effectors, interneurons, 55 neurotransmitters, hormones, and a host of structures through which to orchestrate anatomic homeostasis, in 56 57 tandem with the endocrine system. The neuron is the central functional unit of the nervous system tasked 58 with synchronizing action potentials that govern sensory, integrative, and motor functions. This small but 59 rapid and highly efficient communication system regulates processes of learning and memory, sensations (e.g., pain, thermal, tactile, proprioceptive), perception, analgesia, differentiation, development, emotional responses, 60 emotional behaviors, wakefulness and sleep (Tortora & Grabowski, 2003;Massirer et al., 2010). An extensive 61 annual review (not elaborated here) covering the endogenous opioid system reflects its diverse contributions 62 to matters concerning: "behavior, pain, and analgesia; stress and social status; tolerance and dependence; 63 learning and memory; eating and drinking; alcohol and drugs of abuse; sexual activity and hormones; pregnancy; 64 development and endocrinology; mental illness and mood; seizures and neurologic disorders; electrical-related 65 activity, neurophysiology and transmitter release; general activity and locomotion; gastrointestinal, renal, and 66 hepatic function; cardiovascular responses; respiration and thermoregulation; ??and] immunological responses" 67 (Bodnar & Klein, 2006). The activities of each body system are regulated through action potentials generated 68 by the neuron. Extensive alternative splicing in the nervous system is, therefore, likely to play a role in many 69 70 of these physiological processes and conditions (Grabowski & Black, 2001). The endogenous opioid system, 71 which is resident within the mammalian nervous system, plays a significant role in a variety of physiological processes within the mammalian body. The nervous system is naturally resilient. It does not easily succumb to 72 toxicity or insult, instituting neuroadaptive changes and selfrecovery instead. Importantly, the nervous system 73 has a unique integrative capacity to resolve at the molecular level problems that arise at the cellular level. 74 Psychotropic substances of plant origin, such as morphine from the opium poppy (Papaver somniferum), mimic 75 the action of neurotransmitter action of enkephalins, the natural ligand for this receptor. Specialized (sensory) 76 neurons throughout the body mediate pain sensation by regulating the human muopioid receptor (MOR-1) gene 77 (OPRM1) expression. Among natural medicines used for pain, the fig (Ficus carica) plant is commonly not 78 listed. It more often finds prominence relative to conditions such as diabetes, hyperlipidemia, eczema, psoriasis, 79 constipation, skin tumors and warts, and vitiligo ??Jellin et al., 2009), some of which are side effects of opioids 80 (Stephan & Parsa, 2016). Hence, it would be interesting to learn of a role for the fig plant in the mediation of 81 pain, analgesia, and opioid receptor pharmacology. 82

⁸³ 1 Consumption of Pain Medications

Pain drugs are the second most dominant pharmaceutical class in the global market. US market. For
centuries, the alkaloid-derived morphine (Figure 1) has remained the prototypical anti-nociceptive agent ??WHO,
1986;Pasternak, 2001; ??anquelin & von Mentzer, 2007;Yu & Sadee, 1988;Corbett et al., 2006). Its analgesic
superiority underscores the use of morphine as a preferred clinical and non-medical psychotherapeutic drug
(Tremblay & Hamet, 2010). Contrary to controversial reports that the United States alone utilized eighty
percent (80%) of the global supply of morphine (Manchikanti et al., 2006(Manchikanti et al., 2010)

⁹⁰ 2 Cause for Concern: Variability of Response to Drug-Disease ⁹¹ Interactions

The observation that there exist inter-and intraindividual differences in response to prescribed or illicitly used medications reinforces the significance of modernday precision medicine (Samer et al., 2006;Rollason et al., 2008;Dorn & Cresci, 2008). Characteristically, differences in age as well as in drug interactions with cytochrome P450 metabolic enzymes have historically separated subpopulations from generalized use of medications to more patient-centered determinations of appropriate pharmacological treatments (Samer et al., 2006;Rollason et al., 2008; ??inklestein, 2017;Krebs & Milani, 2019). The ongoing discussion of genetic polymorphisms continues to inform this process.

⁹⁹ The current literature on alternative splicing (Figure 2) indicates that this posttranscriptional process is ¹⁰⁰ essential for life but may to contribute inter-and intraindividual variability by altering gene function (House

& Lynch, 2008); switching substrate specificity (Christmas et al., 2001; Bauman et al., 2009); or causing 101 disease (e.g., cancer) through aberrant splicing events (Faustino 2003;Buratti et al., 2006). At least ten 102 alternativelyspliced isoforms (Figure 3) of the human mu-opioid receptor (MOR-1) gene (OPRM1) have been 103 identified ??Pasternak and Pan, 2009). Moreover, each splice variant may exhibit different agonist-induced 104 activation, signal transduction, and protein expression patterns. Within pharmacogenomics, understanding how 105 a person's genetic profile influences his response to a drug is a treasured clinical endeavor, in which is embedded 106 great hope for the improvement in the medical use and administration of drugs across all ages and stages of 107 development ??Finklestein, 2017). Central to these efforts is the drug-receptor (Danhof et al., 2007;Ploeger et 108 al., 2009). 109

110 3 Historical Receptor Theories

The receptor is the smallest pharmacological unit necessary to differentiate between drugs (Kenakin, 2004). The 111 idea that receptors are responsible for drug effects is an evolving theory that developed between the late 19 th 112 century and early 20 th century due to the pioneering work of several scientists. Credited for the concept of 113 "locus of effect," Claude Bernard (1813-1878) pioneered a methodological blueprint for elucidating the specificity 114 and selectivity of drug action (Bernard, 1856). As an offshoot of interest in finding a more rational approach 115 for therapy, Hungarian scientist Rudolf Buchheim (1820-1879) opened the first pharmacology laboratory with 116 the intent of measuring drug effects and their associated mechanisms of action (Hollinger, 1997). In1848, Blake 117 framed the structure activity relationship (SAR). He made observations to correlate the biological effects of a 118 substance with its chemical structure, arguing that a specific component was responsible for the observed change 119 rather than the complex as a whole. He garnered theoretical support from the later work of Arrhenius on 120 electrolytic dissociation, and Crum Brown and Fraser who found differing physiological actions by alternating 121 alkaloid structures. Hans Horst Meyer (1899) and Charles Ernest Overton (1901), independently described lipid 122 123 solubility. Among other discoveries during this period, other scientists were discovering the high physiological specificity of opioids on smooth muscle versus smooth muscles. 124

The existence of receptors was first suggested in 1878 by John Newport Langley (1852-1925), followed in 1905 by him coining the term "receptive substances." Paul Ehrlich (1854-1915) specifically introduced the concept word "receptor" in his medical correspondence wherein he attributed a therapeutic effect only to an agent having "the right sort of affinity" ??Ehrlich, 1923;Hollinger, 1997). Ehrlich envisioned receptors as "sidechains" that interacted with a "combining group of the protoplasmic molecule to which the introduced group is anchored" in mammalian cells (Hollinger, 1997).

As he studied the interactions between enzymes and substrates, Emil Fischer, a German chemist, and enzymologist, was the first to propose a "lock and key" relationship between a drug and its receptor. Fischer postulated that a specific similar geometric configuration of the receptor was necessary for a chemical reaction to proceed from contact between these molecules. The precise fit was required to produce the optimal response (Hollinger, 1997). This theory was consistent with existing science showing that the primary amino acid sequence of a protein determines its three-dimensional structure, and according to Christian Anfinsen, these molecules were capable of unfolding (denaturing) and folding (renaturing) to vary their conformation (Hollinger, 1997).

¹³⁸ 4 Characteristics of the Mu-opioid Receptor

The opioid receptor is a member of Class A of the superfamily of guanosine nucleotide-binding protein (G-protein)-139 coupled receptors (GPCRs) that constitute $\sim 3\%$ of the human genome. They contain a total of 7 extracellular 140 and intracellular transmembrane (7TM) domains linked to three subunits: alpha?, beta?, and gamma?. The beta 141 and gamma subunits are tightly linked, while the alpha subunit more freely associates or dissociates from this 142 dimer. Ligands approach and engage the receptor from the extracellular space, and receptor activation results 143 in coupling to heterotrimeric G-proteins on the intracellular face of the membrane. Binding and hydrolysis of 144 guanosine triphosphate (GTP) to the ?-subunit of the G-protein activates a resting receptor and results in the 145 dissociation of the ?? subunit from the receptor. The ?? subunit in conjunction with downstream effectors or the 146 GTP-bound ?-subunit can trigger a plethora of downstream events. The association of guanosine diphosphate 147 (GDP) with the ?-subunit promotes its further association with the ?? subunits, returning it to an inactive state. 148 Opioid receptor signals are transduced by intracellular inhibitory G-proteins (G i /G 0), which are relatively 149 resistant to tolerance and desensitization (Pasternak, 2001;Pan et al., 1999Pan et al., 2001Pan et al., 2005)). 150 As Alfred Joseph Clark (1885-1941) first proposed, the "receptor occupancy theory" demonstrates the 151 interaction of a first messenger (e.g., a signal molecule such as a drug, chemical, or neurotransmitter) with 152 its specific physiological cellular receptor (e.g., mu-opioid receptor, subtype 1 -MOR-1) to produce a measurable 153 154 biological response (Limbird, 2005). The curve of a dose-response graph resembles a mathematical hyperbole. 155 Subsequent research by Raymond P. ?? hlquist (1914 ?? hlquist (-1983)) led to the discovery of unique differences between alpha-and betaadrenoceptors, and this served one catalyst for Sir James Black's (1988) Nobel winning 156 interrogation of drugs with receptor-selective subtypes. Not too long afterward, Gilman and Rodbell won the 157 Nobel Prize for GPCRs and receptor coupling. These monumental works have moved the field of receptor 158 pharmacology to uncharted heights that continue to influence today's society. A summary of the general 159 characteristics of receptors appears in Table 3. 160

In studies conducted by Kuhar (2010), autoradiographic localization of opiate receptors rendered these microscopic molecules as being saturable, primarily particulate-bound, and accessible in proportion to the high level of activity of opiate drugs, but seemingly unaffected by drugs not of opiate origin. Observed drug effects are a consequence of physiological responses associated with control mechanisms that permit access to the drug via the action of its physiological intermediate.

When an endogenous opioid, such as enkephalin, binds to and activates MOR-1, the ensuing conformational changes help to modulate synaptic transmission in the neuron, ultimately resulting in a cascade of intracellular signaling events that amplify the signal and produce a diverse array of pharmacological outcomes, depending on the tissue (Limbird, 2005; ??enye et al., 2015). The intensity of the response to a signaling 'messenger' molecule (ligand) depends in large part on the specificity with which that ligand attaches to the receptor recognition site (binding pocket).

¹⁷² 5 MOR-1 Alternative Splicing

Although the pharmacological and physiological attributes of morphine and its receptors have been extensively elaborated over the past three decades ??Zadina et al., 1993;Pasternak, 2001 (Pasternak, 2001). Gene expression is regulated at the transcriptional level; hence, contributions by MOR-1 splice variants are of interest. There is also a dearth of information about mechanisms that account for the substantial diminution of the efficacy of morphine, which gives rise to the development of tolerance following longterm use (Yu & Sadee, 1988; ??adina et al., 1993;Taylor & Fleming, 2001;Willner et al., 2014).

Up to sixty percent of the human genome is estimated to contain alternatively-spliced gene isoforms (Lee & 179 Irizarry, 2003). Aberrant splicing events have been implicated in a growing number of diseases, including cancer 180 (Mercadante & Kole, 2000;Braaco & Kearney, 2003;Lee & Irizarry, 2003;Brinkman, 2004). The C-terminus of 181 cell-surface, seven-transmembranes (7TM) receptors is home to the biggest array of splice variants, which may 182 occur at more than one site on the receptor, adding to the complex structure of the gene (Kilpatrick et al., 183 1999). The opioid receptor is one example of a 7TM receptor within the guanine nucleotide-binding proteins (g-184 protein)-coupled receptor (GPCR) family, which transfers signals for hundreds of cellular receptors. This highly 185 diversified family of gprotein receptors execute neurotransmission, cellular differentiation, hormonal activities, 186 signal transduction, metabolism, and other processes (Kilpatrick et al., 1999). The pharmacological significance 187 of newly identified mu-opioid receptor (MOR-1) alternative splice variants (Pasternak & Pan, 2004;House & 188 Lynch, 2008) has not been characterized relative to drug response mechanisms and may inform the issue of 189 morphine tolerance. Among the 70-90% of cancer patients requiring individualized opioid therapy for intense 190 chronic pain, the response to prototypical opiates like morphine is highly variable, necessitating dose escalation 191 with an increased risk of developing tolerance ??WHO, 1986; ??racco and Kearsey, 2003). 192

Given the central role of MOR-1 in pain mediation, brain reward systems, opiate addiction and homeostasis (Cox, 1991;Trujillo & Akil, 1991;Di Chara & North, 1992;Meunier, 1992;Law & Loh, 1999; ??estler & Aghajanian, 2007), a plethora of questions exist as to the functionality of these alternatively spliced variants, selectivity of ligand binding, and the implications of these potential associations in disease and therapy (Braaco & Kearney, 2003;Lee & Irizarry, 2003;Brinkman, 2004). The functional capacity of MOR1 splice variants is unknown, and it is yet unclear whether alternativelyspliced isoforms respond to botanical products like the prototypical ligand for the mu-opioid receptor, morphine.

²⁰⁰ 6 Discovery of the Medicinal Properties of Morphine

Recognition of the pharmacological properties of plants and the medicinal use of morphine date far back to 201 ancient civilizations (e.g., Sumeria, Egypt, Ancient Greece, Roman Empire). Among modern narcotic analgesics, 202 morphine is the oldest and remains the gold standard (prototype) that is the most widely used. Morphine is 203 the principal active ingredient in the opium poppy (Papaver somniferum). The groundbreaking discovery of 204 morphine as the first alkaloid isolated from naturally occurring plant species by Wilhelm Serturner, a German 205 Pharmacist, forever changed organic chemistry, medicine, and history. Notwithstanding, morphine is also present 206 in appreciable amounts in Theriaca, laudanum, Doveri, and paregoric (Benyhe et al., 2015). The recreational use 207 of opium is widely (but not exclusively) practiced in the Middle East and the Far East provinces (e.g., Arabia, 208 Turkey, Iran, India, and China), but the illicit sale and use of opium and its synthetic derivatives have since 209 reached global proportions (Benyhe et al., 2015). 210

Subsequent determination of the chemical formula of morphine (Laurent, 1847), the structure of morphine 211 212 ??Robinson, 1925), and its industrial extraction (Kabay, 1925) have led to the total synthesis of morphine ??1952) 213 ??1953) ??1954) ??1955) ??1956) ??Gates and ??sudi, 1952-1956). Gulland elucidated the stereochemical 214 structure of morphine as having a rigid phenanthrene ring system comprised of five condensed rings (A -phenolic, aromatic; B -cyclohexane; C -cyclohexanol, cyclohexene; D -N-methyl-piperidine, piperidine; and E -a partially 215 saturated furan ring, tetrahydrofuran) ??Gulland and Robinson, 1925). The phenolic makeup of the A-ring 216 makes it a weak acid ??Lemke, 2003). Primary and secondary alcohol (-OH) group substitutions at the A-ring 217 C3 and the D-ring C6 positions, respectively, confer chemical reactivity on the molecule. Morphine has five chiral 218 centers at carbon-5 (C5), C6 C9, C13, and C14 positions. The piperidine constituency of the D ring renders 219 morphine the classification of a weak base (Benyhe et al., 2015). By this latter classification, morphine "does not 220

readily donate its electrons and forms an unstable ammonium ion that dissociates readily with a large dissociation constant (K a), and thus has a small pK a " (Lemke, 2003).

²²³ 7 Brief History of the Fig Plant

224 The

225 8 Herb-Disease Interactions

The therapeutic efficacy of Ficus carica has also not been demonstrated; neither is it established as having 226 potential drug-herb interactions with narcotic drugs or nutrients ??Jellin et al., 2009) Studies assessing the ?-227 tocopherol, flavonoid, and phenol contents relative to the antioxidant activity of fig leaves have established the 228 antioxidant capacity of Ficus carica leaf extracts and raised hopes for the role of ?-tocopherol in clarifying 229 its mechanism of action (Konyahoglu et al., 2005). Phytosterols have, in part, been credited for the 230 hypocholesterolemic effect observed in Mission fig (Jeong & Lachance, 2001). In Ghana, the Ficus plant is a 231 popular galactagogue (Bekoe et al., 2018). Also, the nutritive value of the high dietary fiber and high mineral 232 content of figs is superior to many other fruits. There is an established high correlation between total polyphenols, 233 or total anthocyanins, and the antioxidant capacity 234

235 9 Herb-Drug Interactions

Individual Ficus carica cultivars reportedly vary in the antioxidant capacity ??Crisosto et ?? 1997, 2001, 2005,
2011) is important. Comparatively, opportunities to remove nutritional deficiencies through supplemental use of
Ficus carica may become necessary. These attributes support the assumption that the leaves may also possess
a high nutritional and medicinal value that warrants further exploration. Nine cultivars grown in the United
States were selected for further interrogation in the present research (Table ??).

²⁴¹ 10 PCR Comes of Age

One celebrated outcome of the Human Genome Project is its propulsion of the field of molecular genomics 242 into the research spotlight. Innovations in molecular biology and pharmacogenomics, as well as technological 243 advances in quantitative real-time polymerase chain reaction (qRT-PCR), have led to the identification of several 244 new human mu-opioid receptor (MOR-1) splice variants that to date have not been fully characterized (Saiki 245 et al., 1985(Saiki et al., 1988;;Watson, 1990;Olson, 1993;Collins et al., 1998;Pollock, 2002). The polymerase 246 chain reaction (PCR) is a sensitive technology which was discovered by Kary Mullis in 1983 for the original 247 248 purpose of improving DNA quantification (Mullis et al., 1986;Bartlett & Starling, 2003). However, PCR has 249 also led to our improved knowledge of biological processes such as RNA transcription, cellular growth, and proliferation, differentiation, development Advances in PCR technology have advanced the field of gene expression 250 analysis for over twenty-five years, namely: the introduction of real-time PCR (Williams, 2009), discovery of 251 reverse transcription (Baltimore, 1970; Temlin & Mizutani, 1970); qRT-PCR and the emergence of sophisticated 252 instrumentation to detect vanishingly small quantities of nucleic acids (Saiki et al., 1985(Saiki et al., 1988 253 PCR capitalizes on the well-established significance of DNA in living cells following elucidation of the genetic 254 code (Watson & Crick, 1953) as well as the central dogma of molecular biology which posits that the uni-255 directional flow of genetic information is from DNA to RNA, via transcription, and from mRNA (the product 256 of transcription) to protein, via translation (Crick, 1958). Transcription is the first and rate-limiting step in 257 the process of gene expression. The term 'gene expression' is synonymous with 'messenger ribonucleic acid 258 (mRNA) levels.' Quantitative real-time polymerase chain reaction precisely and reliably measures gene expression 259 levels of specific nucleic acid sequences (Kaltenboeck & Wang, 2005;Bustin, 2000Bustin, , 2010;;Bustin & Nolan, 260 2004;Bustin et al., 2005). Close examination of emerging patterns of gene expression can provide insight into 261 physiological responses to cellular stressors or signals, or whether the genes are functionally related (Pollock, 262 2002). The evident superiority of qRT-PCR surpasses older technologies (e.g., Northern blot, RNase protection 263 assays) and affirms its designation as the "gold standard" or method-of-choice for analyzing gene expression of 264 modest numbers of genes (Nedelman, 1992). 265

The biological significance of qRT-PCR to modern biology and biomedical sciences is irrefutable. In the 266 aftermath of discoveries made in the Human Genome Project, scientists have begun to explore more intensely 267 the molecular underpinnings of sickness, chronic disease, and drug interactions in the body (Snider et al., 268 269 2001;Bernard & Wittwer, 2002;Pollock, 2002;Kaltenboeck & Wang, 2005). Answers to elusive medical conditions, 270 such as cancer and drug tolerance, can be elucidated at the molecular level to shed greater insight into the 271 nature of these conditions as well as the mechanisms by which they occur (Braaco & Kearney, 2003;Brinkman, 272 2004;Kaltenboeck & Wang, 2005). The use of this generalized PCR equipment to characterize the plant genome is not new. The efficiency of quantitative real-time PCR in detecting posttranscriptional changes in human cells 273 induced by natural plant products gives way to future consideration of the mechanistic actions of plant extracts, 274 as well as drug-herb interactions. The enhanced capacity for comparative analysis of critical neurological systems, 275 such as the opioid system, using this technology, as well as the potential discovery of relevant interventions to 276 eliminate the cause of chronic diseases, gives hope for the future of medicine. 277

²⁷⁸ 11 Using Ancient Medicinal Herbs to Counteract an Age-Old ²⁷⁹ Enigma

With the recent advances of receptor polymorphisms and gene splicing, several variant forms of MOR-1 have recently been identified. Through the use of polymerase chain reaction (PCR) technology, the fields of molecular genetics and pharmacology have begun to converge and so enable a deeper understanding of the mechanistic basis of opioid-related diseases, like opioid dependence, addiction, and withdrawal, which are chronic outcomes of morphine tolerance.

The present study examined the prototypic effects of morphine on MOR-1 variant mRNA expression compared that of fig (Ficus carica) leaf extract. The objective of this study was to employ advanced molecular genetics techniques, including real-time qRT-PCR, to assess the ability of fig leaf extract, in the presence or absence of morphine, to interact with MOR-1 receptor alternatively-spliced variants (ASVs) and to modify its transcriptional machinery, a rate-limiting step in MOR-1 protein synthesis and functionality of the muopioid receptor (OPRM1).

²⁹⁰ 12 II. Materials and Methods

²⁹¹ 13 a) General Chemical Reagents and Pharmacologic

Agents Isopropanol, chloroform, morphine, distilled water, and consumable supplies were supplied by Sigma-Aldrich (St. Louis, MO, USA). Absolute ethanol (100%) was obtained in-house.

²⁹⁴ 14 i. qRT-PCR Chemicals

The DNAse treatment and removal kit were purchased from Ambion (Foster City, CA, USA). iQ? SYBR Green Supermix and iScript cDNA Synthesis System were ordered from Bio-Rad Laboratories (Hercules, CA).

297 ii. ??).

²⁹⁸ 15 b) Methods

²⁹⁹ 16 i. Human Neuroblastoma (SH-SY5Y) Cell Line

Human neuroblastoma (SH-SY5Y) cells are epithelial cells that were derived from the bone marrow of a 300 metastasized tumor originating in the brain of a 4year old girl. SH-SY5Y cells are stable neuroblasts that were 301 thrice-cloned from the original SK-N-SH cell line ??Ross et al., 1983). The expression of mu-opioid receptors 302 in SK-N-SH cells was determined to be five times higher than that of delta-opioid receptors ??Yu et al., 1986), 303 which is reproduced in SH-SY5Y subclones ??Yu et al., 1986). SH-SY5Y cells are a reproducible cell model for 304 studying the biochemical correlates of opiate efficacy and tolerance (Yu and Sadee, 1988). Additionally, SH-SY5Y 305 cells can express several distinct phenotypes, including immature neuroblast forms that differentiate into mature 306 neurons ?? Ross et For centuries, morphine has been utilized as the prototypical analgesic drug in the treatment of 307 chronic and intractable pain. Morphine exerts its pain-relieving effects primarily through the mu-opioid receptor 308 (MOR-1). Moreover, ancient civilizations have used the fig (Ficus carica L.) plant for wound healing, digestive 309 clearance, and as a hypolipidemic agent in diabetes. Since morphine induces constipation and hyperglycemia, we 310 hypothesized that fig leaf extract could attenuate or abrogate these adrenergic effects of morphine, as well as its 311 central effects. 312

following treatment with retinoic acid (RA) ??Pahlman et al., 1984). It is advantageous to use an in vitro model of specialized nervous system cells (i.e., neurons) in this study because isolation of the effects of chemicals at the molecular level is complicated by the heterogeneity of in vivo nervous system networks within tissues.

ii. Cell Culture Human neuroblastoma (SH-SY5Y) cells (ATCC, Bethesda, MD) were maintained under
sterile conditions in Dulbecco's Modified Eagle's Medium (DMEM)/Ham's F-12 (1:1) (Gibco Laboratories, Grand
Island, NY) supplemented with 10% fetal bovine serum (FBS) and penicillin/streptomycin (Sigma, St. Louis,
MO). The cells were stored in a humidified incubator at 37°C and 5% CO 2. Cells were grown to 70-80%
confluence (15 x 10 6 cells) before differentiation with retinoic acid (RA).

321 17 RA Differentiation of SH-SY5Y Cells

322 Human neuroblastoma SH-SY5Y cells were the first neuronally-derived cell line suitable for studying chronic opiate (morphine) effects ??Zadina et al., 1993). Two unique properties of SH-SY5Y cells that auger well for 323 324 their use in opioid research are their constitutive expression of the mu opiate receptor in measurable quantities 325 ??Toll, 1990; ??are et al., 1994; ??dsjo et al., 2007) and the rare ability of this cell line to be induced to express the neuronal phenotype by addition of retinoic acid (Sidell 1982; ??idell et al., 1983; ??adina et al., 1993; ??u and 326 Sadeee, 1988; ??orner et al., 2007). Differentiation of SH-SY5Y cells ensued after addition of all-trans retinoic 327 acid (10 mM), dissolved in absolute ethanol, to fresh culture medium (final concentration: 10µM). At 70-80% 328 confluence (~15 x 10 6 cells), the cells were exposed to RA for 48 hrs before all media was removed and refreshed. 329

 $_{330}$ $\,$ The cells were reintroduced to RA (10 $\mu M)$ for a further 24-hr period.

331 18 Treatment of SH-SY5Y Cells

Three categories of treatments were used in the experiments, including morphine (the prototypical, full opioid receptor agonist and potent analgesic), fig leaf extract (a crude botanical product), or control (media alone). Plated cells, at or near confluence, were randomly assigned to the respective treatment groups.

³³⁵ 19 Differentiated but Untreated SH-SY5Y Cells [Experimental ³³⁶ Controls]

The measurement outcome (dependent variable) of these experiments is "gene expression." The independent variables examined in this study are "treatment," "time," and "genotype." Control (media alone) received no chemical treatment but a supplemental volume of media only. Triplicate control plates accompanied each set of independent experiments. We evaluated MOR-1, MOR1-A, MOR1-B1, MOR1-B2, MOR1-B3, MOR1-B4, MOR1-B5, MOR1-K1, and ?-ACT genotypes (n=8) in each sample at one of three time-points (n=3), 24hr, 48hr, and 72hr. Plated cells were randomly allocated to the respective treatment groups at or near confluency. A total of 9 plates comprising a single experimental unit were analyzed.

³⁴⁴ 20 Phase Contrast Microscopy

We monitored the growth of SH-SY5Y cell cultures differentiated with RA using phase-contrast microscopy to ensure the progression of neuritogenesis under experimental conditions.

³⁴⁷ 21 c) Primer Design

Primer pairs (Table 2) were designed to recognize and amplify the specific region within the cterminus of the MOR-1 gene where the variant is located. Invitrogen's OligoPerfect? Designer online system was used to design forward and reverse oligonucleotide primers (Invitrogen, Carlsbad, CA), no more than 26 base pairs in length.

³⁵¹ Primers to detect the expression of transcription factors are listed in Table 3.

352 22 d) RNA Isolation

All procedures were performed according to the manufacturer's protocols. Trizol extraction (Invitrogen, Carlsbad, CA) and DNase treatment of total ribonucleic acid (RNA) were used to purify the sample for reverse transcription (Turbo DNA-free? Kit, Ambion, Foster City, CA). The final concentration of the reaction mixture was 10 µg of RNA/50 µl DNase cocktail. Total RNA concentrations were measured before proceeding with the remaining procedures. Samples with Nanodrop? A 260 /A 280 absorbance ratios ?1.8 and adequate RNA concentrations were selected for amplification by realtime quantitative polymerase chain reaction (qRT-PCR) using the Bio-Rad iCycler/MyIQ? (Bio-Rad, Hercules, CA).

³⁶⁰ 23 e) cDNA Synthesis

First-strand cDNA was reverse-transcribed from purified RNA using a 20 µl reaction mixture (iScript? cDNA
Synthesis Kit, Bio-Rad, Hercules, CA) containing 5 µl (1 µg) of RNA. Samples were incubated in the thermocycler
for 30 minutes (25°C, 5 min; 42°C, 15 min, twice; 85°C, 5 min) before storage at -70°C.

³⁶⁴ 24 f) Real-Time Quantitative Polymerase Chain Reaction ³⁶⁵ (qRT-PCR)

Reverse-transcribed cDNA was amplified by RT-qPCR using iQ SYBR Green Supermix®? (Bio-Rad, Hercules, 366 CA) with human forward/reverse primer-probe sets for ?-actin (housekeeping gene), human MOR1A (HMOR-367 1A), HMOR-1B1, HMOR-1B2, HMOR-1B3, HMOR-1B4, HMOR-1B5 and HMOR-1Y genes (25 µl A stock 368 solution of morphine (10 mM) was prepared according to manufacturer's instructions. SH-SY5Y cells were 369 treated with morphine (10μ M), fig leaf extract (3μ L/ 30μ L media), or both for 48 hours as independent triplicate 370 samples. On harvesting, the SH-SY5Y cells were thrice washed with 1X PBS then stored at -70°C until analysis. 371 SYBR, 20 µl water, 3 µl primer, 2 µl cDNA). Optimization of the thermal profile at 95°C (5 min) was followed by 372 a 2-step amplification and melt process over 40 cycles (95°C, 10 sec; 55°C, 45 sec). The thermal cycler (Figure 4) 373 was set to proceed at 95°C (1 hr) followed by 55°C (1 hr), and finally, 55°C (10 sec). The specificity of qRT-PCR 374 was checked by examining melt curves generated for each set of triplicate control, treated, and standard curve 375 samples. 376

377 25 i. Standard Curve

Relative gene expression levels were determined using the standard curve method. For each primer pair (forward and reverse), the amplification efficiency for each gene of interest was based on a fourpoint, 5-fold sample dilution
series. Signal threshold cycle, or C t , values were logarithmically transformed to extrapolate the level of MOR-1
variant mRNA relative to ?-actin (reference gene). Relative expression of an individual gene of interest was

defined as the percentage ratio of log-transformed C t values for treated (C t -treat) samples to the C t value for ?-actin, relative to controls (C t -control).

³⁸⁴ 26 g) Quality Assurance/Control

The specificity of qRT-PCR was checked by examining melt curves generated for each set of triplicate control, treated, and standard curve samples. Before each use, the Nanodrop? and analytical scale were sanitized and calibrated between after each use according to standard laboratory procedures. Microvolumes of each sample were loaded onto the pedestal as RNA purity and quantity were assessed spectrophotometrically.

³⁸⁹ 27 h) Normalization of qRT-PCR Data

Replicate samples should be run at least in triplicate assays, and the experiments repeated at least thrice. Relative gene expression is calculated based on the standard curve method (as above) and normalized by housekeeping and control genes. The data should be subjected to dual normalization based on the ratio of 'log base two' equivalent values for target and control genes. For example, the proportion of 'target gene: housekeeping gene' and 'target gene: control gene' were computed.

³⁹⁵ 28 i) Statistical Analysis

The measurement outcome (dependent variable) of these experiments is "gene expression," quantified as relative messenger RNA (mRNA) levels. The independent variables examined in this study are "treatment" and "genotype." For qRT-PCR, MOR-1 and selected variant forms (i.e., MOR-1A, MOR-1B1, MOR-1B2, MOR-1B3, MOR-1B4, MOR-1B5, and MOR-1K1), as well as ?-ACT genotypes (n=8) were evaluated.

The data (mean \pm SEM) represent triplicate assays of samples obtained from three independent experiments. The small sample size represents a limitation on this pilot study that does not appear to deface the quality of the data. Dataset organization and basic descriptive statistics were calculated using Microsoft Excel®. The data

403 were then normalized to ?actin mRNA and control values.

Statistical analyses and graphics were performed using the Prism 6.0? software. Statistical significance of t-tests was set at an alpha level of p<0.05.

406 29 III. Results

407 30 a) RA Differentiation in SH-SY5Y Cells

Retinoic acid (RA) induced differentiation of native SH-SY5Y cells into cells morphologically classifiable as neuronal cells, as confirmed by the presence of dendritic formations, neurite outgrowths, and axonal extensions.

⁴¹⁰ 31 i. Expression of MOR-1 ASVs in Experimental Control

411 SH-SY5Y Cells Untreated but RA-differentiated (control) cells exhibited significant (p<.0001) constitutive, 412 differential expression of all MOR-1 alternative splice variants as well as beta-actin (Figure 5). MOR-1B4 was 413 undetected.

414 **32** b) Tolerogenic Effect of Morphine on Differentiated SH-415 SY5Y Cells

Based upon our present preliminary screen of mRNA extracted from RA-differentiated human neuroblastoma 416 (SH-SY5Y) cells and analyzed by qRT-PCR using Bio-Rad Thermocycler/MyIQ® software, prototypical opioids 417 induced measurable tolerogenic effects within 48 hours of opioid exposure. Treatment with morphine alone 418 significantly down-regulated MOR-1B1 (77.32%, p<.0001), MOR-1B2 (70.10%, p<.0001), MOR-1B3 (92.96%, 419 p<.005), and MOR-1K1 (82.18%, p<.0001) mRNA levels relative to controls in BACTnormalized samples. In 420 contrast, MOR-1A (179.7%, p < .05) and MOR-1B5 (109.3%, p < .0001) in these samples were significantly up-421 regulated following morphine treatment (Figure 6). Compared to the responses of the other variants in morphine-422 treated samples, the effect on MOR-1A may be an outlier as an artifact of a small sample size. MOR-1B4 was 423 undetected. 424

425 **33** c) Effect of Fig Leaf Extract on Differentiated SH-SY5Y 426 Cells

Treatment of SH-SY5Y cells with Alma fig leaf extract for 48hr substantially amplified the expression of MOR-1A (396.1%, p<.0001), MOR-1B1 (440.1%, p<.05), MOR-1B2 (239.1%, p<.05), MOR-1B5 (259.1%, p<.005), and MOR-1K1 (230.2%, p<.05), relative to controls. There was inadequate evidence of MOR-1B3 down-regulation by the Alma fig cultivar (Figure 7).

Compared to the responses of the other variants in Alma fig leaf extract-treated samples, the effect on MOR 1B3 may be an outlier as an artifact of a small sample size. MOR-1B4 was undetected. On examining patterns of

expression following administration of Alma fig only, the inflated mRNA values suggest a synergistic interactionwith endogenous opiates.

435 34 d) Combined Effect of Morphine and Fig Leaf Extract on 436 MOR-1 ASV Expression

Cells initially treated with morphine were subsequently treated with Alma fig leaf extract. In the morphine/Alma fig treatment group, MOR-1B1 (459.6%, p<.05), MOR-1B2 (228.1%, p<.05), MOR-1B5 (301.8%, p=.0069), and MOR-1K1 (156.6%, p<.005) mRNA levels were found to be up-regulated, whereas MOR-1A1 (65.4%, p>.05) and MOR-1B3 (77.02%, p<.0001) mRNA levels were down-regulated (Figure 8).

Compared to the responses of the other variants in morphine-treated samples, the effect on MOR-1A and
 MOR-1B3 may be an outlier as an artifact of a small sample size. MOR-1B4 was undetected.

The addition of Alma fig extract completely abrogated the tolerogenic effects of morphine on MOR-1B1, MOR-443 1B2, and MOR-1K1. When morphine was administered alone, there was an observed characteristic attenuation 444 of mRNA levels. The marked inflation of mRNA levels in morphine/fig samples suggests that Alma fig leaf 445 extract may indeed have "inverse agonist" properties, as it is customary for inverse agonists to elicit the opposite 446 effect to that of an agonist to the receptor. This pattern of opposites was observed relative to MOR-1A, MOR-447 1B1, MOR-1B2, and MOR-1K1 when comparing "morphine"-treated to "morphine/fig"-treated samples. Also 448 prominent were the double to triple amplification of MOR-1B5 signals, approximating additive effects (morphine 449 alone -109.3%; Alma fig alone -259.10%; morphine+Alma fig -301.8%). 450

⁴⁵¹ 35 IV. Discussion and Conclusion a) Model Selection

452 Human neuroblastoma (SH-SY5Y) cells were the first neuronally-derived cell line deemed suitable for the in vitro

453 study of chronic opiate (morphine) effects ??Zadina et al., 1993). SH-SY5Y cells also continue to be a reliable

454 model for its current use in the expression of mu-opioid receptor variants ??Toll, 1990; ??are et al., 1994; ??dsjo 455 et al., 2007) due to its high constitutive expression of this receptor and its ability to be induced by retinoic acid

to express the neuronal phenotype ??Sidell et al., 1983; ??adina et al., 1993;Yu and Sadee, 1988).

457 36 b) Pilot Study

This study confirms our hypothesis that muopioid receptor (MOR-1) alternatively spliced variants are sensitive 458 and differentially responsive to prototypical opioids as well as botanical products (i.e., Ficus carica leaf extract). 459 Relative to ligand binding, these data indirectly suggest that some constituent in the fig (Ficus carica) leaf 460 appears compatible with the mu-opioid receptor, can bind to the MOR-1 binding site, and is capable of triggering 461 a signaling cascade that elicits genetic effects at successive DNA, RNA and posttranscriptional levels. This 462 constituent is probably structurally similar to morphine or one of its precursors. Given the dependency of gene 463 expression on the tightly regulated, successive steps of transcription, it is reasonable to conclude that there is 464 evidence for functional modulation of MOR-1 in neurons. 465

466 37 c) Limitations

467 Due to the small sample size of this pilot study, expanded analyses under experimental conditions are warranted.
468 The data confirm the efficacy of customdesigned primers for targeting specific regions of the OPRM1 gene and
469 the distinctive value of individual Ficus cultivars in interacting with the mu-opioid receptor.

470 **38** d) Conclusions

Alma fig leaf extract targets specific exons within the mu-opioid receptor (MOR-1) gene (OPRM1) to reverse morphine-induced down-regulation of MOR-1 alternatively-spliced variants. The differential expression of MOR-1 isoforms in response to Alma fig and/or morphine/Alma fig leaf extract, as well as the appearance of additive, synergistic and inverse interactions between these botanicals and human cells, suggests a potential future role in resolving inter-and intra-individual differences in response to morphine. The current finding brings us a little closer to an approach for discriminating the functions of individual MOR-1 ASVs and may play a future role in identifying herb-drug interactions that affect medical prescribing and medication management practices.

Further work is needed to characterize Alma fig leaf extract and its implications for cancer and pain therapy. 478 Added attention to the reversing effects of Alma fig leaf extract following morphine treatment is needed as 479 this outcome may prove useful for reversing morphine-induced side effects, such as constipation and tolerance 480 scientific communications signature on the bedrock of my professional training that will last a lifetime. Funding 481 sources included the National Institutes of Health (NIH) grants (RR08111 and RR03020), the FAMU Title III 482 Program, as well as minimal personal support. This research is associated with the American Association for 483 Cancer Research (AACR) 101 st An authentic receptor should be recoverable in its natural (non-metabolized) 484 form. If the gene for such receptor is isolated and expressed, it should be exactly similar to the cloned receptor 485 of the natural receptor. 1 Adopted from Hollinger (1997) 2. 486

487 Figure Legends

D) CONCLUSIONS 38

488

Receptors can contain secondary modifications of carbohydrate and be selectively embedded into the lipid membrane bilayer (Norman & Litwack, 1997). 3 Regardless of where they have been isolated from, studies 489 show that neurotransmitter and peptide hormone receptors are localized on the cell surface. All receptors have 490 an effector domain that "recognizes" the presence of the hormone bound to the ligand domain and that then 491 $1 \ 2$ initiates the generation of the biological response(s) (Norman & Litwack, 1997).



Figure 1:

492

¹Review & Pilot Study

 $^{^{2}}$ \odot 2020 Global Journals Neuromodulation of Mu-opioid Receptor (MOR-1) Gene (OPRM1) Alternatively-Spliced Variants Following Exposure to Morphine with Alma Fig (Ficus carica) Leaf Extract in Human Neuroblastoma (SH-SY5Y) Cells: Review & Pilot Study



Figure 2:



Figure 3:

point for moderate to service point
+/-non cpioid +/- adjuvant
pain persisting or increasing
opioid for mild to moderate pain +/-non opioid +/- adjuvant
pain persisting
or increasing
non opioid +/- adjuvant

Figure 4:



Figure 5:



Figure 6:



Figure 7:



1

Figure 8: Figure 1 :



Figure 9: Figure 2 :



3

Figure 10: Figure 3 :

 $\mathbf{4}$



Figure 11: Figure 4 :



Figure 12: Figure 5 :



Figure 13: Figure 6 :



Figure 14: Figure 7 :

Figure 15:

Figure 16:

Researchers have examined the weight-ofevidence of herb-drug interactions pertaining to the fig leaf and have concluded that concern is both relevant and valid based on available literature (i.e., nonrandomized clinical trial [RCT]; non-quantitative systematic review; lower quality RCT; clinical cohort study; case-control study; historical control; or epidemiologic study) (Jellin et al., 2009). The severity of interactions of fig leaf with two drugs has been rated as "moderate," and caution is advised with these combinations. Clinical research or pharmacokinetic data in humans suggests that this interaction is "probable," meaning that it will occur in a significant portion of patients. Fig leaf may interact with anti-diabetic drugs or insulin. In both cases, fig leaf lowers blood glucose levels by enhancing the effect of hypoglycemic drugs (Jellin et al., 2009). Mechanistically, fig leaf is capable of improving glucose update by skeletal muscle (Jellin et al., 2009). However, there was no clear evidence as to the implications of other herb-drug interactions [such as motor function drugs (e.g., skeletal muscle relaxants benzodiazepines; anti-seizure drugs -phenobarbital), or centrally acting drugs that affect smooth muscles (e.g., morphine)] or herb-disease interactions [such as drugs affecting skeletal muscle tone in Parkinson's Disease and other movement disorders]. Finally, an analysis of the mineral content of the fructus and folium of Ficus carica L. revealed superior concentrations of calcium, potassium, magnesium, phosphorus and sulfur in folium $(27,611 \pm 152 \text{ µg/g})$; $16,000 \pm 234 \ \mu g/g; 3,565 \pm 174 \ \mu g/g; 1,285 \pm 31 \ \mu g/g;$ and $1,150 \pm 67 \,\mu\text{g/g}$ respectively) versus fructus (6,006

 \pm 613 µg/g; 13,892 \pm 415 µg/g; 1,381 \pm 186 µg/g; 1,054 \pm 44 µg/g; and 536.1 \pm 7.5 µg/g respectively) (

Figure 17:

Figure 18:

		Review & Pilot Study	
Nero	Leaf has a cordate	Large, reddish black fig;	ItalDark
	base and		green
(Barnisotte)	5 lobes with the mid-	eye is medium-sized and	(pH)
	dle one		5.78,
			5.79)
	being spatulate and	open; shape is turbinate-	
	the		
	others latate.	pyriform,	sometimes
		oblique with a broad apex.	
4 Jeong & Lachance	(2001).		
5 Bekoe et al. (2011))		

NR = not recorded

[Note: 1]

Figure 19:

 $\mathbf{2}$

Primer	Туре	Sequence $(5' \text{ to } 3')$	Base	sG/C
				Count
hMOR-1 1F	Forward (Sense)	ATGCCAGTGCTCATCATTAC	20	9
hMOR-1 1R	Reverse (Antisense)	GATCCTTCGAAGATTCCTGTC	QT	11
hMOR-1A 1F	Forward (Sense)	CAGGTACGCAGTCTCTAGAAT	725GG	$\mathbf{G}12$
hMOR-1A 1R	Reverse (Antisense)	TTCCCTCCATTCTCATCCTC	20	10
hMOR-1B1 1F	Forward (Sense)	TCAAAAGTCATCTTTACTCAA	C26G]	₽ G
hMOR-1B1 1R	Reverse (Antisense)	GCTTCCAATCTTATATTCTTTC	CACG	9
$hMOR-1B2 \ 1F$	Forward (Sense)	AAAGAAGACAGAAATCTGACT	C C GT	A9A
hMOR-1B2 1R	Reverse (Antisense)	GCAAGCCGGATCACTAGG	18	11
hMOR-1B3 1F	Forward (Sense)	TTTGTTGCTGACCAACTTGC	20	9
$hMOR-1B3 \ 1R$	Reverse (Antisense)	GGTCGTTTTTTTTTGTGTTGAGG	G21	10
$hMOR-1B5 \ 1F$	Forward (Sense)	GGAATTGAACCTGGACTGTCA	A21	10
$hMOR-1B5 \ 1R$	Reverse (Antisense)	AAGCCTTCGCAAACTCAAAA	20	8
hMOR-1K1 1F	Forward (Sense)	CTGGGTAGGAAAGTGGCAAA	20	10
hMOR-1K1 1R	Reverse (Antisense)	TGACCTTGGTGCTCAAGAAG	$\Gamma 21$	10

Figure 20: Table 2 :

3

Type

38 D) CONCLUSIONS

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