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In-Vitro Anti-Microbial and Anti-Urolithiatic Models for Extract of Parmelia Perlata Lichen: An Evaluation of Prophylactic 2 Management Against Kidney Stone 3 Δ

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Abstract 7

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Abstract- Background: Phyto-medicine is regaining interest owing to its advantages over 8 conventional drugs and increasing cases of drug resistance. Moreover, recurrence and 9 persistent side effects of present-day treatment for urolithiasis restrict their use, so an 10 alternate solution using phytotherapy is being sought. The lichen species Parmelia Perlata 11 (family Parmeliaceae) mentioned in India Materia Medica are useful in treating several 12 ailments, and they are being used in large quantities as a food supplements in India. 13 Objective: Up to date, the scientific documentation regarding in-vivo antiurolithiatic and 14 antimicrobial activity of Parmelia perlata has been reported, although it has not reported for 15 invitro antiurolithiatic as well as an antimicrobial activity up till now. The present study 16 attempted to evaluate phytochemical screening, antimicrobial activity, and anti urolithiatic 17 activity of methanolic lichen extracts of Parmelia Perlata, which called ?Chandila? by in vitro 18 model. Material and Methods: In this study, the crude extracts were obtained from the 19 Parmelia perlata by cold extraction method using methanol as a solvent. The phytochemical 20 tests were being carried out on the extract of lichen. The antimicrobial efficacy was being 21 investigated against various pathogenic bacterial and fungal strains. In vitro antiurolithiatic 22 models were investigated through nucleation assay and aggregation assays by 23 spectrophotometric technique. Cystone was being used as a standard drug in the in vitro 24 model for anti urolithiatic activity. Result: The result of the phytochemical tests showed the 25 presence of several biologically active phytochemicals with the highest quantity of alkaloids, 26 flavonoids, and phenols in methanolic extract. The methanolic extract had the highest activity 27 against P.vulgaris and C. tropicalis at 160 mcg/mL concentration. The maximum percentage 28 of dissolution of existing calcium oxalate crystal was found to be 35 29

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- parmelia perlata, lichen, phytochemicals, antimicrobial, anti-urolithiatic. Index terms-

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In-Vitro Anti-Microbial and Anti-Urolithiatic Models for Extract of Parmelia Perlata Lichen: An Evaluation 32 of Prophylactic Management against Kidney Stone Dhara Patel?, Ritu Sapra?, Grishma Patel? & Dhananjay 33 34 Mesharm ? Phyto-medicine is regaining interest owing to its advantages over conventional drugs and increasing 35 cases of drug resistance. Moreover, recurrence and persistent side effects of present-day treatment for urolithiasis 36 restrict their use, so an alternate solution using phytotherapy is being sought. The lichen species Parmelia Perlata (family Parmeliaceae) mentioned in India Materia Medica are useful in treating several ailments, and they are 37 being used in large quantities as a food supplements in India. 38

Objective: Up to date, the scientific documentation regarding in-vivo antiurolithiatic and antimicrobial 39 activity of Parmelia perlata has been reported, although it has not reported for invitro antiurolithiatic as well 40 as an antimicrobial activity up till now. The present study attempted to evaluate phytochemical screening, 41 antimicrobial activity, and anti urolithiatic activity of methanolic lichen extracts of Parmelia Perlata, which 42

43 called "Chandila" by in vitro model. Material and Methods: In this study, the crude extracts were obtained 44 from the Parmelia perlata by cold extraction method using methanol as a solvent. The phytochemical tests 45 were being carried out on the extract of lichen. The antimicrobial efficacy was being investigated against various 46 pathogenic bacterial and fungal strains. In vitro antiurolithiatic models were investigated through nucleation 47 assay and aggregation assays by spectrophotometric technique. Cystone was being used as a standard drug in 48 the in vitro model for anti urolithiatic activity.

Result: The result of the phytochemical tests showed the presence of several biologically active phytochemicals 49 with the highest quantity of alkaloids, flavonoids, and phenols in methanolic extract. The methanolic extract 50 had the highest Introduction rolithiasis, the formation of kidney stone presence of one or more calculi in any 51 location within the urinary tract, is one of the oldest and wide spread diseases known to man. [1] It is a serious, 52 debilitating problem in all societies throughout the world, affecting approximately 12% of the population and, 53 men are three times more prone than women. [2] It is more prevalent between the ages of 20 and 40 in both 54 sexes. [3] Etiology is multifactorial and is strongly related to dietary lifestyle habits or practices. [4] Increased 55 rates of hypertension and obesity also contribute to an increase in stone formation. [5] Kidney stones are hard, 56 solid particles that form in the urinary tract. In many cases, the stones are very small and can pass out of the 57 58 body without any problems. However, if a stone (even a small one) blocks the flow of urine, excruciating pain 59 may result, and prompt medical treatment may be needed. Recurrent stone formation is a common part of the 60 medical care of patients with stone disease. Calcium-containing stones, especially calcium oxalate monohydrate, 61 calcium oxalate dihydrate and, basic calcium phosphate are the most commonly occurring ones to the extent of 75-90% followed by magnesium ammonium phosphate (Struvite) to the extent of 10-15%, uric acid 3-10% 62 and cystine 0.5-1% [6]. The present-day medical management of nephrolithiasis is either costly or not without 63 side-effects. Invasive procedures for the treatment of nephrolithiasis may cause serious complications and also 64 impose a burden of costs on the healthcare system [7]. 65

Lichens are being used since ancient times as one of the natural drugs [8]. Lichens represent a symbiotic association of a fungus with an algal partner and are important constituents of the ecosystem. Lichens produce characteristic secondary metabolites such as aliphatic, aromatic, and terpenic components which have considerable biological activities such as antiviral, antibacterial, antifungal, antitumor, antioxidant, etc. [9,10]. Parmelia perlata is a well-known lichen of family Parmeliaceae. A lichen is an association of an alga and fungus living together in a symbiotic relationship.

living together in a symbiotic relationship.
 Parmelia perlata is commonly called a Stone flower or Chadila. In India it is mainly found in Himachal Pradesh

73 and West Bengal. It is used as food, fodder and medicine. It is a good pain reliever and is being used as a remedy for the early healing of wounds. It cures many skin diseases and is considered to be an expectorant, astringent, 74 resolvent, laxative, carminative and, aphrodisiac. It is also used in the treatment of fever, cough, dysentery 75 and, renal calculi. This lichen exhibits antimicrobial [11][12], antiviral [13], antitumor [14], antispasmodic 76 [15], antioxidant [16] and antipyretic [17] activities. Its hepatoprotective action [18] has also been reported. 77 Phytochemical studies of Parmelia perlata have led to the isolation of various chemical constituents such as 78 atranorin, chloroatranarin, salazinic acid [19], lecanoric acid ??20], imbricaric acid [21], lecanora. The study 79 has been undertaken to evaluate Parmelia perlata different lichen extracts and cystone as a standard for their 80 possible potential to dissolve experimental kidney stone using a modified in vitro model [22][23][24][25] to isolate 81 the chemical constituent responsible for the activity. 82

83 **1 II.**

⁸⁴ 2 Material and Methods

3 a) Source of Plant Materials

The plant material Parmelia perlata (lichen) was collected from the hills of Himachal Pradesh (India). The collected lichen species were identified as Parmelia perlata (Rajnegi and Gadgil, 1996) (Figure 1).

⁸⁸ 4 b) Source of microorganisms

89 The

⁹⁰ 5 c) Extraction of lichen

The collected lichen materials were brought to the laboratory, air-dried for three days, cleaned free of any other plant materials or mosses, and then washed under running tap water. They were oven-dried at 40°C for 42 h and grounded into powder by using a mixer. The powdered samples were stored in sterilized specimen bottles until when needed. Lichen constituents were extracted by cold extraction.

95 6 d) Cold extraction

 $_{\rm 96}$ $\,$ 10 g of lichen powder was added to 200 ml of acetone. The mixture was timed thoroughly by using a shaker

water bath for five hours, then left at room temperature overnight and filtered using Whatman No. 1 filter paper.
The filtrate was collected, and the solvent was removed using rotary evaporator and about 200 mg residues were

recovered. The lichen powder that remained on the filter paper was dried and again extracted using 200 ml
methanol. From this solvent, about 162 mg of residue was recovered [26].
Phytochemical analysis for the qualitative detection of alkaloids, glycosides, reducing sugar flavonoids, tannins,
and saponins were performed with the extracts.

¹⁰³ 7 f) Determination of Antimicrobial Activity

The antimicrobial activity of the lichen extracts was determined using the agar well diffusion method [28] by following the known procedure. Briefly, Nutrient agar was inoculated with the given microorganisms by spreading the bacterial, and fungal inoculums on the media. Wells were made in the agar using the stainless steel borer of 8 mm and filled with 80, 120 and 160 µl of plant extracts. Control wells containing neat solvents (negative control) were also run parallel on the same plate. The plates were incubated at 37°C for 72 hours, and the antimicrobial activity was assessed by measuring the diameter of the zone of inhibition.

¹¹⁰ 8 g) Evaluation for Anti-Urolithiatic Activity

Behind this activity, the idea was to know the role of plant extract in dissolving the already formed stones nucleus in the renal system. For this artificial calcium oxalate crystals were prepared in the laboratory [29] and semi-permeable membrane was prepared from the egg using standard methods [30][31][32].

Step-1: Preparation of experimental kidney stones (Calcium oxalate stones) by homogenous precipitation 147 In 1.47gm of calcium chloride dihydrate was dissolved in 100 ml distilled water, and 1.34 gm of sodium oxalate was 159 dissolved in 100 ml of 2N H 2 SO 4. Both solutions were mixed equally in a beaker to precipitate out calcium 170 oxalate with stirring. An equimolar solution of calcium chloride dehydrate (AR) in distilled water and Disodium 171 hydrogen phosphate (AR) in 10 ml of (2N H 2 SO 4) was allowed to react in a sufficient quantity of distilled 172 water in a beaker. The resulting precipitate was calcium phosphate. Both precipitates freed from traces of H 2 173 SO 4 by ammonia solution. Washed the precipitates with distilled water and dried at 60 0 C for 4 hours.

¹²¹ 9 Step -2: Preparation of semi-permeable membrane from farm

eggs

The semi-permeable membrane of eggs lies in between the outer calcified shell and the inner contents like albumin & yolk. The apex of eggs was punctured by a glass rod to squeeze out the entire content. Empty eggs were washed thoroughly with distilled water and placed in a beaker consisting of 2 M HCl for an overnight, which caused complete decalcification. Further, washed with distilled water, placed in ammonia solution for neutralization of acid traces in the moistened condition for a while & finally rinsed with distilled water (Figure ??) and stored in the refrigerator at a pH of 7-7.4.

129 10 Step-3:

130 Estimation of Calcium oxalate by Spectrophotometrically:

Group I: 1ml of calcium oxalate (1mg/ml) + 1ml of distilled water Group II: 1ml of calcium oxalate (1mg/ml) 131 + 1ml of Cystone solution Group III: 1ml of calcium oxalate (1mg/ml) + 1ml of the cold extract of Parmelia 132 perlata All groups were packed together in egg semipermeable membrane tied with thread at one end and were 133 suspended in a conical flask containing 150 ml, 0.1 M Tris buffer each. At another end of thread tied by a stick 134 placed on the mouth of the conical flask and covered with aluminum foil. All groups were kept in an incubator, 135 pre heated to 37°C for 4 hours, kept for three days. The entire content of each group was removed from the 136 sutured semi permeable membrane and was transferred into the test tube individually. 4ml of 1N H 2 SO 4 and 137 60-80µl of 0.02M KMnO 4 were added and kept aside for 2 hours. Color change from dark pink to colorless was 138 observed after 2 hours. The change of color intensity was measured against 620 nm spectrophotometrically. 139

The concentration of undissolved calcium was determined from a standard calibration curve of calcium oxalate by using the measured absorbance readings.

¹⁴² 11 h) Nucleation assay (Turbidity method):

143 The inhibitory activity of the extracts on the nucleation of calcium oxalate crystals was determined by a 144 spectrophotometric assay. Crystallization was initiated by adding 100 µl of 4 mM calcium chloride and 100 µl of 50 mM sodium oxalate solutions to 0.5 ml of normal human urine, both prepared in a buffer containing 145 0.5 ml of 0.05 mM Tris buffer and 0.5 ml of 0.15mM NaCl solution at pH 6.5 and 37° C and adjusted to volume 146 by adding 1.5 ml of distilled water. The rate of nucleation was determined by comparing the induction time of 147 crystals (time of appearance of crystals that reached a critical size and thus became optically detectable) in the 148 presence of the extract and that of the control with no extract. The optical density (OD) was recorded at 620nm, 149 and the percentage inhibition calculated as (1-OD (experimental)/OD (control))/100. 150

¹⁵¹ 12 i) Aggregation assay

The rate of aggregation of the calcium oxalate crystals was determined by a spectrophotometric assay with slight modifications. The calcium oxalate monohydrate (COM) crystals were prepared by mixing both the solutions of calcium chloride and sodium oxalate of 50 mM each. Both solutions were then equilibrated. The solutions were then cooled to 37°C and then evaporated. The COM crystals were then dissolved with 0.5ml of 0.05mM Tris buffer and 0.5 ml of 0.15mM NaCl solution at pH 6.5 to a final concentration of 1 mg/ml. Absorbance at 620 nm was recorded. The rate of aggregation was estimated by comparing the slope of turbidity in the presence of the extract against control.

159 **13 III.**

160 14 Results

¹⁶¹ 15 a) Qualitative Phytochemical Analysis

The present study reveals that lichen extract shows the presence of phytochemical constituents like alkaloids, flavonoids, carbohydrates, glycosides, proteins, saponins, tannins, terpenoids, reducing sugar and volatile oil in solvent extracts as shown in Table 1.

165 16 b) Antimicrobial Activity

The potential sensitivity of the extract was obtained against all the microorganisms tested, and the zone of inhibition was recorded. The results obtained were compared against standard antibiotic kanamycin and amphotericin B and presented below in the tabulation drawn (Table ??).

¹⁶⁹ 17 c) Estimation of Calcium oxalate

In kidney stones formation, calcium oxalate and calcium phosphate or other chemicals in the urine form crystals on the inner surfaces of kidneys. This stage is called an initial mineral phase formation. Over the period, crystals may combine to form a small, hard mass called as stones, and the stage is referred to crystal growth. Calcium oxalate stones have been classified into two types, i.e., calcium oxalate monohydrate stones (COM) and calcium oxalate dihydrate stones (COD).

175 18 d) Spectrophotometric estimation of calcium oxalate

The extract of Parmelia perlata has greater capability to dissolve calcium oxalate as the foremost element for stone forming in urinary tract. Lower percentage indicates more potency in the dissolution of calcium oxalate crystals as shown in Table ??.

179 19 e) Nucleation assay

Urine supersaturation attributes to calcium oxalate particles crystallization within the urinary tract. This is a 180 nucleation process where stone-forming salts begins to unite into clusters with the addition of new constituents. 181 Cystone standard solution exhibited stronger inhibition activity than the extract of Parmelia perlata in the 182 nucleation of calcium oxalate salts. As in vitro crystallization study was performed, since nucleation is an 183 important first step for the initiation of crystals, which then grow and form aggregates, extract of Parmelia 184 perlata inhibited crystallization by inhibiting the nucleation of calcium oxalate through disintegrating into smaller 185 particles with increasing concentrations of the fraction. The results of nucleation assay confirmed that the extract 186 contained nucleation-preventing agents (Figure ??). 187

¹⁸⁸ 20 f) Aggregation assay

Calcium oxalate crystals begin to grow, aggregate with other crystals and, retained in the kidney. This is an 189 aggregation process that causes renal injury. The extract of Parmelia perlata inhibited formation of COD crystals 190 slightly better compared to Cystone standard solution. COM has a stronger affinity with cell membranes; it may 191 lead to become a higher potential risk for renal calculi formation. This is may be due to the high content of 192 saponins present in Parmelia perlata. It has several polyphenolics, e.g., alkaloids, saponins, phenolics, flavonoids, 193 and other phytoconstituents. Saponins are well known to have anti-crystallization properties by disaggregating 194 195 the suspension of mucoproteins as crystallization promoters. The present investigation will be supportive as 196 additional information to the scientific evidences regarding in-vitro studies (Figure 4). Since the mechanism of anti-urolitholytic activity in the extract is exactly unknown to date, the correlation between in vitro and in 197 vivo studies should be further investigated to reveal the phytochemicals of the extract which are responsible for 198 dissolving or disintegrating renal calculi and for knowing better understanding in the molecular mechanism of 199 lithiolysis. 200

201 IV.

202 21 Conclusion

The present study conclusively demonstrates that Parmelia perlata is a good source of various phytochemicals 203 like alkaloids, flavonoids, carbohydrates, glycosides, saponins, tannins, Terpenoids. This study evaluates the 204 antiurolithiatic and antimicrobial activity of methanolic extracts of Parmelia perlata Lichen. The work was 205 performed by using an in vitro antiurolithiatic model for calculating the percentage dissolution of kidney 206 stones. This study has given primary evidence for Parmelia perlata as the lichen which possesses lithotriptic 207 property. From the result Tables, it is also clear that a positive correlation exists between individual extracts 208 and concentration used, in the study. Out of four concentration used we can observe that activity increase as we 209 increase the concentration and, at one point, further no increase in the activity observed. The plant used in the 210 above study also showed good activity when it was compared with the standard drug cystone. The methanolic 211 extract found to be more potent in terms of activity and the authors of the above work recommends the lichen 212 extract for further studies by conducting the in vivo model.



Figure 1:

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Figure 2: Figure 1 :



Figure 3: Figure 2 : Figure 3 :



Figure 4: Figure 4 :

1

Extract Alkaloid Flavanoids Saponin Terprnoid Tannin Glycoside Reducing sugar Volatile oil Methanolic + + + + + + + +



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Extract

Cystone

Figure 6: Concentration (mg/ml) Aggregation Assay

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