Global Journal of Medical Research: B
Pharma, Drug Discovery, Toxicology & Medicine
Volume 20 Issue 6 Version 1.0 Year 2020
Type: Double Blind Peer Reviewed International Research Journal
Publisher: Global Journals Inc. (USA)
Online ISSN: 2249-4618 & Print ISSN: 0975-5888

Phytochemical Analysis, Antimicrobial and Radical Scavenging Properties of Methanol Extracts of *Dracaena Deisteliana* (Dracaenaceae) and *Sporobolus Indicus* (Poaceae)

By Afagnigni A.D., Nkonpa R.K. & Mofor C.T.
University of Yaounde I, Cameroon

Abstract- *Dracaena deisteliana* and *Sporobolus indicus* are medicinal plants with broad use in Cameroonian folk medicine to treat several infectious diseases. This study aimed to investigate the phytochemical composition, the antimicrobial and antiradical properties of methanol extracts of the leaves, stem and whole plant of *D. deisteliana,* and *S. indicus.* The phytochemical test was undertaken using standard methods. Agar well diffusion was used for sensitivity test while the microdilution method was used to determine the minimum inhibition concentrations (MICs) and the minimum bactericidal/fungicidal concentrations (MBCs/MFCs). The antiradical property of the plant extracts was performed using the 2.2-diphenyl-1-picrylhydrazyl (DPPH) scavenging assay.

Keywords: *Dracaena deisteliana, Sporobolus indicus, phytochemical, antibacterial, antifungal, antiradical.*

GJMR-B Classification: *NLMC Code: QV 704*

Phytochemical Analysis, Antimicrobial, and Radical Scavenging Properties of Methanol Extracts of *Dracaena Deisteliana* (Dracaenaceae) and *Sporobolus Indicus* (Poaceae)

Afagnigni A.D. [α], Nkonpa R.K. [σ] & Mofor C.T. [ρ]

Abstract- *Dracaena deisteliana* and *Sporobolus indicus* are medicinal plants with broad use in Cameroonian folk medicine to treat several infectious diseases. This study aimed to investigate the phytochemical composition, the antimicrobial and antiradical properties of methanol extracts of the leaves, stem and whole plant of *D. deisteliana,* and *S. indicus.* The phytochemical test was carried out using standard methods. Agar well diffusion was used for sensitivity tests, while the minimum inhibitory concentrations (MICs) and the minimum bactericidal/fungicidal concentrations (MBCs/MFCs) were determined through microdilution method. The antiradical property of the plant extracts was investigated using the 2.2-diphenyl-1-picrylhydrazyl (DPPH) scavenging assay. The results revealed the presence of phenols, saponins, terpenoids, anthraquinones, alkaloids, and flavonoids in all the extracts. *D. deisteliana* extracts inhibited the growth of six bacteria strains with the inhibition zones varying from 8.50 ± 0.28 to 18.00 ± 00 mm and the MIC values ranging between 6.25 and 100 mg/mL. The leaf extract of *D. deisteliana* exhibited a higher effect on *Klebsiella oxytoca* and *Escherichia coli* with the MIC of 6.25 mg/mL. The stem and whole plant extracts showed similar activities on *Escherichia coli* and *Enterobacter cloacae*. *S. indicus* was most active on *Acinetobacter sp* and *Bacillus cereus*. *D. deisteliana* stem extract exhibited higher activity with EC_{50} of 491 µg/mL, while *S. indicus* showed an EC_{50} of 550.5 µg/mL. This study indicates that *D. deisteliana* and *S. indicus* possess antimicrobial and antiradical compounds and provides scientific evidence for their traditional uses to treat several infections.

Keywords: *Dracaena deisteliana, Sporobolus indicus, phytochemical, antibacterial, antifungal, antiradical.*

Corresponding Author α: *PhD. Researcher, Department of Biochemistry, Faculty of Science, University of Yaounde I; PO BOX: 812 Yaounde, Cameroon. e-mail: afagnigni2007@yahoo.fr*

Author σ: *PhD. Senior Lecturer, Department of Biological Sciences, Higher Teachers' Training College, University of Yaounde I. PO BOX: 812 Yaounde, Cameroon.*

Author ρ: *PhD. Associate Professor, Department of Biochemistry, Faculty of Science, University of Yaounde I. PO BOX: 812 Yaounde, Cameroon. e-mail: cteugwa@yahoo.fr*

I. Background

Infectious diseases remain a serious public health concern worldwide [1]. Despite the significant increase in the comprehension of the pathogenesis and management of infectious diseases, they remain one of the causes of mortality and morbidity, particularly in developing countries [2]. The indiscriminate use of antimicrobials and the poor management of infections lead to a new upsurge in the loss of drugs and the increase of resistant pathogenic microorganisms in recent years [3]. Approximately 700,000 people died yearly due to antibiotic resistance, and an estimated 10 million lives may be at risk by 2050 if nothing is done to solve the problem of antimicrobial resistance [4]. This situation increases the frequency of therapeutic failures and leads to economic liability, coupled with the undesired side effects of synthetic antimicrobials which complicate treatment [5].

During infection, highly reactive free radical and oxygen species are produced. This leads to high oxidative stress, which can provoke cancer, auto-immune, degenerative, and cardiovascular diseases [6]. Synthetic antioxidants widely used in cosmetics, food, and therapeutic industries are being restricted due to their carcinogenicity [7]. At the time, the current steroidal and non-steroidal anti-inflammatory drugs present adverse side effects [8]. The need to challenge these problems, coupled with the limited number of drugs, motivates the intensive searches for novel, effective, and affordable medicines from different sources [9].

Herbal products are extensively used in African traditional medicine to manage various illnesses [10]. Natural products from plants have been recognized as a reservoir of novel drugs with possible new mechanisms of action [11-13]. The use of medicinal plants is increasing worldwide, especially in advanced countries where many people rely on plants as primary healthcare modality due to limited access to modern medicine [15, 16].

Dracaena deisteliana belongs to the family of Dracaenaceae, which includes more than 480 species distributed, principally in tropical and sub-tropical

regions [17]. The resin of *D. deisteliana* is used in Arab medicinal tradition to treat diarrhea, fracture, stomach, intestinal fever, and toothache [18]. In Cameroon, *D. deisteliana* leaf is used to treat infertility [19] and typhoid [20]. The stem is used to treat toothache [21]. In Nigeria, this plant is used to treat cough [22]. The pharmacological properties of *D. deisteliana* include the antileishmania, anti molluscoidal, antimalarial, antibacterial, and antifungal activities [20, 23]. The phytochemical studies of *D. deisteliana* lead to the isolation of numerous compounds with biological properties [24, 25].

S. indicus belonging to the family of Poaceae is a perennial grass that grows in dense tufts. It is represented by approximately 45 species that generally grow in tropical and sub-tropical regions all over the world [26]. The sirup of *S. indicus* prepared with fruits is used to fight chronic diarrhea. The astringent bark decoction of *S. indicus* is a medicine against scabies, ulcers, and dysentery. The leaves and bark are used as a febrifuge [27]. However, phytochemical and biological activities of these plants are less or not investigated. Therefore, this study was undertaken to investigate the phytochemical composition, the antibacterial and antifungal, and the radical scavenging properties of *D. deisteliana* and *S. indicus*.

II. Materials and Methods

a) Collection and Identification of Plant Materials

D. deisteliana and *S. indicus* plant materials (Figure 1) were collected at Ngousso-Yaounde in the Centre region of Cameroon in April 2012 and May 2013, respectively. The plant identification was made at the Cameroon National Herbarium by comparison with specimen number 55004/HNC (*Dracaena deisteliana)* and 15719/SRF Cam (*Sporobolus indicus).*

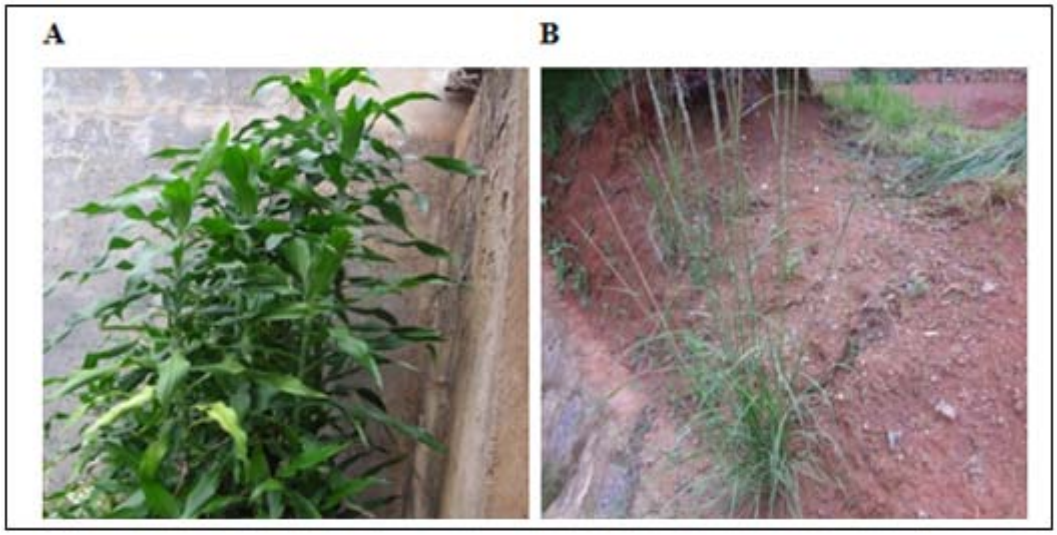

Figure 1: Photographs of selected plants: (A) *Dracaena deisteliana* and (B) *Sporobolus indicus*

b) Preparation of Methanol Extracts

The different parts and the whole plant of *D. deisteliana* and *S. indicus* were air-dried during two weeks at shade at room temperature. The samples were ground separately in a mortar, and 500 g of dried powder of each sample were soaked for 72 h in methanol (1:10 w/v) with constant stirrings. The resulting supernatant was filtered through Whatman no.1 filter paper and concentrated using a rotary evaporator at 55°C. The resultant extracts were transferred into pre-weighed labeled glass vials. The process was repeated twice on the marc to exhaustively extract the plant material. The extraction yield of each plant extract was determined by dividing the total extracted mass by dried plant mass used for extraction. Resultant extracts were air-dried and kept at 4°C for further use.

c) Phytochemical Screening

Qualitative methods were used to determine different classes of phytochemicals (phenolic compound, tannins, saponins, alkaloids, anthocyanins, terpenoids, glycosides, cardiac glycosides, phlobatannins, and flavonoids), as previously described by Trease and Evans [28], and Sofowora [29].

d) Antimicrobial Assays

i. Microbial Strains and Culture Media

Twelve bacteria including ten Gram-negative (*Enterobacter cloacae, Klebsiella oxytoca, Klebsiella pneumoniae, Morganella morganii, Bacillus cereus, Escherichia coli, Proteus vulgaris, Citrobacter freundii, Pseudomonas aeruginosa, Acinetobacter sp)* and two Gram-positive (*Staphylococcus aureus* and *Streptococcus faecalis*) and four yeasts (*Candida albicans*, *Candida krusei, Candida parapsilosis,* and

Cryptococcus neoformans). These clinical isolates were obtained as a donation from the Laboratory of Microbiology, Faculty of Science, University of Yaounde I. All strains were cultured 24 h on Mueller Hinton Agar (MHA) for bacteria and Sabouraud dextrose agar (SDA) for fungi before any test. The Mueller Hinton broth (MHB) and Sabouraud dextrose broth (SDB) were used as liquid medium for the determination of the minimum inhibitory concentrations (MICs) and the minimum bactericidal/fungicidal concentrations (MBCs/MFCs). Gentamicin 1 mg/mL (bacteria) and Fluconazole 100 mg/mL (fungi) were used as positive control.

ii. *Preparation of Microbial Inoculum*

The microbial inoculum was prepared using a direct colony suspension method. Suspensions of bacteria and yeasts were prepared in normal saline from 24 h grown on fresh MHA or SDA at 37°C. The bacterial suspension formed was adjusted with a spectrophotometer to a McFarland standard of 0.5, which is approximately 1.5 x 10^8 CFU/mL. The turbidity of fungal strains was adjusted to a standard of 0.9 to give 1-5 x 10^7 CFU/mL. Each suspension was then diluted 1:100 by transferring 0.1 mL of the bacterial suspension to 9.9 mL of sterile MHB while preparing for experiments [30].

iii. *Agar Well Diffusion Assay for Antibacterial Screening*

The antibacterial activity was performed using the agar well diffusion method according to the modified Kirby Bauer diffusion technique [31]. The agar plates were swabbed with overnight bacterial suspensions of each strain. Then, wells were bored into the agar medium with heat sterilized 6 mm cork borer. A 75 µL of the methanolic extracts (100 mg/mL) was dispensed into the wells, and the plates were left for 30 min before being incubated for 24 h at 37 °C. Each zone of inhibition around the wells was measured using a vernier caliper.

iv. *Determination of the minimum inhibitory concentrations (MICs) and minimum bactericidal/ fungicidal concentrations (MBCs/MFCs) of the plant extracts*

The MICs values of the plant extracts for bacteria and yeasts were determined using serial dilution microplate methods [32, 33]. Two-fold serial dilution of the extract (dissolved in MHB or SDB) was made in a 96-wells microplate for a final concentration ranging from 100 to 97.65 x 10^{-3} mg/mL. An equal volume (100 µL) of the 1.5 x 10^6 CFU/mL bacterial inoculum or 1-5 x 10^5 CFU/mL fungal inoculum prepared in MHB or SDB was then added. The plates were covered with a sterile plate sealer and then incubated for 24 h at 37°C (48 h for fungi). After incubation, 40 µL of 2,3,5-triphenyltetrazolium chloride 0.01 % w/v (TTC) was added in each well of the plates and incubated for 30 min at 37°C. The MIC, defined as the lowest sample concentration that prevented the growth of the bacteria, was then detected by any observed color change. The MBCs/MFCs of each fraction were determined by sub-culturing the sample (50 µL) taken from the wells without growth during MIC determination to 150 µL of MHB or SDB. The plates were incubated at 37°C for 48 h (72 h for fungi). The MBC (or MFC) was regarded as the lowest concentration of extracts with the absence of growth that prevented the color change of the medium after the addition of TTC as mentioned above.

e) Free Radical Scavenging Assay

The DPPH (2.2-diphenyl-1-picrylhydrazyl) radical-scavenging potential of *D. deisteliana* and *S. indicus* extracts was determined following a modified method of Brand-Willians et al. [34]. A 10 µL of each extract prepared in methanol at different concentrations was added into 1990 µL of DPPH solution (0.04 mg/mL) in different tubes for final concentrations of 5 µg/mL; 10 µg/mL; 15 µg/mL; 20 µg/mL; 25 µg/mL; 30 µg/mL. After vortexing, the tubes were kept in the darkness at room temperature for 30 min. The absorbance at 517 nm was taken. The percentage inhibition was calculated from $[(A_0-A_1)/A_0]$ x 100, where A_0 is the absorbance of the control at 30 min (DPPH solution), and A_1 is the absorbance of the extract/reference. Ascorbic acid was used as a reference. The inhibition curves were prepared, and EC_{50} (Efficient concentration of the sample (g) to scavenge 50 % of the DPPH free radical) values were calculated.

f) Statistical Analysis

Data were represented as mean ± standard deviation (SD) of three replicates and subjected to one way analysis of variance (ANOVA) using the Fisher test at the threshold of $p < 0.05$ with Stat graphics plus 5.0 for windows. Linear regression analysis was used to calculate EC_{50} values. Microsoft Excel was used to enter and capture data.

III. Results and Discussion

a) Extraction Yields of the Plant Extracts

The plant material was extracted using methanol as solvent. The highest yield was obtained with *D. deisteliana* leaf extract (8.89 %). The least yield of extraction was obtained with *S. indicus* extract (3.46 %) (Table 1). It has been shown that the type of solvent used in extraction procedure determines the success of isolated compounds from the plant material [35]. The yield of extraction of the stem extract of *D. deisteliana* was 5.63, which is higher than 0.95 previously obtained by Kougan et al. [25].

Table 1: Yield percentage (%) of different extracts of plants used in the study

Plant	Part used	Solvent used	Yield of extraction (%)
D. deisteliana	Leaf	Methanol	8.89
	Stem	Methanol	5.63
	Whole plant	Methanol	6.37
S. indicus	Whole plant	Methanol	3.46

b) Phytochemical Screening

The results of the phytochemical screening carried out with the crude extracts of *D. deisteliana* and *S. indicus* are presented in Table 2. Results showed that all extracts are rich in phenols, saponins, anthraquinones, and alkaloids. Terpenoids and flavonoids were present at a moderate level in the stem and whole plant extracts of *D. deisteliana*, while abundant amount was found in the leaf extract. It has been shown that plants belonging to the genus Dracaena contain steroidal saponins and flavonoids [36, 37]. The phytochemical investigations of *D. deisteliana* leaf extract by Kougan et al. [25] reported the presence of steroidal saponins and saponins. Anthocyanins and tannins were found in the crude extracts of *S. indicus.* In a previous study, it has been reported that *S. indicus* is rich in tannins [27]. Several classes of secondary metabolites found in these plant extracts have been reported to posses' antimicrobial activities [38-40].

Table 2: Phytochemical composition of *D. deisteliana* and *S. indicus* extracts

Plant constituents		*D. deisteliana*			*S. indicus*
		Leaf	Stem	Whole plant	
Phenolic	Ferric chloride test	+	+	+	+
	Potassium dichromate	+	+	+	+
Tannins	Ferric chloride test	-	-	-	+
Anthocyanins	Ammonia HCl test	-	-	-	+
Saponins	Frothing test	+	+	+	+
Flavonoids	Alkaline reagent test	+++	++	++	+
	Lead acetate test	+	+	+	+
Alkaloids	Tannic acid test	+	+	+	+
	Mayer's test	+	+	+	+
	Wagner's test	+	+	+	+
Terpenoids	Salowski test	+++	++	++	+
Anthraquinones	Borntrager's test	+	+	+	+
Glycosides (Free sugar)	Legal's test	-	-	-	-
Cardiac glycosides	Killer Killani test	-	-	-	-
Phlobatanins	Hydrochloride test	-	-	-	-

+++ = abundant; ++ = moderate ; + present ; - completely absent

c) Diameter of Inhibition Zones

The presence of inhibition zones after incubation showed that the Gram-positive, Gram-negative, and fungi isolates exhibited a varied degree of susceptibility to each of the plant extracts that can be considered as a plant with a broad spectrum of activity (Table 3). Considering the susceptibility of the isolates to *D. deisteliana*, the inhibition zones ranged between 8.5 ± 0.28 (*K. pneumoniae*) and 15 ± 0.57 mm (*K. oxytoca* and *E. coli*) for the leaf extract while no activity was noted on *B. cereus* and *P. aeruginosa.* In the previous report, the leaf extract of *D. deisteliana* exhibited the inhibition zones of 8.5 ± 00 mm (80 mg/mL) and 12 ± 00 mm (160 mg/mL) on *S. typhi* [20]. The stem extract

was less active on *K. pneumoniae* with inhibition zone of 11 ± 00 mm and exhibited higher activity on *E. cloacae* with inhibition zone of 18 ± 00 mm. The inhibition zones varied between 9.5 ± 0.28 (*K. pneumoniae*) and 16.5 ± 0.28 mm (*E. cloacae*) for the whole plant extract. This study provides additional data on the antimicrobial activity of *D. deisteliana.* The whole extract of *S. indicus* exhibited inhibition zones varying between 7 ± 00 (*E. coli*) and 14 ± 0.43 mm (*Acinetobacter spp*). These results revealed for the first time information on the antimicrobial properties of *S. indicus.* Nevertheless, the antibacterial activity of both the plant extracts was less pronounced compared to the standard antibiotic (gentamicin) with inhibition zones varying between 21 ± 00 and 29.67 ± 0.88 mm.

Table 3: Inhibition zone (mm) of plant extracts against some bacteria species

Sample / Microorganism	*D. deisteliana*			*S. indicus*	Gentamicin
	Leaf	Stem	Whole plant		
E. cloacae	12 ± 1^{a}	18 ± 0.0^{b}	16.5 ± 0.5^{b}	9 ± 0.0^{c}	28 ± 0.0^{d}
K. pneumoniae	8.5 ± 0.5^{a}	11 ± 0.0^{b}	9.5 ± 0.5^{a}	0 ± 0.0^{c}	30 ± 0.0^{d}
S. faecalis	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	25 ± 0.0^{b}
S.aureus	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	27 ± 0.0^{b}
K. oxytoca	15^{a}	11 ± 1^{b}	14 ± 2^{a}	0 ± 0.0^{c}	27 ± 0.0^{d}
Acinetobacter sp	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	14 ± 075^{b}	23.33 ± 0.57^{c}
E. coli	15 ± 1^{a}	13 ± 1^{a}	18 ± 0.0^{b}	7 ± 0.0^{c}	23.33 ± 0.57^{d}
P. vulgaris	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	8 ± 0.0^{b}	21 ± 0.0^{c}
P. aeruginosa	0 ± 0.0^{a}	12.5 ± 0.5^{b}	11 ± 0.0^{b}	0 ± 0.0^{a}	28 ± 0.0^{c}
M. morganii	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	9 ± 0.0^{b}	29.67 ± 0.57^{c}
C. freundii	0 ± 0.0^{a}	0 ± 0.0^{a}	0 ± 0.0^{a}	7 ± 1^{b}	26.67 ± 1.15^{c}
B. cereus	0 ± 0.0^{a}	12.5 ± 1.5^{b}	14 ± 1^{b}	12 ± 0.0^{b}	23 ± 0.0^{c}

The results are expressed as means ± standard deviation of three determinations. Values with different letters in the same line are significantly different at $p<0.05$.

d) Minimum Inhibitory Concentrations (MICs) and Minimum Bactericidal/Fungicidal Concentrations (MBCs/MFCs)

In this study, the extracts obtained from both the plant extracts displayed varying antimicrobial activities according to their MICs (6.25-50 mg/mL for bacteria and 1.56-50 mg/mL for yeasts) as reported in Table 4. The leaf and the whole plant extracts of *D. deisteliana* were the most active (MIC = 6.25 mg/mL) against *E. cloacae* and *E. coli* (Gram-negative) while both bacteria had the same degree of susceptibility (MIC = 50 mg/mL) to the stem extract. The whole extract of *S. indicus* (MIC = 6.25 mg/mL) had remarkable activity against *B. cereus* (Gram-positive). The sensitivity of Gram-negative and Gram-positive bacteria could be due to the difference in their membrane morphology [41]. The phospholipidic bilayer of the outer membrane of bacteria is the target of interactions with antimicrobial compounds. Damages on the bacterial membrane could increase permeability to ions, the release of intracellular constituents, deterioration of the enzymatic system of bacteria, and even dead [42, 43]. All the tested extracts of *D. deisteliana* were most active on *C. albicans* and *C. krusei* (MIC = 1.56 µg/mL). A similar activity was observed with the leaf and the whole-plant extracts on *C. parapsilosis. C. neofarmans* had the least susceptibility (MIC = 50 µg/mL) to all the extracts of *D. deisteliana.* The antimicrobial activity can be classified as interesting (CMI < 100 µg/mL), moderate (100 < CMI < 625 µg/mL) and weak (CMI > 625 µg/mL) [44, 45]. Therefore, all the plant extracts have weak activity on the tested microorganisms. The weak antibacterial activity exhibited by all the plant extracts could be correlated to the few amounts of secondary metabolites since it has been proven that the concentration, the nature, and the origin of active compounds present in plant extracts may influence the antimicrobial activity [40, 46]. The antimicrobial mechanism of active ingredients may vary with species, chemical composition, cell wall composition, and genetic material of each microorganism [38, 41, 47].

According to Mims et al. [48], the leaves extracts of *D. deisteliana* had a bactericidal effect on *S. aureus, K. pneumoniae, B. cereus,* and *K. oxytoca.* In comparison, the stem extracts had bacteriostatic effect on *S. aureus, K. pneumoniae, E. cloacae,* and *E. coli*. The whole-plant extract of *D. deisteliana* exhibited a bactericidal action on *S. aureus, K. pneumonia,* and *B. cereus.* At the same time, the bacteriostatic effect was observed on *K. oxytoca, E. cloacae,* and *E. coli*. The whole-plant extract of *S. indicus* exhibited a bacteriostatic effect on *K. pneumoniae, E. coli, B. cereus,* and *P. vulgaris* while the bactericidal effect was observed on the rest. All the extracts of *D. deisteliana* have a bactericidal effect on all the yeast strains used in this study.

Table 4: Inhibition parameters (MICs, MBC/MFCs) of methanol extracts from *D. deisteliana* and *S. indicus* (mg/mL) and reference drugs (μg/mL)

	Microorganisms	Parameters	*D. deisteliana*			*S.indicus*	Ref*
			Leaf	Stem	Whole plant		
Bacteria	*S. aureus*	MIC	50	50	50	25	10
		MBC	100	100	100	50	20
		MBC/MIC	2	2	2	2	2
	K. pneumoniae	MIC	50	50	50	12.5	0.39
		MBC	100	100	100	50	0.78
		MBC/MIC	2	2	2	4	2
	Acinetobacter spp	MIC	/	/	/	25	10
		MBC	/	/	/	25	20
		MBC/MIC	/	/	/	1	2
	B. cereus	MIC	25	25	25	6.25	0.19
		MBC	50	100	50	25	0.39
		MBC/MIC	2	4	2	4	2
	K. oxytoca	MIC	25	25	25	25	10
		MBC	50	100	100	25	20
		MBC/MIC	2	4	4	1	2
	S. faecalis	MIC	/	/	/	50	10
		MBC	/	/	/	50	20
		MBC/MIC	/	/	/	1	2
	E. coli	MIC	6.25	12.5	6.25	12.5	3.12
		MBC	50	50	50	50	12.5
		MBC/MIC	8	4	8	4	4
	P. vulgaris	MIC	/	/	/	12.5	10
		MBC	/	/	/	50	20
		MBC/MIC	/	/	/	4	2
	P. aeruginosa	MIC	/	/	/	50	10
		MBC	/	/	/	50	20
		MBC/MIC	/	/	/	1	2
	M. morganii	MIC	/	/	/	50	5
		MBC	/	/	/	50	10
		MBC/MIC	/	/	/	1	2
	C. freundi	MIC	/	/	/	25	10
		MBC	/	/	/	50	20
		MBC/MIC	/	/	/	2	2
	E. cloacae	MIC	6.25	12.5	6.25	25	15.6
		MBC	50	50	50	50	31.2
		MBC/MIC	8	4	8	2	2
Yeasts	*C. albicans*	MIC	1.56	1.56	1.56	/	0.78
		MFC	1.56	1.56	1.56	/	0.78
		MFC/MIC	1	1	1	/	1
	C. krusei	MIC	1.56	1.56	1.56	/	0.39
		MFC	1.56	1.56	1.56	/	0.39
		MFC/MIC	1	1	1	/	1
	C. parapsilosis	MIC	1.56	50	1.56	/	0.39
		MFC	3.12	100	3.12	/	0.39
		MFC/MIC	2	2	2	/	1
	C. neofarmans	MIC	50	50	50	/	0.78
		MFC	50	100	50	/	0.78
		MFC/MIC	1	2	1	/	1

Legend: MIC = Minimal Inhibitory Concentration; MBC/MFC = Minimal Bactericidal/Fungicidal Concentration; Ref* = reference drugs: gentamicin (for bacteria) and fluconazole (for yeasts); / = Not determined

e) Antiradical Properties of Plant Extracts

The free radicals scavenging properties of the plant extracts are reported in Figure 2. The crude extracts of *D. deisteliana* and *S. indicus* exhibited radical scavenging properties in concentration-dependent manners. The inhibition percentages of the stem (26 %) and the whole plant extract of *D. deisteliana* (28 %) and *S. indicus* (24 %) were most pronounced than that of the leaf extract of *D. deisteliana* (14 %) at the concentration of 1 mg/mL. The higher inhibition percentage was observed with the stem extract of *D. deisteliana* (62 %) at the concentration of 5.5 mg/mL. It can be observed that the DPPH activity *D. deisteliana* and *S. indicus* were found to be increasing in concentration-dependent manner.

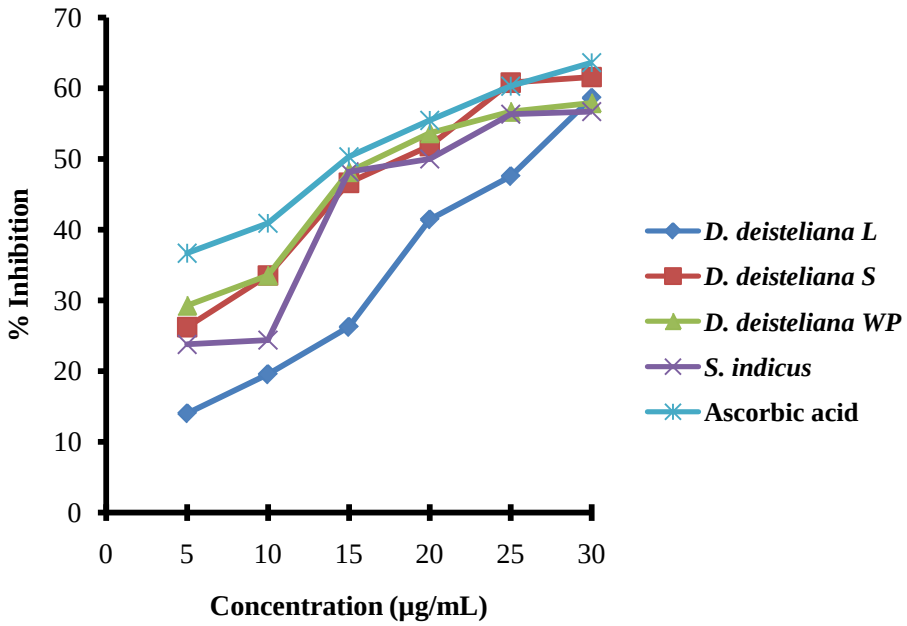

Figure 2: DPPH free radical scavenging activities of plant extracts. L: leaf; S: stem; WP: whole plant.

From each graph, the EC_{50} of each extract was determined. The EC_{50} is the concentration of the samples, which scavenges 50 % of free radicals. Figure 3 shows the scavenging activity of the crude extracts of *D. deisteliana* and *S. indicus* in comparison with that of ascorbic acid. The EC_{50} obtained showed that among the crude extracts of *D. deisteliana,* the leaf extract exhibited the lowest activity (646.75 µg/mL), while the stem extract had the higher one (491 µg/mL). The whole plant extract of *S. indicus* had an EC_{50} of 550.5 µg/mL, while the EC_{50} value of the standard was found to be 411 µg/mL. Numerous previous studies show the correlation between antiradical activity and the phenolic compounds [49]. These studies have confirmed that the phenolic compounds contribute significantly to the antioxidant activity [50]. The antiradical activity depends on the content in phenolic compounds that give up hydrogen to the free radicals and interrupt the chain of lipid oxidative reaction in the first step of inhibition [51]. This higher efficiency of the phenolic compounds to scavenge free radicals like singlet oxygen, superoxide, and hydroxyl radicals is due to their hydroxyl phenolic group [52]. Flavonoids and tannins found in these plant extracts possess a large spectrum of antiradical properties [53]. However, these activities may be due to the synergistic action of the chemical compounds presents in the extracts [54].

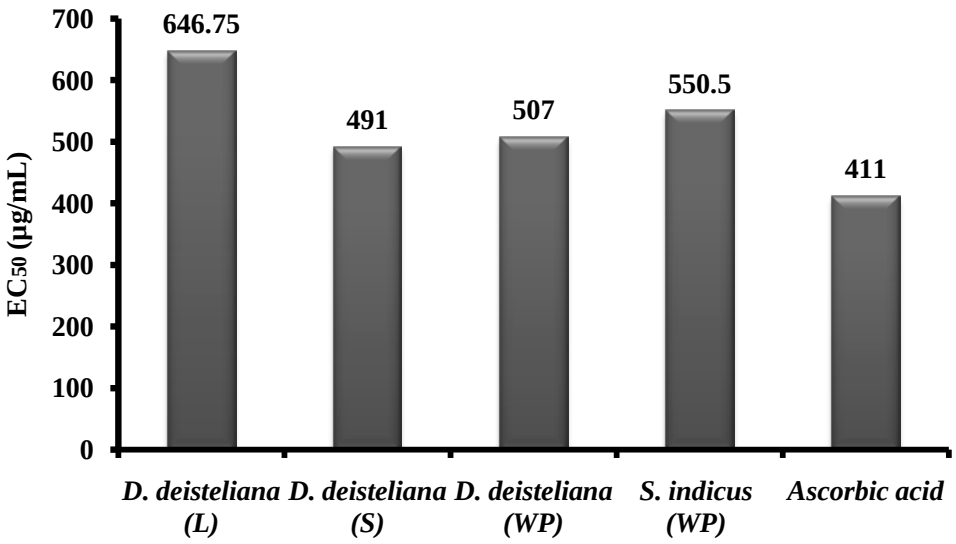

Figure 3: Free radical scavenging activity of plant extracts and reference. L-: leaf- ; S-: stem- ; WP-: whole-plant

IV. Conclusion

Overall, the phytochemical screening of the crude extracts of *D. deisteliana* and *S. indicus* revealed the presence of several classes of secondary metabolites with known antimicrobial and antioxidant activities. The three extracts of *D. deisteliana* have bactericidal activity on *K. pneumoniae* and *S. aureus* and all the fungi tested. *S. indicus* have bactericidal activities on all the bacteria tested. All the plant extracts exhibited weak antibacterial activity. Nevertheless, these results support their traditional uses for the treatment of infections. *D. deisteliana* and *S. indicus* posses' important antiradical activities and can be used to scavenge free radicals. Further studies are still required, namely to isolate active ingredient from these plants to increase their activities and elucidate their potential mechanism of action.

Ethics approval and consent to participate
Not applicable

Consent for publication
Not applicable

Availability of data and material
The datasets used and analyzed during the current study are available from the corresponding authors on reasonable request.

Competing interests
The authors declare that they have no competing interests.

Funding
None

Author's contributions
RNK and CTM designed the study, supervised experiments, and critically revised the paper and intellectual content. ADA carried out the experiments, analyze data, and wrote the manuscript. All authors read and approve the final manuscript.

Acknowledgements

The authors acknowledge the National Herbarium of Cameroon; the Laboratory of Organic Chemistry of the Higher Teachers' Training College and the Laboratory of Microbiology of the Department of Microbiology, University of Yaounde I. Authors are grateful to Prof. Maximilienne Ascension Nyegue for providing facilities.

References Références Referencias

1. Tchinda CF, Sonfack G, Simo IK, et al. Antibacterial and antibiotic-modifying activities of fractions and compounds from *Albizia andianthifolia* against MDR Gram-negative enteric bacteria. BMC Complement Altern Med. 2019; 19: 120. doi: 10.1186/s12906-019-2537-1.
2. Mekonnen B, Asrie AB, Wubneh ZB. Antidiarrheal activity of 80 % methanolic leaf extract of *Justicia schimperiana*. Evid Based Complementary Altern Med. 2018 Article ID 3037120:1-10.
3. Khan MS, Ahmad I, Cameotra SS. Phenyl aldehyde and propanoids exert multiple sites of action towards cell membrane and cell wall targeting ergosterol in *Candida albicans*. AMB Express. 2013; 3:54. doi: 10.1186/2191-0855-3-5
4. Tagliabue A, Rappuoli R. Changing priorities in vaccinology: Antibiotic resistance moving to the top. Front Immunol. 2018;9:1068. doi: 10.3389/fimmu.2018.01068
5. Aslam B, Wang W, Arshad MI, et al. Antibiotic resistance : A rundown of a global crisis. Infect Drug Resist. 2018; 11: 1645-1658. doi: 10.2147/IDR.S173867

6. Mzid M, Ben Khedir S, Ben Salem M, Regaieg W, Rebai T. Antioxidant and antimicrobial activities of ethanol and aqueous extracts from Urtica urens. Pharm Biol. 2017; 55(1):775-81. https://doi.org/10.1080/13880209.2016.1275025
7. Islam MZ, Hossain T, Hossen F, Mukharjee SK, Sultana N, Paul SC. Evaluation of antioxidant and antibacterial activities of *Crotalaria pallida* stem extract. Clin Phytoscience. 2018;4:8. https://doi.org/10.1186/s40816-018-0066-y
8. de Oliveira RG, Mahon CP, Ascêncio PG, Ascêncio SD, Balogun SO, Martins DT. Evaluation of anti-Inflammatory activity of hydroethanolic extract of *Dilodendron Bipinnatum* Radlk. J Ethnopharmacol. 2014;155(1):387-95. doi: 10.1016/j.jep.2014.05.041.
9. Chitemerere, TA, Mukanganyama S. Evaluation of cell membrane integrity as a potential antimicrobial target for plant products. BMC Complement Altern Med. 2014;14:278. doi: 10.1186/1472-6882-14-278.
10. Nyegue MA, Afagnigni AD, Ndam NY, Djova SV, Fonkoua MC, Etoa FX. Toxicity and activity of ethanolic leaf extract of *Paullinia pinnata* Linn (Sapindaceae) in *Shigella flexneri*-induced diarrhea in Wistar rats. J Evid-Based Integ Med. 2020;25:1-9. doi:10.1177/2515690X19900883.
11. Afagnigni AD, Nyegue MA, Djova SV, Etoa FX. LC-MS analysis, 15-lipoxygenase inhibition, cytotoxicity, and genotoxicity of Dissotis multiflora (Sm) Triana (Melastomataceae) and *Paullinia pinnata* Linn (Sapindaceae). J Trop Med. 2020, 1-8. https://doi.org/10.1155/2020/5169847.
12. Kuete V, Fokou WF, Karaosmanoğlu O, Beng VP, Sivas H. Cytotoxicity of the methanol extracts of *Elephantopus mollis, Kalanchoe crenata* and 4 other Cameroonian medicinal plants towards human carcinoma cells. BMC Complement Altern Med. 2017;17:280.
13. Famuyide IM, Aro AO, Fasina FO, Eloff JN, McGaw LJ. Antibacterial and antibiofilm activity of acetone leaf extracts of nine underinvestigated south African Eugenia and Syzygium (Myrtaceae) species and their selectivity indices. *BMC Complement Altern Med*. 2019; 19:141. https://doi.org/10.1186/s12906-019-2547-z
14. Kudumela RG, Mazimbab O, Masoko P. Isolation and characterisation of sesquiterpene lactones from *Schkuhria pinnata* and their antibacterial and anti-inflammatory activities. S Afr J Bot. 2019; 126: 340-344.
15. Odeja O, Ogwuche CE, Elemike EE, Obi G. Phytochemical screening, antioxidant and antimicrobial activities of *Acalypha ciliata* plant. Clin Phytoscience. 2016;2:12. doi 10.1186/s40816-016-0027-2
16. James PB, Wardle J, Steel A, Adams J.¨ Traditional, complementary and alternative medicine use in Sub-Saharan Africa: a systematic review. BMJ Glob Health. 2018;3(5): e000895. doi: 10.1136/bmjgh-2018-000895
17. Mimaki Y, Kuroda M, Takaashi Y, Sashida Y. Steroidal saponins from the stems of *Dracaena concinna.* Phytochemistry. 1998;47(7):1351-1356.
18. Yung L, Hui W, Huerong X, Weuli M, Haofu D. Antioxidant phenolic compounds of *Dracaena cambodiana.* Molecules. 2010;15: 8904-8914
19. Telefo PB, Lienou LL, Yemele MD, Lemfack MC, Mouokeu C, Goka CS, Tagne SR, Moundipa FP. Ethnopharmacological survey of plants used for the treatment of female infertility in Baham, Cameroon. *J Ethnopharmacol.* 2011;136(1):178-187. doi: 10.1016/j.jep.2011.04.036.
20. Tsobou R, Mapongmetsem PM, Voukeng KI, Van Damme P. Phytochemical screening and antibacterial activity of medicinal plants used to treat typhoid fever in Bamboutos division, West Cameroon. J App Pharm Sci. 2015;5(06):34-49. doi: 10.7324/JAPS.2015.50606
21. Jiofack T, Ayissi I, Fokunang C, Guedje N, Kemeuze V. Ethnobotany and phytomedicine of the upper Nyong valley forest in Cameroon. Afr J Pharm Pharmacol. 2009;3(4):144-150.
22. Adjanohoun EJ, Aboubakar N, Dramane K, et al. Traditional medicine and pharmacopoeia. Contribution to ethnobotanical and floristic studies in Cameroon. Organisation of African unity. Scientific, Technical and research commission (OAU/STRC) Lagos, Nigeria. 1996.
23. Okunji CO, IWU MM, Jackson JE, Tally JD. Biological activity of saponins from two Draceana species. Adv Exp Med Biol. 1996;28:404-415.
24. He L, Wang Z, Tu P, Hou H. Advances in study on chemical constituents and pharmacological activities in plants of Dracaena vand. Ex L. Chin Tradit Herb drugs. 2004;35:221-228.
25. Kougan GB, Miyamoto TT, Paululat T, Mirjolet JF, Duchamp O, Sondengam BL, Lacaille-Dubois MA. Steroidal saponins from two species of Dracaena. J Nat Prod. 2010;73(7):1266-1270. doi: 10.1021/np100153m
26. Peterson PM., Webster RD, Valdes-Reyna J. Genera of New World Eragrostideae (Poaceae: Chloridoideae). Smithson contrib bot. 1997;1-54. https://doi.org/10.5479/si.0081024X.87
27. Global Invasive Species Database. Species profile: *Sporobolus africanus*. http://www.iucngisd.org/gisd/species.php?sc=1587. Accessed 24/06/2020.
28. Trease G.E., Evans W.C. Pharmacognosy. 15th Ed. London: Saunders Publishers; 2002.
29. Sofowora A. Screening plants for bioactive agents. In: Medicinal plants and traditional medicinal in Africa. 2nd ed. Ibadan: Spectrum Books Ltd, Sunshine House. 1993; 134-156.

30. Dzoyem JP, Guru SK, Pieme CA, Kuete V, Sharma A, Khan IA, Saxena AK, Vishwakarma RA. Cytotoxic and antimicrobial activity of selected Cameroonian edible plants. BMC Complement Altern Med. 2013;13:78. doi: 10.1186/1472-6882-13-78
31. Cheesbrough M. Medical Laboratory Manual for Tropical Countries, vol 2 of ELBS Tropical Health Technology, Butterworth-Heinemann, Cambridge, UK. 2002.
32. Eloff JN. A sensitive and quick microplate method to determine the minimum inhibitory concentration of plant extracts for bacteria. Planta Med. 1998; 64:711-713.
33. Masoko P, Picard J, Eloff JN. Antifungal activities of six South African *Terminalia* species (Combretaceae). J Ethnopharmacol. 2005; 99(2): 301-308.
34. Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. Lebensmittel-Wissenschaft und Technologie. 1995; 28: 25-30.
35. Masoko P, Makgapeetja DM. Antibacterial, antifungal and antioxidant activity of *Olea africana* against pathogenic yeast and nosocomial pathogens. BMC Complement Altern Med. 2015; 15:409. doi 10.1186/s12906-015-0941-8
36. Tapondjou LA, Ponou KB, Teponno RB, Mbiantcha M, Djoukeng JD, Nguelefack TB, Watcho P, Cadenas AG, Park HJ. In vivo anti-inflammatory effect of a new steroidal saponin, mannioside A, and its derivatives isolated from *Dracaena mannii*. Arch Pharm Res. 2008;31:653-658
37. Moharram FA, El-Shenawy SM. Antinociceptive and anti-inflammatory steroidal saponins from *Dracaena ombet*. Planta Med. 2007;73(10):1101-1106.
38. Cowan MM. Plants products as antimicrobial agents. Clin Microbiol Rev. 1999; 14:564-584.
39. Olufunmiso OO, Anthony JA. Synergistic interactions of methanolic extract of *Acacia mearnsii* De wild. with antibiotics against bacteria of clinical relevance. Int J Mol Sci. 2012;13:8915-8932.
40. Dzotam JK, Touani FK, Kuete V. Antibacterial activities of the methanol extracts of *Canarium schweinfurthii* and four other Cameroonian dietary plants against multi-drug resistant Gram-negative bacteria. Saudi J Biol Sci. 2016; 23:565-570.
41. Masoko P, Gololo SS, Mokgotho MP, Eloff JN, Howard RL, Mampuru LJ. Evaluation of the antioxidant, antibacterial and antiproliferatory activities of the acetone extracts of the roots of *Senna italica* (Fabaceae). Afri J Trad Compl Altern Med. 2010; 7(2):138-48.
42. Zhao W.H., Hu Z.O., Okubo S., Hara Y., Shimamura T. Mechanism of synergy between epigallocatechin gallate and blactams against methicillin-resistant *Staphylococcus aureus*. Antimicrob Agents Chemother. 2001; 45(6):1737-1742.
43. Perumalla AV, Navam S. Green tea and grape seed extracts-potential applications in food safety and quality. Food Res Intern. 2011;44(4):827-839.
44. Kuete V. Potential of Cameroonian plants and derived products against microbial infections: A review. Planta med. 2010; 76:1479-1491.
45. Kuete V, Efferth T. Cameroonian medicinal plants: pharmacology and derived natural products. Front Pharmacol. 2010; 1:123. doi: 10.3389/fphar.2010.00123.
46. Takeo O, Masato K, Keiko S, Rika O, Junko M, Hiroshi I, Hiroyuki K, Toshi A, Tosshifumi A, Shigeo M. *In vitro* and *in vivo* antimicrobial activities of tricyclic ketolide Te-802 and its analogs. J Antibiotics. 2004; 57: 518-527.
47. Mims CA, Playfair JH, Roitt IM, Wakelin D, Williams R. Antimicrobials and chemotherapy. In: Mims et al. Editors. Medical Microbiology Review, Mosby Europe Ltd, London. 1993; 35:1-34.
48. Tekwu EM, Pieme AC, Beng VP. Investigations of antimicrobial activity of some Cameroonian medicinal plant extracts against bacteria and yeast with gastrointestinal relevance. J Ethnopharmacol. 2012; 142:265-273.
49. Mukherjee S, Pawar N, Kulkarni O, Nagarkar B, Thopte S, Bhujbal A, Pawar P. Evaluation of free-radical quenching properties of standard Ayurvedic formulation Vayasthapana Rasayana. BMC Complement Altern Med. 2011; 11:38.
50. Li HB, Wong CC, Cheng KW, Chen F. Antioxidant properties in vitro and total phenolic contents in methanolic extracts from medicinal plants. LST Food Sci Technol. 2008; 41:0385-390.
51. Kaushik R, Narayanan P, Vasudevan V, Muthukumaran G, Antony U. Nutrient composition of cultivated stevia leaves and the influence of polyphenols and plant pigments on sensory and antioxidant properties of leaf extracts. J Food Sci Technol. 2010;47:27-33.
52. Sawa T, Nakao M, Akaike T, Ono K, Maeda H. Alkylperoxyl radical scavenging activity of various flavonoids and other phenolic compounds: Implications for the antitumor promoter effect of vegetables. J Agric Food Chem.1999; 47:397-492.
53. Mohammed F, Nagendra PK, Kong KW, Amin I. Flavonoid, hesperidine, total phenolic contents and antioxidant activities from Citrus species. Afr J Biotechnol. 2010; 9:326-330.
54. Ali G, Hawa ZE, Jaafar, Asmah R. Antioxidant Activities, Total phenolics and flavonoids content in two varieties of Malaysia young Ginger (*Zingiber officinale* Roscoe). Molecules. 2010; 15(6): 4324-4333.