



## Role of Interleukin-5 in the Pathophysiology and Treatment of Eosinophilic Asthma

By Nightingale Syabbalo

*Medicine Copperbelt University*

**Abstract-** Asthma is a heterogeneous chronic airway disease with several distinct phenotypes characterized by different immune pathological pathways, clinical features, disease severity, physiology, and response to treatment. Approximately 50% of patients with stable chronic asthma have the eosinophilic phenotype, whereas the remainder have the non-eosinophilic asthma. Eosinophilic asthma is the most common phenotype in children with acute severe asthma, but neutrophilic asthma is the most common in adult patients presenting with acute severe asthma. T helper 2 (Th2) cytokines, such as interleukin-5 (IL-5), IL-4, IL-13, IL-25, IL-33, and thymic stromal lymphopoietin (TSLP), play a very important role in the pathophysiology of eosinophilic asthma. Interleukin-5 is the pivotal cytokine responsible for the proliferation, differentiation, activation, and survival of eosinophils; and promotion of eosinophil migration and airway eosinophilia. Activated eosinophils secrete granular cytotoxic cationic proteins, such as major basic protein, eosinophil cationic protein, eosinophil-derived neurotoxin, eosinophil-derived peroxidase, and reactive oxygen species.

**Keywords:** eosinophilic asthma, cytokines, interleukin-5, monoclonal antibodies.

**GJMR-F Classification:** NLMC Code: WF 140



*Strictly as per the compliance and regulations of:*



RESEARCH | DIVERSITY | ETHICS

# Role of Interleukin-5 in the Pathophysiology and Treatment of Eosinophilic Asthma

Nightingale Syabbalo

**Abstract-** Asthma is a heterogeneous chronic airway disease with several distinct phenotypes characterized by different immune pathological pathways, clinical features, disease severity, physiology, and response to treatment. Approximately 50% of patients with stable chronic asthma have the eosinophilic phenotype, whereas the remainder have the non-eosinophilic asthma. Eosinophilic asthma is the most common phenotype in children with acute severe asthma, but neutrophilic asthma is the most common in adult patients presenting with acute severe asthma. T helper 2 (Th2) cytokines, such as interleukin-5 (IL-5), IL-4, IL-13, IL-25, IL-33, and thymic stromal lymphopoietin (TSLP), play a very important role in the pathophysiology of eosinophilic asthma. Interleukin-5 is the pivotal cytokine responsible for the proliferation, differentiation, activation, and survival of eosinophils; and promotion of eosinophil migration and airway eosinophilia. Activated eosinophils secrete granular cytotoxic cationic proteins, such as major basic protein, eosinophil cationic protein, eosinophil-derived neurotoxin, eosinophil-derived peroxidase, and reactive oxygen species. The cytotoxic cationic proteins lead to epithelial injury, inflammation and airway remodeling. Additionally, eosinophils can synthesize and secrete a plethora of pro-inflammatory mediators, such as lipid-derived mediators, cytokines, chemokines, and growth factors, which orchestrate eosinophilic airway inflammation, and airway hyper responsiveness. IL-5 is of paramount importance in eosinophil immunopathological effects. Pharmacologic blockade of IL-5 or its receptor ( $\alpha$ ) has yielded the discovery of biologics, such as mepolizumab, reslizumab, and benralimab, which are useful for the treatment of corticosteroid-resistant eosinophilic asthma.

**Keywords:** eosinophilic asthma, cytokines, interleukin-5, monoclonal antibodies.

## I. INTRODUCTION

Asthma is a significant public health problem, affecting more than 358 million individuals globally,<sup>1</sup> and its prevalence has been increasing during the last 40 years.<sup>1-3</sup> It is the most common chronic respiratory disease in children in developed countries,<sup>4</sup> and its prevalence is steadily increasing in the developing world.<sup>5</sup>

Asthma is a chronic inflammatory airway disease with several distinct phenotypes, characterized by different immunopathological pathways, clinical presentation, severity of the disease, and response to treatment.<sup>6-11</sup> There are several clinical, molecular, and immunogenetic phenotypes of asthma,<sup>12,13</sup> but the

phenotypes of asthma can be simply classified into four phenotypes using induced sputum cytometry.<sup>7,14</sup> The phenotypes of asthma include eosinophilic, neutrophilic, mixed granulocytic, and paucigranulocytic asthma.<sup>7</sup>

Eosinophilic asthma has elevated sputum eosinophil count  $\geq 3\%$ ,<sup>7,14-16</sup> whereas neutrophilic asthma has elevated sputum neutrophil count between  $\geq 61\%$ ,<sup>15</sup> and  $\geq 64\%$ ,<sup>16</sup> depending on the study. Mixed granulocytic phenotype is characterized by increase in both sputum eosinophils ( $>3\%$ ), and neutrophils ( $>61\%$  or  $>64\%$ ).<sup>15</sup> Paucigranulocytic phenotype includes patients with very few eosinophils ( $<3\%$ ), and neutrophils ( $<61\%$  or  $<64\%$ ) in induced sputum.<sup>7,15,17</sup> Non-eosinophilic asthma is the term used to classify patients with low eosinophil numbers ( $<3\%$ ), which include neutrophilic asthma, and paucigranulocytic phenotype.<sup>7</sup>

Eosinophilic airway inflammation play a key role in the pathophysiology of eosinophilic asthma. Activated eosinophils secrete granular cytotoxic cationic proteins, such as major basic protein, eosinophil cationic protein, eosinophil-derived neurotoxin, eosinophil-derived peroxidase, and reactive oxygen species. The cytotoxic cationic proteins lead to epithelial injury, inflammation and airway remodeling. Additionally, eosinophils can synthesize and secrete a plethora of inflammatory mediators, such as leukotrienes, prostaglandins, cytokines, chemokines, and growth factors, which orchestrate eosinophilic airway inflammation, and airway hyper responsiveness (AHR).

Eosinophilic asthma is regarded as T helper 2 (Th2)-driven phenotype. Th2 cytokines, such as interleukin-5 (IL-5), IL-4, IL-13, IL-25, IL-33, and thymic stromal lymphopoietin (TSLP), play a very important role in the pathophysiology of eosinophilic asthma. Interleukin-5 plays a pivotal role in the proliferation, differentiation, migration, activation, and survival of eosinophils. Together with IL-4, IL-13, IL-25, IL-33, and TSLP, it is responsible for airway eosinophilic inflammation, sub epithelial basement membrane fibrosis, extracellular matrix protein deposition, and airway smooth muscle (ASM) cell hyperplasia and hypertrophy.

Most patients with stable eosinophilic asthma are responsive to the step-wise guideline therapies, including high dose inhaled corticosteroids (ICS), long-acting  $\beta$ -2 agonists (LABA), and leukotriene receptor antagonist (LTRA).<sup>1,18,19</sup> However, there is a sub-group of

**Author:** MB., ChB., PhD., FCCP., FRS Professor of Physiology and Medicine Copperbelt University M. C. Sata School of Medicine Kitwe Zambia. e-mail: nightesyab@gmail.com

severe asthma patients with increased airway allergic inflammation despite treatment with high-dose ICS or oral corticosteroids (OCS), and their inflammatory biomarkers remain high even after corticosteroid injections.<sup>20</sup> This sub-group of patients with eosinophilic and steroid-resistant asthma require alternative targeted therapies such as interleukin antagonists (ILA), and bronchial thermoplasty.

## II. EOSINOPHILS

Eosinophils play a pivotal role in the pathogenesis and severity of asthma, and other allergic diseases such as allergic rhinitis, atopic eczema,<sup>21</sup> and eosinophilic granulomatosis with polyangiitis.<sup>22</sup> The eosinophil was first described by Paul Ehrlich in 1879, after he developed the fluorescent dye eosin which coloured basic protein bright red.<sup>23</sup> Hematologic ally, it was identified as eosinophil in an autopsy of a 48-year-old male patient who died of status asthmaticus by Dr. Fraenkel in 1900.<sup>24</sup> In 1953, Houston, et al.<sup>25</sup> demonstrated that patients dying from status asthmaticus had airway mucosa infiltration by activated eosinophils. Thereafter, Bousquet and colleagues,<sup>26</sup> reported that patient with chronic asthma had an increase in eosinophils in peripheral blood, bronchoalveolar lavage (BAL) fluid, and lung biopsy specimens.

Eosinophils are polymorphonuclear cells, with a diameter of about 10-16  $\mu\text{m}$ , and constitutes 1-4% of circulating white blood cells. They have a nucleus usually with two lobes, and have large cytoplasmic granules that stain beautifully deeply red after staining with eosin, using the Romanowsky method.<sup>27</sup> They have a very short life span of about 1-6 hr in the circulation, but can live longer in allergic inflamed tissues, such as lungs and airways.

Eosinophils and other leukocytes are formed from bone marrow CFU-GM progenitor cells during myelopoiesis.<sup>28</sup> The pluripotent myeloid progenitor cells give rise to CD34+ IL-5R $\alpha$  eosinophil progenitor cells, which by the actions of haematopoietic factors, growth factors, and cytokines lead to eosinophils maturation.<sup>28,29</sup> The differentiation of eosinophils is regulated by transcription factors GATA-binding protein 1 (GATA-1), PU.1, and the CCAAT-enhancing binding protein (c/EBP) family.<sup>30</sup> GATA-1 seems to have the most important role, because disruption of GATA1 gene in mice results in a strain completely without eosinophils.<sup>29</sup> Interleukin-4 is essential, because of its requirement to the Th2 cell commitment, and activation via stimulation of key transcription factors, such as GATA3 and STAT6.<sup>31</sup> Interleukin-5, IL-3, and GM-CSF synergistically contribute to the development of mature eosinophils, and other leucocytes, through induction of bcl-xl expression.<sup>29</sup> Interleukin-5 is the most specific and central cytokine in eosinophils, and basophils biology. It

plays a key role in eosinophilic proliferation, differentiation, and the release of eosinophils from the bone marrow into the blood stream, acting synergistically with eotaxin-1, -2, and -3.<sup>31</sup> Eotaxin-1 (CCL11) is a specific chemo attractant of eosinophils and stimulates migration of CD34+ progenitor cells, and release of eosinophils into the peripheral blood, and accumulation in the lungs.<sup>32,33</sup> Eotaxin-1 and its receptor CCR3, may be involved in the survival and other immunological functions of eosinophils.<sup>31</sup>

Migration of eosinophils from the vasculature into lung tissue is facilitated by eosinophils-specific adhesion molecules, such as  $\beta$ 1 integrin very late antigen (VLA-4), the vascular cell adhesion molecule (VCAM-1), and the P-selectin glycoprotein ligand (PSGL-1).<sup>34</sup> VLA-4 is an integrin which is expressed on the membrane of eosinophils after stimulation by eotaxin-1. It ligands with the VCAM-1 integrin expressed on the vascular membrane, resulting in the activation and firm adhesion to eosinophils. Thus, facilitating eosinophil diapedesis through the endothelium into lung tissues.<sup>34</sup>

Recruitment and migration of eosinophils into the airway mucosa is mediated by coordinated action of cytokines, such as IL-5, IL-4, and IL-13, and chemokines, eotaxins-1 (CCL11), eotaxin-2 (CCL24), and eotaxin-3 (CCL26), RANTES, and MCP1/4.<sup>34-36</sup> In particular, eotaxin-1 and its receptor CCR3, play an important role in driving eosinophils into allergic inflamed tissues in the airways. Leukotriene C4 (LTC4) is one of the most potent eosinophil chemo attractant, and is also involved in eosinophil recruitment and cascade. Table 3 summarizes the physiological functions of IL-5 in eosinophil immunobiology.

Eosinophils exhibit chemotaxis, and diapedesis, but they are weak phagocytes. They appear to be used selectively for combating helminth parasitic infections,<sup>37,38</sup> such as *Strongyloides stercoralis*; *Schistosoma haematobium* and *S. mansoni*; *Taenia saginata*; and *Diphyllobothrium latum*. However, they also play an important immunological role in viral infections, especially respiratory viral infections.<sup>39</sup>

## III. EOSINOPHIL SURFACE RECEPTORS

Eosinophils possess a wide repertoire of surface adhesion molecules and receptors, for cytokines, chemokines, and growth factor receptors, lipid mediators receptors, chemo attractant receptors, adhesion receptors, Toll-like receptors, and Fc $\epsilon$ R1 receptors.<sup>40-44</sup> Eosinophilic signaling and inflammatory activity is regulated via cytokines, chemokines, leukotrienes and prostaglandins secreted by Th2 cells, ILC2s, mast cells, and basophils through their respective receptors on the surface of eosinophils.

Eosinophils express the Fc $\epsilon$ R1 receptor for immunoglobulins (Ig), IgE, IgG, IgA, IgM, and IgD, which serves a key role in allergic inflammatory responses.

During allergen exposure, the high-affinity FcεR1 tetramer (αβγγ) receptor on the surface of eosinophils interact with the Fc portion of the IgE molecule leading to activation and degranulation of eosinophils, and release of cationic proteins and other mediators.<sup>44</sup> The most important cytokine receptor on the surface of eosinophils is the heterodimer IL-5 receptor, which plays a key role in eosinophil immunopathology.<sup>44,45</sup> Eosinophils also express other cytokine receptors, including IL-4, IL-13, IL-17, IL-25, IL-33, and TSLP, and growth factor receptors, such as transforming growth factor-β. The above cytokines play a synergistic role in the pathophysiology of eosinophilic asthma.

#### IV. EOSINOPHIL MEDIATORS

Activated eosinophils either by allergic and non-allergic pathways, undergo autolysis and release an array of eosinophil-specific granules found in the extracellular DNA traps.<sup>46</sup> The most predominant bio-active mediators released from the granules are the four cytotoxic cationic proteins, such as major basic protein (MBP), eosinophil cationic protein (ECP), eosinophil-derived neurotoxin (EDN), eosinophil-derived peroxidase (EDPX), and reactive oxygen species.<sup>44,44,47,48</sup> Major basic protein, ECP, and EDPX are toxic to a number of cells, including airway epithelial cells,<sup>49</sup> and ASM cells, and contribute to airway hyper responsiveness (AHR).<sup>50</sup> Eosinophil-derived neurotoxin, and ECP belong to the RNAase A family of granule proteins that have ribonuclease activity.<sup>51</sup> They are associated with host defense against viruses, and may play a role in tissue remodeling.<sup>51</sup> Eosinophil-derived neurotoxin is toxic to nerves,<sup>52</sup> whereas eosinophil peroxidase produce reactive oxygen species, and reactive nitrogen intermediates, which promote oxidative stress in tissue, causing cell death by apoptosis and necrosis.<sup>49</sup>

Eosinophils can synthesize and release a plethora of inflammatory mediators, including lipid-derived mediators, such as histamine, cysteinyl leukotrienes, prostaglandins, thromboxanes, and PAF; cytokines, chemokine, enzymes, and growth factors.<sup>53,54</sup> Leukotrienes and prostaglandins promote airway smooth muscle contraction, mucus secretion, vasodilatation and edema formation, which lead to airway obstruction.<sup>54</sup> They also activate mast cells and basophils through their receptors to secrete more histamine, prostaglandins, and leukotienes, thus amplifying the eosinophilic inflammatory responses.<sup>55</sup>

Additionally, eosinophils synthesize and secrete several Th2 and ILC2 cytokines, such as IL-3, IL-4, IL-5, IL-9, IL-13, IL-15, IL-23, IL-25, IL-33, GM-CSF, TNF-α, and GM-CSF. Interleukin-5, IL-13, and GM-CSF secreted by eosinophils promote eosinophilic function and survival in an autocrine fashion.<sup>53</sup> Interleukin-13, and IL-25 are pro-fibrotic cytokines which lead to subepithelial basement membrane fibrosis, deposition

of extracellular matric proteins, airway remodeling, and fixed airflow limitation. Th2 cytokines, such as IL-5, IL-4, IL-13, IL-25, IL-33, and TSLP are responsible for airway eosinophilia, AHR, and remodeling. They promote airway smooth muscle (ASM) cell proliferation and hypertrophy, amplify ASM cell contraction, and lead to sub mucous glands and goblet cell hyperplasia and mucus hyper secretion. Eosinophils also secrete growth factors, such as TGF-β, VEGF, IL-13, and enzymes, including MMP-9, and TIMP-1 which are responsible for the development of ASM hypertrophy and sub epithelial fibrosis, hence, severe fixed airflow obstruction,<sup>57</sup> and corticosteroid resistance. Table 1 shows the list of pro-inflammatory mediators synthesized and secreted by activated eosinophils.

#### V. CYTOKINES

Pro-inflammatory cytokines play a central role in the pathophysiology of allergic diseases, such as eosinophilic asthma.<sup>58</sup> The cytokines implicated in the pathophysiology of eosinophilic asthma are derived mainly from Th cells.<sup>58</sup> The other sources of cytokines include, mast cells, basophils, eosinophils,<sup>45,59,60</sup> dendritic cells, natural killer cells, and NK T cells.<sup>61-64</sup> Novel T cells, such as ILC2, Th9, Th17, Th22 cells, Treg cells, and nuocytes,<sup>64,65</sup> and structural cells including epithelial cells, fibroblasts and airway smooth muscle cells can also produce cytokines and chemokines.<sup>61</sup>

Notably, there is cross-talk between eosinophils and mast cells, and the cytokines and chemokines networks in orchestration the allergic inflammatory responses.<sup>60,66</sup> Each of these cells produce an array of cytokines and chemokines which promote secretion of more cytokines by the other inflammatory cells, thus establishing paracrine and even autocrine positive feedback loops.

Th2 lymphocyte, ILC2s, mast cell, and eosinophil cytokines possess overlapping biological activities; they can synergize or antagonize the effects of other cytokines. For example, IL-5, IL-4, IL-13, IL-25, and IL-33 are the key drivers of the inflammatory process in eosinophilic asthma; and IL-4 and IL-13 are central Th2 cytokines with distinct overlapping roles, particularly in airway remodeling and bronchial hyperresponsiveness.<sup>58</sup> Similarly, interferon-γ, a Th1 cytokine acts in conjunction with Th2 cytokines (IL-3, IL-4, and IL-5) in maintaining chronic airway inflammation in patients with asthma. The most important cytokines in the pathophysiology of eosinophilic asthma include IL-5, IL-4 IL-13, IL-25, IL-33, and TSLP. IL-5 is the regarded as the master minder cytokine.

#### VI. INTERLEUKIN-5

Interleukin-5 is mainly produced and secreted by Th2 lymphocyte and group 2 innate lymphoid cells (ILC2).<sup>67-71</sup> Th2 cells, and ILC2 secrete IL-5 after been



activated by dendritic cells in response to allergens, and inflammatory mediators.<sup>72</sup> Interleukin-4 is essential for the promotion of Th2 cell differentiation from naive T helper cells (Th0), and activation of Th2 cells leading to the production and release of cytokines, such as IL-4, IL-5, IL-13, IL-25, IL-33, and TSLP.<sup>62</sup> The differentiation of Th2 cells is transcribed by GATA-3 acting as a master signaling factor,<sup>62,73</sup> and STAT6 serving as a key transcription factor.<sup>73</sup> IL-5 secretion from ILC2 is also dependent on GATA-3 activation induced by epithelial "alarmin" cytokines, such as IL-25, IL-33, and TSLP.<sup>73</sup>

Interleukin-5 is a cytokine composed of 134-amino acid proteins that form a 52-kDa homodimer.<sup>74-76</sup> It belongs to the haematopoietic growth factor cytokine family, which also include IL-3 and GM-CSF.<sup>74</sup> It is highly specific for eosinophil formation,<sup>77</sup> by stimulating the production, proliferation, and differentiation of eosinophils from myeloid progenitor cells in the bone marrow.<sup>78,79</sup> IL-5 also aids in the extrusion of eosinophils from the marrow. Peripherally, IL-5 participates in the terminal maturation of the eosinophil in the circulation. Interleukin-5 is important in the recruitment and activation of eosinophils in the lungs, and for eosinophil survival by preventing apoptosis. It plays a critical role in diapedesis of eosinophils by facilitating endothelial adhesion, and promotes chemotaxis in inflamed lung tissues.<sup>80</sup>

The interleukin-5 receptor is a heterodimer composed of a specific subunit, IL-5R $\alpha$ , and a separate motif for binding to the signaling subunit,  $\beta$ c, of the receptor.<sup>76,79</sup> The IL-5R $\alpha$  is specific to IL-5 binding, whereas the  $\beta$ c chain also binds to IL-3, and GM-CSF.<sup>76,79</sup> The IL-5R $\alpha$  subunit is expressed about threefold on eosinophils compared with basophils.<sup>80,81</sup>

Binding of IL-5 to the IL-5 receptor triggers activation of a complex intracellular signaling involving JAK1/2 and STAT1/3/5 modules, p38 and ERK MAP kinases, and NF- $\kappa$ B transcription factor.<sup>82</sup> JAK2, and Lyn and Raf-1 are involved in eosinophil survival by preventing apoptosis, and Raf-1 is specifically involved in stimulating eosinophil activation and degranulation.<sup>82,83</sup> Another IL-5 signaling pathway include activation of intracellular kinases, such as phosphoinositide 3-kinase (PI3K), and mitogen-activated protein kinases (MAPK).<sup>83,84</sup> Through NF-dependent mechanism, p38 MAPK up-regulates eosinophil recruitment into allergic airways, and activates synthesis of pro-inflammatory mediators, including cytokines, chemokines, and leukotrienes, and prostaglandins.<sup>85,86</sup> These mediators orchestrates airway eosinophilic inflammation, subepithelial reticular membrane fibrosis, submucous gland hyperplasia and mucus secretion, and ASM cell proliferation, hyperplasia and hypertrophy.

Interleukin-5 and its receptors (IL-5R $\alpha$ , CD125) expressed on the surface of eosinophils, basophils, and a subset of mast cells are the central players

responsible for airway eosinophilia. Therefore, targeting IL-5 or its receptor subunit IL-5R $\alpha$  is a logical approach for add-on treatment of severe difficult-to-treat eosinophilic asthma, and corticosteroid-resistant asthma phenotypes.<sup>87-89</sup> There are currently two marketed IL-5 monoclonal antibodies (mAb) targeted against IL-5 (mepolizumab, and reslizumab), and one mAb targeted against IL-5R $\alpha$  (benralizumab). Interleukin-5 antagonists bind to distinct epitopes of IL-5 interfering its binding to IL-5 receptors expressed on the surface of eosinophils. Anti-IL-5R antibodies also induce targeted-cell lysis and have been shown to reduce circulating eosinophil counts rapidly.

## VII. MEPOLIZUMAB

Mepolizumab (Nucala®) is an N-glycosylated IgG1/k humanized monoclonal antibody formed by two light chains and two heavy chains bound by a disulphide bond, with a molecular weight of 149.2 kDa.<sup>90</sup> Mepolizumab binds to the  $\alpha$ -chain of IL-5 with both specificity (IC50 <1nm), and affinity (Kd = 4.2 pM),<sup>90</sup> with a dissociation constant of 100 pM, thus preventing it from binding to the  $\alpha$  subunit of the IL-5 receptor expressed on the surface of the eosinophil.<sup>90-92</sup> This results in inhibition of IL-5 signaling and bioactivity which lead to reduction in the production, differentiation, activation and survival of eosinophils. Mepolizumab inhibit eosinophilic activation and the release of myriad of inflammatory mediators from the eosinophils, thus preventing airway eosinophilic inflammation.<sup>92-94</sup>

Mepolizumab (SB-240563, GlaxoSmithKline) was the first biological anti-IL-5 agent to be tested in randomized clinical trials (RCT) in 2000.<sup>95</sup> The first clinical trial of mepolizumab in patients with asthma showed a reduction in sputum and blood eosinophil count but no change in bronchial hyper responsiveness, and no effect on the late asthmatic response.<sup>95</sup> In the phase 2b/3 DREAM (Dose Ranging Efficacy And safety with Mepolizumab in severe asthma) much larger population trial, Pavord et al.<sup>96</sup> confirmed that mepolizumab reduced sputum and blood eosinophil counts, and also significantly reduced asthma exacerbation rates. Additionally, mepolizumab improved the asthma control questionnaire (ACQ) scores, and the asthma quality of life questionnaire (AQLQ) scores.<sup>96</sup>

In the MENSA (MEpolizumab as adjunctive therapy in patients with Severe Asthma) study, Ortega and colleagues,<sup>97</sup> showed that treatment with intravenous (IV) or subcutaneous (SC) mepolizumab decreased the rate of exacerbations by 47% and 53% respectively. It also reduced exacerbations requiring emergency room visits or hospitalization by 32% for IV and 61% for SC mepolizumab. In addition, patients in both IV and SC mepolizumab groups showed significant improvement in the quality of life, and asthma control as assessed by the St. George's Respiratory Questionnaire (SGRQ), and the ACQ-5, and a slight improvement in FEV1.<sup>97</sup>

The SIRUS (Steroid Reduction with mepoliz Umab Study) in patients with severe asthma and peripheral blood eosinophilia while on maintenance corticosteroid revealed that, patients on mepolizumab had a likelihood of reducing corticosteroid-dose 2.37 times greater than patients on placebo.<sup>98</sup> Patients on mepolizumab were also to reduce the corticosteroid dose by 50%, and had lower exacerbation rates, and improved asthma control despite receiving lower doses of ICS or OCS, thus demonstrating a steroid-sparing effect.<sup>98</sup> Recently, Chupp et al.<sup>99</sup> have confirmed a significant change in the St. George's Respiratory Questionnaire score at the 24<sup>th</sup> week of treatment with add-on mepolizumab. Patients receiving mepolizumab showed improvement in symptoms, and health-related quality of life (HRQoL) scores, compared with control subjects receiving placebo. In summary, mepolizumab has a very good safety and tolerability profile. Add-on treatment with mepolizumab has been shown to improve the ACQ scores, AQLQ scores, SGRQ scores, and FEV1. Additionally, add-on mepolizumab has been shown to reduce the rate of exacerbations, and the dosage of corticosteroid or use of other drug modifiers.<sup>96-99</sup>

Mepolizumab was approved by the FDA on March 23, 2015 for add-on treatment of eosinophilic asthma in adults and children aged  $\geq 12$  years.<sup>100</sup> Mepolizumab was also approved by the European Medicines Agency Committee for Medicinal Products for human use in December 2015.<sup>101</sup> Nucala is also indicated for the treatment of eosinophilic granulomatosis with polyangiitis (EPGA/Churg/Strauss Syndrome). Mepolizumab is not indicated for treatment of the relief of acute bronchoconstriction and status asthmaticus or any other eosinophilic syndromes.

The recommended dose is 100 mg administered subcutaneously every 4 weeks, and it is well tolerated and has been found to be safe.<sup>100</sup> The most common adverse effects with Nucala include injection site reaction, headache, backache, fatigue, muscle weakness, nasopharyngitis, and upper respiratory tract infection. Acute and delayed systemic reactions, including anaphylaxis, urticarial rash, angioedema, bronchospasm, and hypotension may occur. Anaphylaxis is rare ( $<1\%$ ), but patients need to be monitored after treatment for these adverse effects.

Eosinophils play an important role in protection against parasitic infection, including helminth infestation. Patients with pre-existing helminth infections should be treated for the infection before mepolizumab therapy. If individuals become infected whilst receiving treatment with Nucala and do not respond to anti-helminth treatment, temporary discontinuation of mepolizumab should be considered.

## VIII. RESLIZUMAB

Reslizumab (Cingair®), previously known as SCH55700 (Scherig-Plough), is a fully humanized, IgG4/k monoclonal antibody with high affinity for IL-5. The monoclonal antibody has an ERRR configuration (glutamine, arginine, arginine, arginine) corresponding to amino acids 89-92 on the IL-5 antibody molecule. This region is critical for its interaction with the IL-5 receptor which results into inhibition of its bioactivity.<sup>102</sup>

Several randomized clinical trials (RCT) have been conducted on the safety and efficacy of reslizumab. Kips et al.<sup>103</sup> in the first phase 2 pilot study, in patients with severe persistent asthma showed that reslizumab lowered sputum and eosinophil levels, and induced a transient increase in FEV1. A larger phase 2 trial, conducted by Castro et al.<sup>104</sup> showed that treatment with reslizumab significantly increased FEV1, and improved symptoms control, especially in patients with very high eosinophilia and concomitant nasal polyps. Two multicentre, parallel, double-blind, randomised, placebo-controlled, phase 3 trials by Castro et al.<sup>105</sup> demonstrated that reslizumab decreased the annual rate of asthma exacerbation by 50-59% in severe asthmatics with blood eosinophil count  $>400$  cells/ml. Reslizumab also improve asthma symptom control and slightly improved FEV1.<sup>105</sup>

Bjermer and colleagues,<sup>106</sup> in phase 3 trial, have shown that therapy with reslizumab resulted in significant increase in pulmonary function (FEV1), including airflow limitation in peripheral airways, i.e., increase in forced expiratory flow at 25-75% of forced vital capacity (FEF25-75%). Treatment also improved self-reported asthma control, and quality of life. Brusselle and colleagues have also reported that reslizumab is able to reduce asthma exacerbations, and improve lung function in patients with late-onset eosinophilic asthma.<sup>107</sup>

Reslizumab was approved on March 23, 2016 by the FDA for patients aged  $\geq 18$  years as add-on maintenance therapy for severe uncontrolled eosinophilic asthma.<sup>108</sup> The approved dosage for reslizumab is 3 mg/kg intravenously infused over 20-50 minutes every 4 weeks. It is safe and well tolerated by the patients. The most common side effects of Cinqair include headache, nasopharyngitis, myalgia, and fatigue. Anaphylaxis occurs in about 0.3% of the patients,<sup>109</sup> and the U.S. Food and Drug Administration recommends that patients should be observed in a setting where health care professionals are available to treat the adverse reactions. If the patient experiences a severe reaction including anaphylaxis, reslizumab infusion should be discontinued immediately, and the patient should be treated for the adverse event.

Eosinophils play an important in combating helminth infections. Treat patients with pre-existing helminth infections before initiating Cinqair. If patients

become infected while receiving treatment with reslizumab and do not respond to anti-helminth treatment, discontinue treatment with reslizumab until parasitosis resolves.

## IX. BENRALIZUMAB

Benralizumab (Fasenra®), formerly called MEDI-563 (AstraZeneca-MedImmune) is a humanized afucosylated IgG1/k monoclonal antibody, developed via hybridoma technology, which selectively recognize the isoleucin-61 residue of domain 1 of human IL-5R $\alpha$ , located near IL-5 binding site.<sup>110,111</sup> As a result, the interaction of benralizumab with its recognition site on IL-5R $\alpha$  block IL-5 binding to target cells, thus preventing hetero-dimerization of IL5R $\alpha$  and  $\beta$ c subunit, and the subsequent activation of IL-5-dependent signaling pathway.<sup>112</sup> Through the constant Fc region, benralizumab bind to the Fc $\gamma$ R1II $\alpha$  membrane receptor expressed by natural killer cells, which upon activation release the pro-apoptotic proteins granzyme D and perforin, which are responsible for eosinophil apoptosis implemented via antibody-dependent cell-mediated cytotoxicity.<sup>113-115</sup> All these effects cause a reduction in eosinophil numbers in the airway mucosa, submucosa, sputum, blood, and bone marrow.<sup>116</sup>

Preliminary RCT have shown that treatment with benralizumab results in a decrease in blood eosinophil count to almost depletion, which is associated with reductions in the rate of exacerbations, and improvement in the ACQ-5 scores.<sup>117, 118</sup> The SIROCCO RCT showed that treatment with benralizumab significantly reduced exacerbation rates, and improved lung function, and asthma control in patients with severe asthma uncontrolled on high-dose inhaled corticosteroids and long-acting  $\beta$ -agonists.<sup>119</sup> FitzGerald and colleagues in the CALIMA study showed that treatment with subcutaneous benralizumab 30 mg every 4 weeks resulted in a 36% reduction in exacerbations, and a significant increase of 125 ml in FEV1.<sup>120, 121</sup>

The ZONDA phase III oral corticosteroid-sparing trial, demonstrated that add-on benralizumab treatment resulted in up to 51% reduction in annual asthma exacerbation rates versus placebo. There was also a significant improvement in lung function as measured by FEV1. The FEV1 increased by 159 ml, and the improvement in lung function was seen as early as 4 weeks after the initiation of the treatment.<sup>122</sup> Additionally, there was a 75% median reduction in daily oral corticosteroid (OCS) use, and discontinuation of OCS in 52% of the eligible patients.<sup>122</sup> Noteworthy, the BORA RTC revealed that long-term use of add-on benralizumab was associated with a very good safety and tolerability profile.<sup>123</sup> In real-life daily clinical practice, the therapeutic effects of Fasenra may even be better than what is observed in randomized, double-blind clinical trials.<sup>124</sup>

Benralizumab was approved by the U.S. Food and Drug Administration on November 14, 2017, as add-on therapy for people with severe eosinophilic asthma aged 12 years and older, and those whose asthma is not controlled with current asthma medication.<sup>124</sup> Benralizumab has a half-life of 15-18 days, and is available as a single-dose pre-filled syringe. The recommended dose is 30 mg/ml injection subcutaneously every 4 weeks for the first three doses, thereafter every eight weeks. The most common adverse effects of Fasenra include headache (8.6%), nasopharyngitis (4%), arthralgia (3.9%, cough (3.3%), injection site reaction (2.2%), urticaria rash. Other rare adverse events include chills, nausea, dysgeusia, asthenia, tremor, dizziness hot flushes, and hyperhidrosis.

It is not known if Benralizumab will influence helminth infestation or response to anti-helminth treatment. The manufacturers recommend treatment of the parasitosis before initiating Fasenra, and if patients become infected while receiving Fasenra and do not respond to anti-parasitic agents, to discontinue benralizumab until the infection resolves.

## X. CRITERIA FOR INITIATION OF INTERLEUKIN-5 ANTAGONISTS

Biologics should be recommended early in the management of established eosinophilic asthma diagnosed using pharmacodynamic biomarkers, such as sputum and blood eosinophil counts, fractional exhaled nitric oxide (FeNO), serum perisostin, dipeptidyl peptidase-4, and osteopontin.<sup>126-130</sup> Long-term treatment with biologics, such as omalizumab (anti-IgE),<sup>131,132</sup> and mepolizumab,<sup>132</sup> significantly reduce airway wall and reticular membrane thickening. Phipps et al.<sup>133</sup> have reported that mepolizumab is associated with significant reductions in tenascin and lumican deposition in the reticular basement membrane in human atopic skin.<sup>133</sup> If mepolizumab and other anti-IL-5 antagonists exhibit similar effects in eosinophilic airways inflammation, they may be capable of preventing subepithelial fibrosis, and progressive decline in lung function in patients with eosinophilic asthma.

The GINA guidelines,<sup>1</sup> and NAEPP<sup>19</sup> yardsticks for step-up treatment for severe refractory asthma, recommend initiation of biologics, such as anti-IgE, and anti-interleukin (IL)-5 monoclonal antibodies for patients with eosinophilic asthma at step 5. The latest ERS/ATS Task Force guidelines,<sup>134</sup> recommend using anti-IL-5 and anti-IL-5 receptor  $\alpha$  for severe uncontrolled adult eosinophilic asthma phenotypes, using a blood eosinophil cut-point of 150 cells. $\mu$ L<sup>-1</sup> to guide anti-IL-5 initiation in adult patients with severe asthma. The guidelines also suggest specific eosinophil  $\geq 260$  cells. $\mu$ L<sup>-1</sup>, and FeNO 19.5 ppb cut-offs to identify adolescents or adults with the greatest likelihood of



response to anti-IgE therapy. Table 3 shows the three anti-IL-5 antagonists and their weekly costs.

There are few reports on the pharmacoeconomical aspects of the newly introduced biologics for the treatment of severe steroid-resistant eosinophilic asthma. Bogart et al.<sup>135</sup> using a hypothetical model estimates that mepolizumab without bronchial thermoplasty (BT) was the most cost-effective option for biological responders, with a 10-year-per-patient cost of US\$116,776. In patients who do not respond to eosinophilic targeted biologics, bronchial thermoplasty is more cost-effective option. Similarly, an indirect comparison of BT with omalizumab in patients with moderate-to-severe allergic asthma in the USA reported greater than 60% chance that bronchial thermoplasty was cost-effective relative to omalizumab and standard therapy at the willingness-to-pay of \$100,000 per quality-adjusted life years (QALY).<sup>136</sup> However, bronchial thermoplasty is a complex sophisticated procedure which requires critical selection of the patients, experienced pulmonologist, and anesthesiologists, excellent bronchoscopic skills, and dedicated intense post-procedural management and follow-up.<sup>137-140</sup>

## XI. CONCLUSION

Eosinophilic asthma is a well characterized phenotype of asthma, which is driven by Th2 cytokines, such as IL-4, IL-4, IL-13, IL-25, IL-33, and TSPL. Interleukin-5 plays a central role in the differentiation, proliferation, maturation, survival, and activation of eosinophils. Activated eosinophils secrete cytotoxic cationic proteins, radical oxygen species, cytokines, chemokines, and growth factors which are responsible for epithelial injury, airway inflammation, AHR, and airway remodeling. Targeting IL-5 and its receptor with biologics, such as mepolizumab, reslizumab, and benralizumab is a novel therapeutic strategy for the treatment of severe refractory, steroid unresponsive eosinophilic asthma. Early use of anti-IL-5 antagonists may prevent the progressive decline in lung function, and improve the quality of daily living.

### Conflicts of interest

The author reports no conflicts of interest in this manuscript.

## REFERENCES RÉFÉRENCES REFERENCIAS

1. Global Initiative for Asthma. Global Strategy for Asthma management and Prevention - updated 2020. April 2020. [www.ginaasthma.org](http://www.ginaasthma.org).
2. Pearce N, Ait-Khaled N, Beasley R, Mallol J, Keil U, Mitchell E et al. Worldwide trends in the prevalence of asthma symptoms: phase III of the International Study of Asthma and Allergies in Childhood (ISAAC). (Survey) Thorax 2007; 62(9):758.
3. The Global Asthma Network. The Global Asthma Report 2014. Available at: <http://www.glob>
4. alasthmanetwork.org/publications/Global\_Asthma\_Report\_2014.pdf [Last accessed 07 August 2018].
5. Asher MI, Montefort S, Bjorksten B, et al. Worldwide time trends in the prevalence of symptoms of asthma, allergic rhinoconjunctivitis, and eczema in childhood: ISAAC Phase One and Three repeat multi-country cross-sectional surveys. Lancet 2006; 368:733-743.
6. Behbehani N, Abul A, Syabbalo NC, Azeem A, et al. Prevalence of asthma, allergic rhinitis, and eczema in 13-to-14 year-old children in Kuwait: an ISAAC study. Ann Allergy Asthma Immunol 2000; 85(1):58-63. DOI: 10.11016/s1081-1206(10)62435-0.
7. Wenzel SE, Schwartz LB, Langmack EL, Halliday JL, Trudeau JB, Gibbs RL. Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and clinical characteristics. Am J Respir Crit Care Med 1999; 160:1001-1008.
8. Simpson JL, Scott R, Boyle MJ, Gibson PG. Inflammatory subtypes in asthma: assessment and identification using induced sputum. Respirology 2006; 11(1): 54-61.
9. Anderson GP. Endotyping asthma: new insights into key pathogenic mechanism in a heterogenous disease. Lancet 2008; 372:1107-1119.
10. Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approach. Nat Med 2012; 18: 716-725.
11. Pavord ID, Beasley R, Agusti A, et al. After asthma: redefining airways disease. Lancet 2018; 391:350-400.
12. Pembrey L, Barreto ML, Douwes J, Cooper P, Hendersen J, Mpairwe H, Ardura-Garcia M, Chico M, et al. Understanding asthma phenotypes: the World Asthma Phenotypes (WASP) international collaboration. ERJ Open Res 2018; 4(3):00013-2018.
13. Loza MJ, Djukanovic R, Chung KF, Horowitz D, Ma K, Branigan P, et al. Validated and longitudinally stable phenotypes based on cluster analysis of the ADEPT study. Respir Res 2016; 17(1):165. doi: 10.1186/s12931-016-0482-9.
14. Lefaudeaux D, De Meulder B, Loza MJ, Pepper N, Rowe A, Baribaud F, et al. U-BIOPRED clinical adult asthma cluster linked to a subset of sputum omics. J Allergy Clin Immunol 2016; 139(6):1797-1807. doi: 10.1016/j.jaci.2016.08.048.
15. Haldar P, Pavord ID. Noneosinophilic asthma: a distinct clinical and pathologic phenotype. J Allergy Clin Immunol 2007; 119:1043-1052.
16. Aleman F, Lim HF, Nair P. Eosinophilic endotype of asthma. Immunol Allergy North Am 2016; 36(3): 559-568.
17. Taylor SL, Leong LEX, Choo JM, Wesselingh S, Yang IA, Upham JW, et al. Inflammatory phenotypes in patients with severe asthma are associated with



- distinct airway microbiology. *J Allergy Clin Immunol* 2018; 141(1): 94-103 e15.
17. Ntontsi P, Loukides S, Bakakos P, Kostikas K, Papatheodorou G, Papathanassiou E, Papaportfyriou A, Konstantellou E, et al. Clinical, functional and inflammatory characteristics in patients with paucigranulocytic asthma. *Eur Respir J* 2016; 18:PA4173; DOI: 10.1183/13993003.congress-2016.PA4173.
18. National Asthma Education and Prevention Program. Guidelines for the Diagnosis and Management of Asthma. ERP-3. 2007. [www.nhlbi.nih.gov/health-topics/guidelines-for-diagnosis-management-of-asthma](http://www.nhlbi.nih.gov/health-topics/guidelines-for-diagnosis-management-of-asthma). Date last accessed: October 28, 2019.
19. Holguin F, Cardet JC, Chung KF, Diver S, Ferreira DS, Fitzpatrick A, Gaga M, Kellermeyer L, et al. Management of severe asthma: A European Respiratory Society/American Thoracic Society guidelines. *Eur Respir J* 2020; 55(1): DOI: 10.1183/13993003.0058-2019.
20. Peters MC, Kerr S, Duncan EM, Woodruff PG, Fajt ML, Levy BD, Isreal E, Phillips BR, et al; National Lung and Blood Institute Severe Asthma Research Program-3 (2018) Refractory airway type-2 inflammation in a large subgroup of asthmatics treated with inhaled corticosteroids. *J Allergy Clin Immunol pii: S0091-6749(18)30390-7*. doi: 10.1016/j.jaci.2017.12.1009.
21. Futura GT, Atkins FD, Lee NA, et al. Changing roles of eosinophils in health and disease. *Ann Allergy Asthma Immunol* 2014; 113:3-8.
22. Vaglio A, Buzio C, Zwerina J. Eosinophilic granulomatosis with polyangitis (Churg-Stauss): state of the art. *Allergy* 2013; 68:261-273.
23. Ehrlich P. Beiträge zur Kenntniss der granulirten Bindegewebszellen und der eosinophilen Leukocythen. *Arch Anat Physiol (Leipzig)*. 1879; 3:166-169.
24. Fraenkel A. Zu Pathologie de Bronchia lasthma. *Deutch Med Wchnschr* 1900; 17:269.
25. Houston JC, De Navasquez S, Trounce JR. A clinical and pathophysiological study of fatal cases of status asthmaticus. *Thorax* 1953; 8:207-213.
26. Bousquet J, Chanez P, Lacoste JY, et al. Eosinophilic inflammation in asthma. *N Engl J Med* 1990; 323:1033-1039.
27. Rosenberg HF, Phipps S, Foster PS. Eosinophil trafficking in allergy and asthma. *J Allergy Clin Immunol* 2007; 119(6):1303-1310.
28. Blanchard C, Rothenberg ME. Biology of eosinophils. *Adv Immunol* 2009; 101:81-121. doi: 10.1016/S0065-2776(08)01003.
29. McBrien CN, Menzies-Gow A. The biology of eosinophils and their role in asthma. *Front Med (Luasanne)* 2017; 4:93. doi: 10.3389/fmed.
30. Du J, Stankiewicz MJ, Liu Y, Xi Q, Schmidt JE, Lekstrom-Hime JA, et al. Novel combinatorial interactions of GATA-1, PU.1, and C/EBP episilon isoforms regulate transcription of the gene encoding eosinophil granule major basic protein. *J Biol Chem* 2002; 277(45):43481-43494. 10. 10074/jb cM204777.200.
31. Lambrecht BN, Hamad H. Immunology of asthma. *Immunol* 2015; 16(1):45-46. doi: 10.1038/ni.3049.
32. Pease JE. Asthma, allergy and chemokines. *Curr Drug Targets* 2006; 7:3-12. doi: 10.2174/138945006775270204.
33. Griffiths-Johnson DA, Collins PD, Rossi AG, Jose PJ, Williams TJ. The chemokine, eotaxin, activates guinea-pig eosinophils in vitro and causes their accumulation into the lung in vivo. *Biochem Biophys Res Commun* 1993; 197:1167-1172. doi: 10.1006/bbrc.1993.2599.
34. Jia G-Q, Gonzale J-A, Hidalgo A, Wagner D, Cybulsky M, Gutierrez-Ramos JC. Selective eosinophil transendothelial migration triggered by eotaxin via modulation of Mac-1/ICAM-1 and VLA-4/VCAM-1 interactions. *In Immunol* 1999; 11:1-10. doi: 10.1093/intimm/11.1.1.
35. Ying S, Meng Q, Zeibecoglou R, Robinson DS, Macfarlane A, Humbert M, et al. Eosinophil chemotactic chemokines (eotaxin-2, RANTES, monocyte chemo attractant protein-3 (MCP-3), and MCP-4, and C-C chemokine receptor 3 expression in bronchial biopsies from atopic and nonatopic (intrinsic) asthmatics. *J Immunol* 1999; 163(11): 6321-6327.
36. Johannson MW, Kelly EA, Busse WW, Jajour NN, Mosher DF. Up-regulation and activation of eosinophil integrin in blood and airway after segmental lung allergen challenge. *J Immunol* 2008; 180(11):7622-7235. doi: 10.4049/jimmunol.180.11.7622.
37. Klion AD, Nutman TB. The role of eosinophils in host defense against helminth parasites. *J Allergy Clin Immunol* 2004; 113:30-37. doi: 10.1016/j.jaci.2003.10.050.
38. Kay AB. The early history of the eosinophil. *Clin Exp Allergy* 2015; 45: 575-582.
39. Jacobson E, Helmer RA, Lee JJ, Lee NA. The expanding role(s) of eosinophils in health and disease. *Blood* 2012; 120:3882-3890. <https://doi.org/10.1182/blood-2012-06-330845>.
40. Spencer LA, Szela CT, Perez SA, et al. Human eosinophils constitutively express multiple Th1, Th2 and immunoregulatory cytokines that are secreted rapidly and differentially. *J Leukoc Biol* 2009; 85(1):117-123.
41. Bandeira-Melo C, Bozza PT, Weller PF. The cellular biology of eosinophil eicosanoid formation and function. *J Allergy Clin Immunol* 2002; 109(3):393-400. doi: 10.1067/mai.2002.121529.

42. Singh RK, Tandon R, Dastidar SG, Ray A. A review of leukotrienes and their receptors with reference to asthma. *J Asthma* 2013; 50(90):922-931. doi: 10.3109/02770903.2013.823447.
43. Horiuchi T, Weller PF. Expression of vascular endothelial growth factor by human eosinophils: upregulation by granulocyte macrophage colony-stimulating factor and interleukin-5. *Am J Respir Cell Mol Biol* 1997; 17(1):70-77. doi: 10.1165/ajcmb.17.1.2796.
44. Gleich GJ. Mechanisms of eosinophil-associated inflammation. *J Allergy Clin Immunol* 2000; 105:651-663.
45. Hogan SP, Rosenberg HF, Moqbel R, Phipps S, Foster PS, Lacy P, Kay AB, Rothenberg ME. Eosinophils: biological properties and roles in health and disease. *Clin Exp Allergy* 2008; 38(5):709-750. doi: 10.1111/j.1365-2222.2008.02958.x.
46. Carmo LA, Bonjour K, Ueki S, Neves JS, Liu L, Spencer LA, Dvorak AM, Weller PF, Melo RC CD63 is tightly associated with intracellular secretory events chaperoning piecemeal degranulation and compound exocytosis in human eosinophils. *J Leukoc Biol* 2016; 100:391-401. doi: 10.1189/jlb.3A1015-480R.
47. Gleich GJ. Eosinophil granule proteins and bronchial asthma. *Annu Rev Med* 1993; 44:85:101.
48. Trulsson A, Byström J, Engström A, Larsson R, Venge P. The functional heterogeneity of eosinophilic cationic protein determined by a gene polymorphism and post-translational modifications. *Clin Exp Allergy* 2007; 37(2):208-218. doi: 10.1111/j.1365-2222.2007.02644.x.
49. Young JD, Peterson CG, Venge P, Cohn ZA. Mechanisms of membrane damage by human eosinophil cationic protein. *Nature* 1986; 321(6070):613-616. doi: 10.1038/321613a0.
50. Gleich GJ, Flavahan NA, Fujisawa T, Vanhoutte PM. The eosinophil as a mediator of damage to respiratory epithelium: A model for bronchial hyperreactivity. *J Allergy Clin Immunol* 1988; 81:776-781. doi: 10.1016/0091-6749(88)90931-1.
51. Rosenberg HF. Eosinophil-derived neurotoxin/RNase 2: connecting the past, the present and the future. *Cur Pharm Biotechnol* 2008; 9:135-140.
52. Morgan RK, Costello RW, Duncan N, Kingham PJ, Gleich GJ, McLean WG. Diverse effects of eosinophil cationic granule proteins on IMR-32 nerve signaling and survival. *Am J Respir Cell Mol Biol* 2005; 33(2):167-177. doi: 10.1165/rcmb.2005-0056OC.
53. Kay AB. Mediators of hypersensitivity and inflammatory cells in the pathogenesis of bronchial asthma. *Eur J Respir Dis* 1983; 129(Suppl):1-44.
54. Peters-Golden M, Henderson WR, Jr. Leukotrienes. *N Engl J Med* 2007; 357(18):1841-1854.
55. Bochner BS, Gleich GJ. What targeting eosinophils has taught us about their role in diseases. *J Allergy Clin Immunol* 2010; 126:16-25. doi: 10.1016/j.jaci.2010.02.026.
56. Spencer LA, Szela CT, Perez SA, et al. Human eosinophils constitutively express multiple Th1, and Th2 and immunoregulatory cytokines that are secreted rapidly and differentially. *Leukoc Biol* 2009; 5(1):117-123.
57. Durrani SR, Viswanathan RK, Busse WW. What effect does asthma treatment have on airway remodeling? Current perspectives. *J Allergy Clin Immunol* 2011; 128(3):439-448. doi: 10.1016/j.jaci.2011.06.002.
58. Barnes PJ. The cytokine network in asthma and chronic obstructive pulmonary disease. *J Clin Invest* 2008; 118(11):3546-3556. DOI: 10.1172/JCI36130.
59. Shakoory B, Fitzgerald SM, Lee SA, Chi DS, Krishnaswamy G. The role of human mast-cell cytokines in eosinophil biology. *J Interf Cytokine Res* 2004; 24:271-281. doi: 10.1089/107999004323065057.
60. Galdiero MR, Varricchi G, Seaf M, Marone G, Levi-Schaffer F, Marone G. Bidirectional mast cell-eosinophil interactions in inflammatory disorders and cancer. *Front Med* 2017; 4:103. doi: 10.3389/fmed.2017.00103.
61. Kips JC. Cytokines in asthma. *Eur Respir J* 2001; 18(Suppl 34):24s-34s.
62. Steinke JW, Borish L. Th2 cytokines and asthma - interleukin-4: its role in the pathogenesis of asthma, and targeting it for asthma treatment with interleukin-4 receptor antagonists. *Respir Res* 2001; 2:66-70.
63. Nussbaum JC, Van Dyken SJ, von Moltke J, Cheng LE, Mohapatra AB, et al. Type 2 innate lymphoid cells control eosinophil homeostasis. *Nature* 2013; 502:245-248. doi: 10.1038/nature12526.
64. Kabata H, Moro K, Koyasu S, Asono K. Group 2 innate lymphoid cells and asthma. *Allergol Int* 2015; 64:227-234. doi: 10.1016/j.alit.2015.03.004.
65. Wisniewski JA, Borish L. Novel cytokine-producing T cells in allergic disorders. *Allergy Asthma Proc* 2011; 32(2):83-94.
66. Minai-Fleminger Y, Elishmereni M, Vita F, Soranzo MR, Mankuta D, Zabucchi G, et al. Ultra structural evidence for human mast cell-eosinophil interactions in vivo. *Cell Tissue Res* 2010; 341:405-415. doi: 10.1007/s00441-010-1010-8.
67. Woodruff PG, Modrek B, Choy DF, Jia G, Abbas AR, Ellwanger A, et al. T-helper type 2-driven inflammation defines major subphenotypes of asthma. *Am J Respir Crit Care Med* 2009; 180:388-395. doi: 10.1164/rccm.200903-0392OC.
68. Brusselle GG, Maes T, Bracke KR. Eosinophils in the spotlight: Eosinophilic airway inflammation in non allergic asthma. *Nat Med* 2013; 19:977-979. doi: 10.1038/nm.3300.

69. Walker JA, Barlow JL, McKenzie AM. Innate lymphoid cells: how did we miss them? *Nat Rev Immunol* 2013; 13:75-87. 10.1038/nri3349.
70. Smith SG, Chen R, Kjarsgaard M, Huang C, Oliveria JP, O'Byrne PM, et al. Increased numbers of activated group 2 innate lymphoid cells in airways of patients with severe asthma and persistent airway eosinophilia. *J Allergy Clin Immunol* 2016; 137:75-86. 10.1016/j.jaci.2015.05.037.
71. Yanagibashi T, Satoh TM, Nagai Y, Koike M, Takatsu K. Allergic diseases: from bench to clinic - contribution of the discovery of IL-5. *Cytokine* 2017; 98:59-70. 10.1016/j.cyto.2016.11.011.
72. Lambrecht BN, Hammad H, Fahy JV. The cytokines of asthma. *Immunity* 2019; 50:975-991. 10.1016/j.immuni.2019.03.018.
73. Lambrecht BN, Hammad H. The immunology of asthma. *Nat Immunol* 2015; 16:45-56. 10.1038/ni.3049.
74. Milburn MV, Hassell AM, Lambert MH, et al. A novel dimer configuration revealed by the crystal structure at 2.4 Å resolution of human interleukin-5. *Nature* 1993; 363(6425):172-176. doi: 10.1038/363172a0.
75. McKinnon M, Bank M, Solari R, Robinson G. Interleukin-5 and the interleukin receptor: target for drug discovery in asthma. In: Sanderson C. (ed), *Interleukin-5: From Molecule to Drug Target for asthma*. New York: Marcel Dekker, Inc., 1999 pp. 299-320.
76. Menzies-Gow A, Flood-Page P, Sehmi R, Burman J, Hamid Q, Robinson D, et al. Anti-IL-5 (mepolizumab) therapy induces bone marrow eosinophil maturational arrest and decreases eosinophil progenitors in the bronchial mucosa of atopic asthmatics. *J Allergy Clin Immunol* 2003; 111:714-719.
77. Greenfender S, Umland SP, Cuss FM, Chapman RW, Egan RW. Th2 cytokines and asthma – The role of interleukin-5 in allergic eosinophilic disease. *Respir Res* 2001; 2:71-79.
78. Clutterbuck E, Hirst E, Sanderson C. Human interleukin-5 (IL-5) regulates the production in bone marrow cultures: comparison and interaction with IL-1, IL-3, IL-6, and GM-CSF. *Blood* 1989; 73(6):1504-1512.
79. Kouro T, Takatsu L. IL-5- and eosinophil-mediated inflammation: from discovery to therapy. *Int Immun* 2009; 21(12):1303-1309. doi:10.1093/intimm/dxp102.
80. Adachi T, Alam R. The mechanism of IL-5 signal transduction. *Am J Physiol* 1998; 275(3 Pt 1):C623-C633.
81. Molfino NA, Gossage D, Kolbeck R, Parker JM, Geba GP. Molecular and clinical rationale for therapeutic targeting of interleukin-5 and its receptor. *Clin Exp Allergy* 2012; 42:712-737. 10.1111/j.1365-2222.2011.03854.x.
82. Alam R, Pazdrak, Stafford S, Forsythe P. The interleukin-5/receptor interaction activates Lyn and JAK2 tyrosine kinase and signals via Ras-Raf-1-MAP kinase and the JAK-STAT pathway in eosinophils. *Int Arch Allergy Immunol* 1995; 107:226-227. DOI: 10.1017/CBO97811074/5324.004.
83. Pazdrak K, Olszewska-Pazdrak B, Stafford S, Garofalo RP, Alam R. Lyn, Jak2, and Raf-1 kinases are critical for the antiapoptotic effect of interleukin-5, whereas only Raf-1 kinase is essential for eosinophil activation and degranulation. *J Exp Med* 1998; 188:421-429. 10.1084/jem.188.3.421.
84. Sano M, Leff AR, Myou S, Boetticher E, Meliton AY, Learoyd J, et al. Regulation of interleukin-5-induced beta2-integrin adhesion of human eosinophils by phosphoinositide 3-kinase. *Am J Respir Cell Mol Biol* 2005; 33:65-70. 10.1165/rcmb.2005-0076OC.
85. Adachi T, Choudhuri BK, Stafford S, Sur S, Alam R. The differential role of extracellular signal-regulated kinases and p38 mitogen-activated protein kinase in eosinophil functions. *J Immunol* 2000; 165:2198-2204. 10.4049/jimmunol.165.4.2198.
86. Ip WK, Wong CK, Wang CB, Tian YP, Lam CW. Interleukin-3, -3, and granulocyte macrophage colony-stimulating factor induce adhesion and chemotaxis of human eosinophils via p38 mitogen-activated protein kinase and nuclear factor kappaB. *Immunopharmacol Immunotoxicol* 2005; 27:371-393. 10.1080/08923970500240925.
87. Varrichi G, Cononica GW. The role of interleukin 5 in asthma. *Exp Rev Clin Immunol* 2016; 12:903-905. doi.org/10.1080/1744666X.2016.1208564.
88. Varricchi G, Bagnasco D, Borriello F, Heffler E, Cononica GW. Interleukin-5 pathway inhibition in the treatment of eosinophilic respiratory disorders: evidence and unmet needs. *Curr Opin Allergy Clin Immunol* 2016; 16:186-200. 10.1097/ACI.0000000000000251.
89. Agumadu VC, Ramphul K, Mejias SG, Sonaye R, Sombans S, Lohana P. A review of three anti-interleukin-5 monoclonal antibody therapies for severe asthma. *Cureus* 2018; 10(8):e3216. doi: 10.7759/cureus.3216.
90. Zia-Amirhosseini P, Minthorn E, Benincosa L, Hart T, Hottenstein C, Tobias L, et al. Pharmacokinetics and pharmacodynamics of SB-240563, a humanized monoclonal antibody directed to human interleukin-5, in monkeys. *J Pharmacol Exp Ther* 1999; 291:1060-1067.
91. Lopez A, Sanderson C, Gamble J, Campbell H, Young I, Vadas M. Recombinant human interleukin 5 is a selective activator of human eosinophil function. *J Exp Med* 1988; 167:219-224.
92. Ohnishi T, Sur S, Collins D, Fish J, Gleich G, Peters S. Eosinophil survival activity identified as interleukin-5 is associated with eosinophil

- recruitment and degranulation and lung injury twenty-four hours after segmental antigen lung challenge. *J Allergy Clin Immunol* 1993; 92:607-615.
93. Gnanakumaran G, Babu KS. Technology evaluation: mepolizumab, GlaxoSmithKline. *Curr Opin Ther* 2003; 5:321-325.
94. Bagnasco D, Caminati M, Ferrando M, Aloè T, Testino E, Canonica GW, et al. Anti-IL-5 and IL-5R: efficacy and safety of new therapeutic strategies in severe uncontrolled asthma. *Biomed Res Int* 2018;5698212. 10.1155/2018/5698212.
95. Leckie MJ, ten Brinke A, Khan J, Diamant Z, O'Connor BJ, Walls CM, et al. Effects of an interleukin-5 blocking monoclonal antibody on eosinophils, airway hyper responsiveness, and the late asthmatic response. *Lancet* 2000; 356: 2144-2148. doi: 10.1016/S0140-6736(00)03496-6.
96. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl L, Keene ON, et al. Mepolizumab for severe asthma (DREAM): a multicenter, double-blind, placebo-controlled trial. *Lancet* 2012; 380:651-659. 10.1016/S0140-6736(12)60988-X.
97. Ortega HG, Liu MC, Pavord ID, Brusselle GG, Fitzgerald JM, Chetta S, et al. Mepolizumab treatment in patients with severe eosinophilic asthma. *N Engl J Med* 2014; 371:1198-1207. 10.1056/NEJMoa1403290.
98. Bel EH, Wenzel SE, Thompson PJ, Prazma CM, Keene ON, Yancey SW, et al. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med* 2014; 371: 1189-1197. 10.1056/NEJMoa1403291.
99. Chupp GL, Bradford ES, Albers FC, Bratton DJ, Wang-Jairaj J, Nelsen LM, et al. Efficacy of mepolizumab add-on therapy on health-related quality of life and markers of asthma control in severe eosinophilic asthma (MUSCA): a randomized, double-blind, placebo-controlled, parallel-group, multi-center, phase 3b trial. *Lancet Respir Med* 2017; 5:390-400. 10.1016/S2213-2600(17)30125-X.
100. Silver Spring, MD U.S. Food and Drug Administration November 4, 2015. Accessed on January 7, 2016 from <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm471031.htm>.
101. Nucala (Mepolizumab) [FichaTécnica] European Medicine Agency; [Accessed January, 2019]. Published December 2015. [http://www.ema.europa.eu/docs/es\\_ES/document\\_library/EPAR\\_ProductInformation/human/003860/WC5001980037.pdf](http://www.ema.europa.eu/docs/es_ES/document_library/EPAR_ProductInformation/human/003860/WC5001980037.pdf).
102. Zhang J, Kuvelkar R, Murgolo NJ, Taremi SS, Chou CC, Wang P, et al. Mapping and characterization of the epitope(s) of Sch 55700, a humanized mAb, that inhibits human IL-5. *Int Immunol* 1999; 11(12):1935-1944. 10.1093/intimm/11.12.1935.
103. Kips JC, O'Connor BJ, Langley SJ, Woodcock A, Kerstjens HA, Postma DS, et al. Effect of SCH55700, a humanized antihuman interleukin-5 antibody, in severe persistent asthma. A pilot study. *Am J Respir Crit Care Med* 2003; 167:1655-1659. 10.1164/rccm.200206-525OC.
104. Castro M, Mathur S, Hargreave F, Boulet LP, Xie F, Young J, et al. Reslizumab for poorly controlled asthma: a randomized, placebo-controlled study. *Am J Respir Crit Care Med* 2011; 184(10):1125-1132. 10.1164/rccm.201103-0396OC.
105. Castro M, Zangrilli J, Wechsler ME, Bateman ED, Brusselle GG, Bardin P, et al. Reslizumab for inadequately controlled asthma with elevated eosinophil count: results from two multicentre, parallel, double-blind, randomised, placebo-controlled, phase 3 trials. *Lancet Respir Med* 2015; 3:355-366. 10.1016/S2213-2600(15)00042-9.
106. Bjermer L, Lemiere C, Maspero J, Weiss S, Zangrilli J, Germinaro M. Reslizumab for inadequately controlled asthma with elevated blood eosinophil levels: a randomized phase 3 study. *Chest* 2016; 150:789-798. 10.1016/j.chest.2016.03.032.
107. Brusselle G, Germinaro M, Weiss S, Zangrilli J. Reslizumab in patients with inadequately controlled late-onset asthma and elevated blood eosinophils. *Pulm Pharmacol Ther* 2017; 43:39-45. 10.1016/j.pupt.2017.01.011.
108. Silver Spring, MD U.S. Food and Drug Administration March 23, 2016. Accessed on April 8, 2016 from: <http://www.fda.gov/NewsEvents/Newsroom/PressAnnouncements/ucm491980.htm>.
109. TEVA Pharmaceutical Ltd. Reslizumab (Cingaero® (reslizumab). 2006. <http://www.ema.europa.eu>. Accessed Mar 2017.
110. Ishino T, Pasut G, Scibek J, Chaiken I. Kinetic interaction analysis of human interleukin 5 receptor  $\alpha$  mutants reveals a unique binding topology and charge distribution for cytokine recognition. *J Biol Chem* 2004; 279:9547-9556. 10.1074/jbc.M309327200.
111. Kolbeck R, Kozhich A, Koike M, Peng L, Andersson CK, Damschroder MM, et al. Medi-563, a humanized anti-IL-5 receptor  $\alpha$  mAb with enhanced antibody-dependent cell mediated cytotoxicity function. *J Allergy Clin Immunol* 2010; 125:13344-1353. 10.1016/j.jaci.2010.04.004.
112. Pelaia C, Paoletti G, Puggioni F, Racca F, Pelaia G, Canonica GW, Heffler E. Interleukin-5 in the pathophysiology of severe asthma. *Front Physiol* 2019; 10:1514. doi: 10.3389/fphys.2019.01514.
113. Shields RL, Lai J, Keck R, O'Connell LY, Hong K, Meng YG, et al. Lack of fucose on human IgG1 N-linked oligosaccharide improves binding to Fc $\gamma$ RIII and antibody-dependent cellular toxicity. *J Biol Chem* 2002; 277:26733-26740. 10.1074/jbc.M202069200.



114. Ghazi A, Trikha A, Calhoun WJ. Benralizumab - a humanized mAb to IL-5 with enhanced antibody-dependent cell-mediated cytotoxicity - a novel approach for the treatment of asthma. *Expert Opin Biol Ther* 2012; 12:113-118. 10.1517/14712598.2012.642359.
115. Bagnasco D, Ferrando M, Varricchi G, Puggioni F, Passalacqua G, Canonica GW. Anti-interleukin 5 (IL-5) and IL-5Ra: Efficacy, Safety, and Future Perspectives in Severe Eosinophilic Asthma. *Front Med (Lausanne)*. 2017; 4:135. doi: 10.3389/fmed.2017.00135.
116. Tan LD, Bratt JM, Godor D, Louie S, Kenyon NJ. Benralizumab: a unique IL-5 inhibitor for severe asthma. *J Asthma Allergy* 2016; 9:71-81. doi: 10.2147/JAA.S78049.
117. Castro M, Wenzel SE, Bleecker R, et al. Benralizumab, an anti-interleukin 5 receptor  $\alpha$  monoclonal antibody, versus placebo for uncontrolled eosinophilic asthma: a phase 2b randomized dose-ranging study. *Lancet* 2014; 2:879-890.
118. Laviolette M, Gossage DL, Gauvreau G, Leigh R, Olivenstein R, Katal R, et al. Effect of benralizumab on airway eosinophil in asthmatic patients with sputum eosinophilia. *J Allergy Clin Immunol* 2013; 132(5):1086-1095.e5. doi: 10.1016/j.jaci.2013.05.020.
119. Bleecker ER, Fitzgerald JM, Chanez P, Papi A, Weinstein SF, Baker P, et al. Efficacy and safety of benralizumab for patients with severe asthma uncontrolled with high-dose inhaled corticosteroids and long-acting  $\beta$ -agonists (SIROCCO): a randomized multicentre, placebo controlled phase 3 trial. *Lancet* 2016; 388:2115-2127. 10.1016/S0140-6736(16)31324-1.
120. FitzGerald JM, Bleecker ER, Nair P, Korn S, Ohta S, Lommatzsch M, et al. Benralizumab, an anti-interleukin-5 receptor monoclonal antibody, as add-on treatment for patients with severe asthma, uncontrolled eosinophilic asthma (CALIMA): a randomized, double-blind, placebo-controlled phase 3 trial. *Lancet* 2016; 388: 2128-2141. 10.1016/S0140-6736(16)31322-8. 6736(16):1-14. doi: 10.1016/S0140-6736(16)31322.
121. FitzGerald JM, Bleecker ER, Menzies-Gow A, Zangrill JG, Hirch I, Metcalfe P, Newbold P, Goldman M. Predictors of enhanced response with benralizumab for patients with severe asthma: Pooled analysis of the SIROCCO and CALIMA studies. *Lancet Respir Med* 2018; 6:51-64. doi: 10.16/S2213-2600(17)30344-2.
122. Nair P, Wenzel S, Rabe KF, Bourdin A, Lugogo NL, Kuna P, et al. Oral glucocorticoid-sparing effect of benralizumab in severe asthma. *N Engl J Med* 2017; 375:2448-2458. 10.1056/NEJMoa1703501.
123. Busse E, Bleecker R, FitzGerald JM, Ferguson GT, Barker P, Sproule S, et al. Long-term safety and efficacy of benralizumab in patients with severe, uncontrolled asthma: 1-year results from the BORA phase 3 extension trial. *Lancet Respir Med* 2019; 7:46-59. 10.1016/S2213-2600(18)30406-5.
124. Pelaia C, Busceti MT, Vatrella A, Rago GF, Crimi C, Terracciano R, et al. Real-life rapidity of benralizumab effects in patients with severe allergic asthma: assessment of blood eosinophils, symptom control, lung function and oral corticosteroid intake after the first drug dose. *Pulm Pharmacol Ther* 2019; 58:101830. 10.1016/j.pupt.2019.101830.
125. <https://www.astrazeneca.com/media-centre/press-releases/2017/fasenra-benralizumab-receives-us-fda-approval-for-severe-uncontrolled-eosinophiliic-asthma-14112017.html>.
126. Schleich F, Sophie D, Renaud L. Biomarkers in the management of difficult asthma. *Curr Top Med Chem* 2016; 16(22):2521.
127. Wan XC, Woodruff PG. Biomarkers in severe asthma. *Immunol Allergy North Am* 2016; 36(3): 547-557.
128. Yancy SW, Keene ON, Albers FC, Ortega H, Bates S, Bleecker ER, Pavord I. Biomarkers in asthma. *J Allergy Clin Immunol* 2017; 140(6):1509-1518. doi: 10.1016/j.jaci.2017.10.005.
129. Tiotia A. Biomarker in asthma. *State of the art. Asthma Res Pract* 2018; 4: 10.
130. Syabbalo N. Biomarkers for the diagnosis and management of eosinophilic asthma. *Ann Clin Med Res* 2020; 1:1003.
131. Hoshino M, Ohtawa J. Effects of adding omalizumab, an antiimmunoglobulin E antibody, on airway wall thickening in asthma. *Respiration* 2012; 83:520-528.
132. Riccio A, Dal Negro R, Micheletto C, De Ferrari L, Folli C, Chiappori A, et al. Omalizumab modulates bronchial reticular basement membrane and eosinophilic infiltration in severe persistent allergic asthma patients. *Int J Immunopathol Pharmacol* 2012; 25:475-484.
133. Phipps S, Flood-Page P, Menzies-Gow A, Ong Y, Kay A. Intravenous anti-IL-5 monoclonal antibody reduces eosinophils and tenascin deposition in allergen-challenged human atopic skin. *J Invest Dermatol* 2004; 122:1406-1412.
134. Holguin F, Cardet JC, Chung KF, Diver S, Ferriera DS, Fitzpatrick A, Gaga M, Kellermeyer L, et al. Management of severe asthma: A European Respiratory Society/American Thoracic Society guidelines. *Eur Respir J* 2020; 55(1): DOI: 10.1183/13993003.0058-2019.
135. Bogart M, Roberts A, Wheeler S. Cost-effectiveness of refractory asthma treatment strategies: a decision tree analysis. In: *ISPOR 20<sup>th</sup> Annual Meeting*, Philadelphia, PA, USA.

136. Niven RM, Simmonds MR, Cangelosi MJ, Tilden DP, Cottrell S, Shargill NS. Indirect comparison of bronchial thermoplasty versus omalizumab for uncontrolled asthma. *J Asthma* 2017; 55(4):443-451. doi: 10.1080/02770903.2017.1337789136.
137. Bonta PI, Chanez P, Annema JT, Shah PI, Niven R. Bronchial thermoplasty in severe asthma: best practice recommendations from an expert panel. *Respiration* 2018; 95:289-300. doi.org/10.1159/000488291.
138. d'Hooghe JNS, Ten Hacken NHT, Weersink EJM, Sterk PJ, Annema JT, Bonta PI. Emerging understanding of the mechanism of action of bronchial thermoplasty in asthma. *Pharmacol Ther* 2018; 181:101-107.
139. Thomson NC. Bronchial thermoplasty as a treatment for severe asthma: controversies, progress and uncertainties. *Exp Rev Respir Med* 2018; 12(4):269-282. doi: 10.1080/17476348.2018.1444991.
140. Syabbalo N. Clinical features and management of neutrophilic asthma. *J Pulm Med Respir Res* 2020; 6:036. DOI: 10.24966/PMRR-0177/100036.

*Table 1:* Pro-inflammatory mediators synthesized and secreted by eosinophils

Major basic protein (MBP)
Eosinophil cationic protein (ECP)
Eosinophil-derived neurotoxin (EDN)
Eosinophil-derived peroxide (EDPX)
Reactive oxygen species: superoxide, peroxide, and hypobromite
Prostaglandins: PGD <sub>2</sub>
Cysteinyl leukotrienes: LTC <sub>4</sub> , LTD <sub>4</sub> , LTE <sub>4</sub>
Thromboxane B <sub>2</sub> : TXB <sub>2</sub>
Platelet activating factor (PAF)
Cytokines: IL-2, IL-3, IL-4, IL-5, IL-9, IL-13, IL-23, IL-25, IL-33, and TNF- $\alpha$ , GM-CSF
Chemokines: eotaxin-1, -2, and -3, RANTES, P-selectin, MIP-1, MCP-3, MCP-4
Enzymes: histaminases, arylsulfatase, MMP-9, TIMP-1
Growth factors: TGF- $\beta$ , VEGF, PDGF
Abbreviations: LT, leukotriene; IL, interleukin; MMP, matrix metalloproteinases; TIMP, tissue inhibitors of metalloproteinases; MIP, macrophage inflammatory protein; MCP, monocyte chemoattractant protein; GM-CSF, granulocyte macrophage colony-stimulating factor; TGF- $\beta$ , transforming growth factor- $\beta$ ; VEGF, vascular endothelial growth factor; PDGF, platelet-derived growth factor.

*Table 2:* Physiological functions of IL-5 in eosinophil immunobiology

Stimulation of proliferation, and differentiation of IL-5R $\alpha$ -expressing eosinophil committed progenitor cells
Stimulation of maturation of eosinophils
Facilitation of exit of eosinophils from bone marrow
Promotion of eosinophil navigation and migration in the circulation
Aids eotaxins, and adhesion molecules in diapedesis of eosinophils through the endothelial cells
Facilitation of migration of eosinophils to inflamed lung tissues and airways
Activation of eosinophils, and release of pro-inflammatory mediators
Prolongs eosinophil survival synergistically with other anti-apoptotic cytokines

**Table 3:** Approved interleukin-5 and IL-5R $\alpha$  antagonists for eosinophilic asthma IL-5 antagonist Mepolizumab Reslizumab Benralizumab

Date of approval	Nov 4, 2015	March 23, 2016	Nov 14,2017
Mechanism of action	IL-5 antagonist	IL-5 antagonist	IL-5R $\alpha$ blocker
Route of Administration	Subcutaneous	Intravenous	Subcutaneous
Route/frequency	100 mg/4 wk	3mg/kg/4 wk	30 mg/4 wk
Injection	100 mg powder	100 mg/10 mL	30 mg/1 mL
Eosinophil cut-point	$\geq 150$ cells/ $\mu$ L	$\geq 400$ cells/ $\mu$ L	$\geq 150$ cells/ $\mu$ L
FeNO cut-offs	$\geq 19.5$ ppb	$\geq 19.5$ ppb	$\geq$ ppb
Cost/injection	\$2,868	\$2,580	\$4,752
FDA-approved interleukin-5, and IL-5R $\alpha$ antagonists, and the cost of treatment. Adapted from The Medical Letter on Drugs on Therapeutics 2018; 60:33; and the European Respiratory Society/American Thoracic Journal Task Force. European Respiratory Journal 2020; 55(1):1900588.			

