

MicroRNAs as Potential Regulators of Docosahexaenoic Acid Benefits in Alzheimer's Disease

Vic Shao-Chih Chiang

Received: 8 April 2021 Accepted: 5 May 2021 Published: 15 May 2021

Abstract

Alzheimer's disease (AD) is a highly prevalent neurodegenerative disease that imposes a prodigious burden on the society. Docosahexaenoic acids (DHA) are known to be beneficial in AD, in part through their anti-inflammatory properties. MicroRNAs (miRs) are important regulators of brain functions and this regulation becomes disrupted in AD. Objectives: The purpose of this article is to propose the involvement of miRs in the antiinflammatory effects of DHA on AD. Methods: The literature surrounding this topic is extensively researched: miR involvement in the pathophysiology of AD, the mechanism of action of DHA, the effects of DHA on miRs and potential future therapeutic strategies for AD involving miRs. Results: AD results in a disrupted miR network that relates to inflammation, but the altered miRs vary between studies. The effects of DHA on AD are generally positive but the mechanism remains enigmatic. Emerging studies demonstrate that one of the potential mechanisms of action of DHA is modulation of miRs.

Index terms— Omega-3; Diet; Anti-inflammatory; Inflammation; Nutrigenetics; Food-derived microRNA; Exogenous microRNA.

Introduction sporadic Alzheimer's disease (AD) is the age-related neurodegeneration leading to memory impairments, sensory and locomotive dysfunctions, apathy, aggression and eventually leading to splanchic and peripheral system failure due to diminished networks within the central nervous system (CNS). 1 Currently, one in nine elderly above 65 years of age in the United States (US) has AD, excluding cases of preclinical AD. 2 For elderly above 85 years of age, this prevalence becomes as high as one in three.

Author: e-mail: vic.chiang.15@alumni.ucl.ac.uk AD is proposed to be the third leading cause of death in the US. 3 It administers a heavy burden on the society which reaches approximately \$200 billion in the US every year. 4 With the forecast of AD affecting more than 100 million people worldwide by 2050, AD will create a significantly greater burden on the society. 4 The etiology of AD is multifactorial and several hypotheses have been proposed including aberrant amyloid precursor processing into neurotoxic amyloid beta (A β) metabolites, hyper-phosphorylation of the microtubule-stabilizing protein tau, apoptotic alterations of synaptic operations and mitochondrial dysfunction-mediated neurotoxicity. 5,6 There is also significant evidence that supports an inflammatory hypothesis for AD. 7,8 This inflammation is likely to be both the cause 9,10 and the consequence 11,12 of AD. Microglia are macrophages for the CNS and they are found to be excessively activated in AD. 13 Inflammation appears to be an important trigger of this phenomenon. 13 This activation leads to further microglial release of pro-inflammatory mediators which creates a vicious self-perpetuating inflammation cycle that exacerbates AD. 13 Currently approved pharmacotherapies of AD include the use of cholinesterase inhibitors (e.g. rivastigmine) and N-methyl-D-aspartate antagonist (e.g. memantine) to counteract neurotransmission impairments. 14 However these pharmacotherapies only slow the progression but do not cure AD. Many other AD pharmacotherapies are being developed and investigated but most fail during clinical trials. Therefore, there remain major demands for any strategies that can prevent, retard, halt or cure AD.

Dietary modifications offer key strategies against AD as observed from beneficial dietary effects from vitamins, phytochemicals, Mediterranean diet and Souvenaid®. [15][16][17][18] In contrast, diet may also exacerbate AD,

as witnessed for high-fat diets and excessive intake of food contaminated with metals such as cadmium, lead and arsenic. 15,19 Due to the underlying inflammation in AD, antiinflammatory strategies have exhibited promising strategies against AD such as the use of non-steroidal anti-inflammatory drugs and anti-tumor necrosis factor alpha (TNFA) from human observational studies and animal trials. 8 Docosahexaenoic acid (DHA) is well-established for its anti-inflammatory properties and it has been observed in the literature to improve AD. 20 However, its mechanism of action remains enigmatic. Recent evidence suggests potential involvements of the important post-transcriptional gene regulator, microRNAs (miRs). 21 II.

1 MicroRNAs

MiRs were first identified in 1993 by the Ambros and Ruvkun laboratory from *Caenorhabditis elegans* experiments. 22 They were then identified to be conserved phylogenetically across the plant and animal kingdoms which then triggered a revolution in these newly classified non-coding RNAs.

For intergene-or exon-derived miRs, their canonical biogenesis pathway initiates with nuclear transcription of miR-coding genes into primary miR. 23 Cleavage via the microprocessor complex then occurs to form precursor miR (pre-miR), which is transported into the cytoplasm via exportin5. Dicer cleavage leads to the formation of mature double stranded miR duplex, that is incorporated into the RNA-induced silencing complex (RISC), followed by the degradation of the passenger strand. This is slightly different for intron-derived miRs and other emerging non-canonical miR biogenesis pathways are being increasingly recognized. 24 The most well-known function of miRs are its gene-silencing effects on messenger RNA (mRNA) through Watson-Crick interactions between the miR 5' seed sequence nucleotides and the mRNA 3' untranslated region (3'UTR). 23 This leads to either repression of protein translation or degradation of the mRNA transcript. Only partial complementarity is required for gene-silencing effects and therefore single miR can target multiple mRNAs and vice versa. The presence of "isomiRs" 23 and other miR functions 25 further adds to the conundrum of miRs.

The regulation of miRs themselves are extremely complex and far from being understood. They can be regulated through changes in the miR biogenesis components, at the transcription level (transcriptional factors), the post-transcriptional level (pre-miR degradation and modification) as well as at the post-translational level (miR turnover and endogenous sponge activity). 24 Some of these miRs have been determined to be specifically expressed or enriched in certain tissues. For the brain, a number of enriched miRs include miR-128, miR-129, miR-133a, miR-138, miR-153, miR-181a, miR-181b, miR-218, and miR-219. 24 Within neurons, there appears to be an enrichment of miR-125b, miR-128, miR-32, miR-134 and miR-139 in the synaptic and dendritic regions compared to the soma. 24 MiRs have been reviewed to play essential roles in the development and proper functioning of the brain. 26 At the molecular level, miRs are involved in the lineage determination, maturation, survival and neurotransmission. 24 A troika of miRs, miR-134, miR-132 and miR-138 were recently reviewed for their imperative functions in neurons including dendritogenesis, morphogenesis, neuron plasticity, synapse formation, dendritic spine size, cell migration and axon regeneration. 27 They also participate in the differentiation, activation and polarization of microglia. 28 Using dicer knockout experiments, miRs have shown to be crucial for memory formation within the brain. 29 This is further supported by recent research that miR-34a and miR-132 as well as miR-138 regulate memory in rats and humans, respectively. 30,31 These pivotal roles of miRs convey their profound potential as paramount strategies against AD.

2 III. The Roles of MicroRNAs in

Alzheimer's Disease Inflammation

MiRs are known to participate in human AD, and the studies conducted to date are summarized in Table 1. (Insert Table 1 here) The roles of miRs in AD have been reviewed previously. 32 The present review discusses newer studies conducted since the earlier review and reinforces the anti-inflammatory aspects as a proposal for novel strategies against AD. Since inflammation is implicated in AD and miRs are known to participate in inflammation, 33 it is feasible to hypothesize some of the dysregulated miRs in AD are related to inflammation. From the clinical AD and miR studies conducted to date, several miRs were found to be differentially expressed in multiple studies. Some of these have shown to relate with inflammation and presents opportunities for counteracting the inflammation etiology underlying AD. MiR-132 has been consistently found to be down-regulated. [34][35][36] Further work in some of these studies discovered phosphatase in homolog (PTEN), forkhead box (FOX) O3a, FOXO1a and p300 as its direct targets. 35,36 These genes participate in the inflammation-related phosphoinositide 3-kinase pathway (PI3K). 37 (Wong et al., 2013) demonstrated a -3.8 fold change in miR-132 within their Braak VI stage AD patients. 36 However, (Lau et al., 2013) found a more diminished miR-132 down-regulation of approximately -1.6 fold. 35 The robustness of the results from (Lau et al., 2013) study is greater due to higher statistical significance ($p < 0.00001$) and verification of their Nanostring miR data (Nanostrings) with locked-nucleic acids (Exiqon) quantitative real time polymerase chain reaction (qPCR). 35 Disagreement was presented by (Bekris et al., 2013) where they found miR-132 to be up-regulation in AD. 38 However, their study failed to divide samples into Braak stages and adopted inappropriate age-matched controls. Furthermore, their results did not persist following TaqManqPCR (Applied Biosystems) validation.

3 b) miR-146a

MiR-146a has been described extensively as an inflammation-related miR. It was identified to be upregulated in earlier studies, but observed to be downregulated in a more recent study. It regulates nuclear factor kappa B (NFkB), which is a vital pro-inflammatory transcription factor. The disparity between these two AD studies may arise from the difference in the tissues used, where (Muller et al., 2014) profiled only the hippocampus but (Sethi & Lukiw, 2009) profiled the whole temporal lobe neocortex. In addition to that, it was shown by (Muller et al., 2014) that there was actually an up-regulation of miR-146a at Braak III, followed by a decrease in Braak VI.

Since (Sethi & Lukiw, 2009) did not provide Braak staging for their AD patients, (Muller et al., 2014) contributes to more valuable miR-146a changes that allows its surveillance across different Braak stages within AD.

4 c) miR-15a

The up-regulation of miR-15a in AD was determined, but contrasting results were identified in earlier studies [43][44][45]. MiR-15a is validated to target inflammation-related genes including peroxisome proliferated-activated receptor (PPAR)-delta and coactivator-associated arginine methyltransferase 1.

Studies that found miR-15a down-regulation revealed a -1.5 magnitude with statistical significance $p < 0.01$, based on microarray (LC Sciences & Ambion) data. Comparatively, the methodology used by (Bekris et al., 2013) was more robust in that they had three phases of miR screening using TaqManqPCR (Applied Biosystems) and normalization to three housekeeping genes. Although, valid comparison was further complicated by the use of different brain sections in these studies as well as the absence of considerations for Braak staging.

5 d) miR-181c

Down-regulation of miR-181c was detected in three AD studies. [43][44][45] It was shown to be up-regulated in the preliminary screening by (Bekris et al., 2013), but their statistics were not significant enough to proceed to the third stage validation of this study. MiR-181c has been reviewed in terms of its involvement in inflammation in aspects of their regulatory roles on immunosenescence, T cells activity and mitochondrial encoded cytochrome c oxidase 1. Within the AD studies that found miR-181c down-regulation, the resulting magnitudes were discrepant. [43][44][45] This is likely attributable to the use of different brain tissues as well as RNA extraction methodology. In terms of the miR profiling, two of these studies adopted a microarray (LC Sciences & Ambion) approach but did not proceed with conventional qPCR validation.

(Geekiyana & Chan) practiced a comparatively more robust methodology with quantification using single-plexmiScriptqPCR (Qiagen) and their AD cohort being more defined to specifically profile samples at Braak V. They found a -2.5 fold change within the frontal neocortex.

6 e) miR-27

Up-Regulation of miR-27 was recognized in two of the AD miR studies with similar fold changes around 1.7. It was ascertained in other literature to target the anti-inflammatory genes. IL-10 and PPARG expression. Since the Braak stages in AD patients investigated by (Absalon et al., 2013) were only at Braak III, and (Lau et al., 2013) at Braak VI, this may lead to the perception that miR-27 change occurs at an early stage of AD. Nonetheless, several differences between these two studies prohibit this inference to be made. Variations exist for their brain sections profiled, sample size and miR profiling methods.

While (Lau et al., 2013) had a much greater sample size of 41 and extra miR purification steps, the confirmation of miR-27 up-regulation found in their nCounter miR assay (Nanostrings) failed to persist with miRCURY locked nucleic acid qPCR (Exiqon) validation.

7 f) miR-9

Two studies demonstrated miR-9 downregulation. It has been revealed to activate microglia through NFkB signaling and targeting PPARG (Geekiyana & Chan, 2011). It found higher down-regulation of -3.3 compared to -1.3 found by (Hebert et al., 2008). As mentioned previously, the methodology presented by (Geekiyana & Chan, 2011) has higher credibility. Albeit, differences may likewise derive in that the temporal cortex was profiled by (Hebert et al., 2008) in contrast to the frontal neocortex used by (Geekiyana & Chan, 2011).

No gold standard currently exists for miR work, and therefore the comparison between AD miR studies are difficult. This can be further complicated by the discrepancies in the use of brain sections, Braak stages, ethnicity, genotypes, gender and polymorphisms in miR target sites. Nevertheless, these provide valuable grounding for future research.

It is equally important to validate the inflammation-related mRNAs targeted by the differentially expressed miRs in future AD studies to facilitate development of anti-inflammatory strategies against AD.

IV.

8 The Effects of Docosahexaenoic Acids On Alzheimer's Disease

Docosahexaenoic acids (DHA) are well known for its anti-inflammatory properties. It is along chain omega-3 polyunsaturated fatty acid (n3) made up of 22 carbons and 6 cis double bonds in a homoallylic arrangement. The brain is made up of 60% lipids and 15% of these are DHA, which implicates its essential roles within the brain. Since n3 cannot be synthesized de novo, they must be obtained from external sources such as seafood or in the form of alpha linolenic acid (ALA) within certain plant foods. ALA undergoes elongase and desaturase mediated metabolism into eicosapentaenoic acid (EPA) and then into DHA. They can then be carried across the endothelial cells lining the blood brain barrier via MFSD2A and then esterified into phospholipids at the stereospecific number-2 position. Its liberation from neuronal membranes can be made via phospholipase to mediate intracellular anti-inflammatory actions by inhibiting activity of NFkB. Current hypotheses of these actions are proposed to involve the docosanoid pathway via lipoxygenase conversion into resolvins, protectins and maresins or the activation of G-coupled protein receptor 120 and PPARG. DHA is also important for neuronal membrane fluidity, long term memory, neurotransmission and synaptic plasticity. A systematic review concluded from metaanalysis of 18 observational studies, that n3 was beneficial for AD. Furthermore, higher levels of direct n3 biomarkers in the elderly were associated with superior brain white matter, less executive decline and generally positive brain characteristics. In addition, a meta-analysis of animal trials with direct n3 supplementation revealed improvements of AD-related pathophysiology including reduced A β , diminished neuronal loss and improved cognitive function. In contrast to these, in a recent meta-analysis of 34 human clinical trials, n3 did not benefit cognition or AD in the elderly. The inconsistency of these results may emanate from differences in the n3 consumed in terms of the food source, food processing and form of supplementation. These disparities can all alter the chemistry of n3 such as its isomerism, homoallylic arrangement, oxidation and stereospecific numbering. Oxidized n3 can lead to development of various diseases through damage to physiological systems and 62% of marketed n3 supplements have been found to be significantly oxidized. As described earlier, the population used within the study is also paramount and supplementation at later stages of AD appeared to be less effective. These are key considerations for newer n3 clinical trials, but decades of evidence do suggest DHA to be beneficial for the CNS. Through elucidation of the underlying miR mechanism, anti-inflammatory strategies of DHA against AD can be optimized.

V.

9 The Effects of Docosahexaenoic Acids on Micrnas

MiR studies investigating dietary modifications in the CNS are extremely exiguous and none of these are directly relevant to AD. With regards to the relationship between DHA and miR, only eight published studies exist and these findings are summarized in Table 2. The effects of DHA on miR have been reviewed partially in 2012, but many more miR studies of DHA have been generated since then. Glioma cells U87 MG (treated with radiation) miR-146; miR-181a

Most of these were addressed in other physiological systems with only two conducted within the CNS. These studies in other physiological systems still provide relevant insights to potential miR mechanisms in DHA anti-inflammatory effects on AD. For example, one study illustrated an up-regulation of miR-107 and their further validation revealed the amyloid processing enzyme, beta-secretase 1, as a miR-107 target. a) Farago et al., 2011 Study In this CNS study, the highly fatal malignant glioma was investigated. Their study explored the effects of multiple PUFAs, including DHA based on previous literature evidence that these can combat gliomas. Their aim was to elucidate the miR mechanism of DHA action on gliomas.

For this, they treated glioblastoma cells (U373, GBM2, GBM5) with 50 & 100 μ M DHA for 24 hours and then extracted for their miR (Roche). The miRs were profiled using a megaplex TaqMan qPCR (Applied Biosystems) followed by further validation with singleplex TaqMan qPCR (Applied Biosystems). They also quantified levels of selected apoptotic mRNA targets that were predicted for the differentially expressed miR using qPCR.

The only miR that was found to change in all three cell lines was miR-145, but the direction of change was discrepant for U373. This suggests possible specificity of DHA action on different cell lines. This notion is further supported by disparities in the magnitude of miR-145 change between GBM2 of -1.5 fold with GBM5 of -4.7 fold. Therefore, it would be more appropriate to discuss these cell lines separately.

In GBM2, other miRs that were altered include down-regulation of miR-30c and up-regulation of miR-143. Down-regulation of miR-22, miR-30c and miR-143 were discovered for GBM5 as well as up-regulation of miR-20b. In U373, aside from the miR-145 up-regulation, miR-22 was found to be down-regulated.

In terms of the mRNA targets, correlation was made to their complementary miR. Successful inverse relationships were found for miR-20b with tumor protein p53 inducible nuclear protein 1 (TP3INP1), miR-22 with sirtuin 1 (SIRT1), miR-30c with integrin, beta3 (ITGB3), miR-143 with v-Ki-ras2 oncogene (KRAS) and prostaglandin-endoperoxide synthase 2 (COX2) as well as miR-145 with insulin receptor substrate 1 (IRS1). These miR associations with apoptotic genes suggest DHA involvement in apoptosis, which is closely related with inflammation. b) ?ntal et al., 2014 Study This CNS study was conducted by the same research group and they again investigated glioma but this time they focused on DHA enhancement of radiotherapy against glioma cells. The mechanism of this radiotherapy enhancement by DHA remains enigmatic and therefore the researchers intended to determine the miR mechanism to optimize radiotherapy against glioma.

They subjected U87 glioblastoma cells under 10Gy cobalt irradiation and then treated with 25 μ M DHA for 48 hours. RNA was extracted from cells (Bioneer) and selected miR expression were quantified using singleplexTaqManqPCR (Applied Biosystems). Candidate mRNAs were similarly quantified using qPCR.

The combination of DHA with radiation did not alter any of the miRs that were measured (miR-34a; miR-96; miR-146; miR-181a; miR-148a; miR-148b and miR-152), but DHA alone up-regulated miR-146a and miR-181a. With DHA treatment alone, it was sufficient to pose significant negative effects on U87 cells. In the case of mRNAs, they found up-regulation of oxidative stress related genes including anti-inflammatory heme oxygenase (decycling) 1 (HMOX1) and pro-apoptotic NAD(P)H dehydrogenase, quinone 1 (NQO1). Genes related to endoplasmic reticulum (ER) stress were also altered encompassing the pro-survival G protein-coupled receptor 78 (GPR78) and pro-apoptotic DNA damage-inducible transcript 3 (DDIT3). Early growth response protein 1 (EGR1) is an early-response gene in radiotherapy that coordinates cell differentiation and growth. It was up-regulated with DHA treatment. The Notch signaling pathway was likewise altered through up-regulation of NOTCH1. Despite validation of direct miR-146a and miR-181 targets were not performed in this study, the up-regulation of multiple genes indicates probable involvement of these miRs in these pathways. Many of these pathways including oxidative stress, ER stress and Notch signaling are known to relate with inflammation. Portions of the DHA-regulated miRs similarly correspond to miRs that were altered in AD (Table 1). Some of these have also been discussed earlier to engage in inflammation including miR-15, miR-27, miR-181 and miR-146. Aside from these inflammation-related miRs already discussed, some of the other DHA-regulated miRs are likewise known to associate with inflammation. These include RECK, lipid metabolism, oncology and stress pathways. Through this elucidation of DHA anti-inflammatory mechanisms, they offer valuable insight as a strategy against AD.

The two studies described demonstrate the potential for DHA to affect miR expression within CNS. While they do provide beneficial preliminary intuition, the cell specificity of DHA effects signifies the importance of establishing their effects on AD-related neuronal and neuroglial miR expression.

The objective to investigate their underlying anti-inflammatory mechanism in AD requires thorough considerations. For in vitro studies, the considerations for AD-relevant cell lines or primary cells are necessary. As described above, the parameters of the DHA supplemented is equally important. Furthermore, the miR methodology demands the adoption of robust procedures that are comparable to high quality studies. Animal models and human studies will also be useful. As addressed above, regards need to be paid for the characterization of AD patients, brain section that is profiled and sample size.

10 VI.

How MicroRNAs Contribute to Future Strategies against Alzheimer's Disease

By understanding miRs that are responsible for anti-inflammatory effects of DHA on AD, improvements can be made to existing tactics as well as development of novel strategies. The first miR therapy is presently being tested under clinical trial to explore miR-34 replacement as an anti-tumor therapy. Exogenous miRs offer possible solution where exogenous miR transfer from dietary origin was first documented in 2012 that discovered the presence of rice-derived miRs in human serum. It is feasible for miRs to survive gastrointestinal digestion due to their well-appreciated stability. This property is ascribed to their selective cellular export into various transport mechanisms. The extracellular miRs can then be taken up by recipient cells to mediate function distally. Newer studies further support food-mediated miR transfer into human, porcine and murine biological fluids from plants and milk. [79][80][81] By contrast, there are similarly studies that refute this notion as shown in bees, mice, macaques and human. The presence of miRs from other species have been recently reported to possibly arise from undesirable contamination or artefacts of sequencing methodologies. Based on these arguments, dietary transfer of miRs remains to be concluded. Additional considerations need to be made whether the amount of exogenous miR transferred translate into biological significance. Nevertheless, these demonstrate the feasibility of using exogenous miRs as strategies against AD.

The concept of exogenous miR supplementation for AD requires knowledge of miR mechanisms of how DHA antagonizes AD inflammation. The miRs that are responsible can be supplemented to create novel food products or nutraceuticals. Genetic engineering can likewise be adopted to enhance levels of these miRs within foods. In an alternative perspective, the efficiency of miR-modulation by DHA in AD can be enhanced through understanding which aspects of the DHA molecule (e.g. unsaturation, stereospecificity, MicroRNAs as Potential Regulators of Docosahexaenoic Acid Benefits in Alzheimer's Disease allylism) are responsible for its anti-inflammatory effects. This can lead to fabrication of optimized DHA and derivatives to maximize their miR effects to antagonize AD inflammation.

11 Medical

AD is afflicted with a pathological state of inflammation and this can be counteracted through anti-inflammatory effects of DHA. The underlying mechanism likely involves miRs and through its elucidation, novel strategies can be developed to combat AD. AD is highly prevalent, affecting 1 in 3 elderlies above 85 years of age. It is the third leading cause of death in United States and attributes to a financial burden of \$200 billion annually.

280 Any prevention, retardation, termination or reversal strategies against AD will reduce the significant and rapidly
281 growing societal burden attributed by AD. ¹

¹© 2021 Global JournalsMicroRNAs as Potential Regulators of Docosahexaenoic Acid Benefits in Alzheimer's Disease

Author	Clinical Samples	Up-regulated microRNA 1	Down-regulated MicroRNA 1	
Wang et al. (2008) 85	Cerebral cortex n=6		miR-103; miR-107; miR-23b	
Hebert et al. (2008) 43	Anterior temporal cortex n=5	miR-520h; miR-197; miR-511; miR-320; miR-516-3p;	let-7i; miR-22; miR-93; miR-26b; miR-9; miR-488; miR-363; miR-181c; miR-106b; miR-101; miR-210; miR-15a; miR-19b; miR-29b;	
Sethi&Lukiw (2009) 40	Temporal lobe neocortex n=6	miR-9; miR-125b; miR-146a		
Nunez-Iglesias et al. (2010) 44	Parietal lobes n=5	617; miR-30184; miR-671; miR-188; miR-miR-185; miR-382; miR-432; miR-486; miR-19790; miR-28648; miR-572; miR-18895; miR-35456; miR-320; miR-134; miR-45605; miR-10939; miR-10912; miR-	miR-101; miR-20b; miR-08570; miR-29b; miR-374; miR-582; miR-05109; miR-12504; miR-12497; miR-30e-5p; miR-376a; miR-44608; miR-181c; miR-368; miR-95; miR-20546; miR-148b; miR-02532; miR-42448;	17 Year 2021
Shioya et al. (2010) 86	Frontal lobes n=7	06383; miR-765; miR-575; miR-23974; miR-601;	miR-15a; miR-130a; miR-29c;	Volume XXI
Geekiyana& Chan (2011) 45	Frontal cortices n=7		miR-598; miR-494; miR-29a miR-153	Is- sue V
Long et al. (2012) 87	Frontal cortex n=5			Ver- sion I
Lau et al. (2013) 35	Hippocampus n=41			(D D D D)
Wong et al. (2013) 36	Temporal cortex n=6		miR-132; miR-212	Research Medical
Bekris et al. (2013) 38	Cerebellum n=21 Hip-	miR-138; miR-208b; miR-181c; miR-152; miR-126; miR-330-3p; miR-184; miR-191; miR-328; miR-342-3p; miR-370; miR-501; miR-331-3p; miR-139-5p; miR-149; miR-132; miR-98; miR-204; 3p; miR-139-5p; miR-149; miR-132; miR-miR-138; miR-208b; miR-181c; miR-152; miR-126; miR-330-3p; miR-191; miR-328;	miR-184	Global Jour- nal of
Bekris et al. (2013) 38	pocampus n=21			

2

Author	Model	Up-regulated microRNA	Down- regulated MicroRNA
Davidson et al. (2009) 66	Sprague-Dawley rats colon (induced colon tumour by azoxymethane)	Let-7d; miR-15b; miR- 107; miR-191; miR-324- 5p	
Farago et al. (2011) 67	GBM2 Glioma cells	miR-143	miR-30c; miR-145

Figure 2: Table 2 :

[Alzheimers Dement ()] , *Alzheimers Dement* 2014. 10 p. .

[Shioya et al. ()] ‘Aberrant microRNA expression in the brains of neurodegenerative diseases: miR-29a decreased in Alzheimer disease brains targets neurone navigator 3’. M Shioya , S Obayashi , H Tabunoki , K Arima , Y Saito , T Ishida . *Neuropathol Appl Neurobiol* 2010. 36 p. .

[Siddesha et al. ()] ‘Acetylsalicylic acid inhibits IL-18-induced cardiac fibroblast migration through the induction of RECK’. J M Siddesha , A J Valente , Svp Sakamuri , J D Gardner , P Delafontaine , M Noda . *J Cell Physiol* 2014. 229 (7) p. .

[Lee et al. ()] ‘Activated human microglia stimulate neuroblastoma cells to upregulate production of beta amyloid protein and tau: implications for Alzheimer’s disease pathogenesis’. M Lee , E Mcgeer , P L Mcgeer . *Neurobiol Aging* 2015. 36 (1) p. .

[Lau et al. ()] ‘Alteration of the microRNA network during the progression of Alzheimer’s disease’. P Lau , K Bossers , R Janky , E Salta , C S Frigerio , S Barbash . *EMBO Mol Med* 2013. 5 p. .

[Hebert et al. ()] ‘Alzheimer disease in the United States (2010-2050) estimated using the 2010 census’. L E Hebert , J Weuve , P A Scherr , D A Evans . *Neurology* 2013. 80 (19) p. .

[Alzheimer’s disease: a systematic review and metaanalysis J Alzheimers Dis ()] ‘Alzheimer’s disease: a systematic review and metaanalysis’. *J Alzheimers Dis* 2012. 28 p. .

[Henson and Bratton ()] ‘Antiinflammatory effects of apoptotic cells’. P M Henson , D L Bratton . *J Clin Invest* 2013. 123 (7) p. .

[Hotchkiss and Nicholson ()] ‘Apoptosis and caspases regulate death and inflammation in sepsis’. R S Hotchkiss , D W Nicholson . *Nat Rev Immunol* 2006. 6 p. .

[Liang et al. ()] ‘Assessing the survival of exogenous plant microRNA in mice’. G F Liang , Y L Zhu , B Sun , Y Shao , A Jing , J Wang . *Food Sci Nutr* 2014. 2 (4) p. .

[Adlakha and Saini ()] ‘Brain microRNAs and insights into biological functions and therapeutic potential of brain enriched miRNA-128’. Y K Adlakha , N Saini . *Mol Cancer* 2014. 13 p. .

[Baselga-Escudero et al. ()] ‘Chronic administration of proanthocyanidins or docosahexaenoic acid reverses the increase of miR-33a and miR-122 in dyslipidemic obese rats’. L Baselga-Escudero , A Arola-Arnal , A Serrano , A Ribas-Latre , E Casanova , M J Salvado . *PLoS ONE* 2014. 8 (7) p. e69817.

[Zhang et al. ()] ‘Circulating mitochondrial DAMPs cause inflammatory responses to injury’. Q Zhang , M Raoof , Y Chen , Y Sumi , T Sursal , W Junger . *Nature* 2010. 44 p. .

[Virtanen et al. ()] ‘Circulating omega-3 polyunsaturated fatty acids and subclinical brain abnormalities on MRI in older adults: the cardiovascular health study’. J K Virtanen , D S Siscovick , R N Lemaitre , Longstreth Jr , W T Spiegelman , D Rimm , EB . *J Am Heart Assoc* 2013. 2 p. e000305.

[Liu et al. ()] ‘Coactivator-associated arginine methyltransferase 1 targeted by miR-15a regulates inflammation in acute coronary syndrome’. X Liu , L Wang , H Li , X Lu , Y Hu , X Yang . *Atherosclerosis* 2014. 233 p. .

[Luchtman and Song ()] ‘Cognitive enhancement by omega-3 fatty acids from childhood to old age: findings from animal and clinical studies’. D W Luchtman , C Song . *Neuropharmacology* 2013. 64 p. .

[Antal et al. ()] ‘Combination of unsaturated fatty acids and ionizing radiation on human glioma cells: cellular, biochemical and gene expression analysis’. O Antal , L Hackler , J Shen , I Man , K Hideghety , K Kitajka . *Lipids Health Dis* 2014. 13 p. .

[James et al. ()] ‘Contribution of Alzheimer disease to mortality in the United States’. B D James , S E Leurgans , L E Hebert , P A Scherr , K Yafe , D A Bennett . *Neurology* 2014. 82 (12) p. .

[Wong et al. ()] ‘De-repression of FOXO3a death axis by microRNA-132 and -212 causes neuronal apoptosis in Alzheimer’s disease’. H A Wong , T Veremeyko , N Patel , C A Lemere , D M Walsh , C Esau . *Hum Mol Genet* 2013. 22 (15) p. .

[Brenna and Carlson ()] ‘Docosahexaenoic acid and human brain development: evidence that a dietary supply is needed for optimal development’. J T Brenna , S E Carlson . *J Hum Evol* 2014. 77 p. .

[Gil-Zamorano et al. ()] ‘Docosahexaenoic acid modulates the enterocyte Caco-2 cell expression of microRNAs involved in lipid metabolism’. J Gil-Zamorano , R Martin , L Daimiel , K Richardson , E Giordano , N Nicod . *J Nutr* 2014. 144 (5) p. .

[Siddesha et al. ()] ‘Docosahexaenoic acid reverses angiotensin II-induced RECK suppression and cardiac fibroblast migration’. J M Siddesha , A J Valente , T Yoshida , Svp Sakamuri , P Delafontaine , H Iba . *Cell Signal* 2014. 26 p. .

[Yang et al. ()] ‘Effect of docosahexaenoic acid on hippocampal neurons in high-glucose condition: involvement of PI3K/AKT/NFkB-mediated inflammatory pathways’. R H Yang , J Lin , X H Hou , Rf Yu , H Q Liu , A L Ji . *Neuroscience* 2014. 274 p. .

- [Jiao et al. ()] 'Effect of n3 PUFA supplementation on cognitive function throughout the life span from infancy to old age: a systematic review and meta-analysis of randomized controlled trial'. J Jiao , Q Li , J Chu , W Zeng , M Yang , S Zhu . *Am J Clin Nutr* 2014. 100 p. .
- [Cheng et al. ()] 'Emerging roles of the γ -secretase-notch axis in inflammation'. Y L Cheng , Y Choi , C G Sobey , T V Arumugam , D G Jo . *Pharmacol Ther* 2015. 147 p. .
- [Beydoun et al. ()] 'Epidemiologic studies of modifiable factors associated with cognition and dementia: systematic review and meta-analysis'. M A Beydoun , H A Beydoun , A A Gamaldo , A Teel , A B Zonderman , Y Wang . *BMC Public Health* 2014. 14 p. .
- [Nicolia et al. ()] 'epigenetics and neurodegeneration: Focus on nutrition in Alzheimer's disease'. V Nicolia , M Lucarelli , Fuso A Environment . *Exp Gerontol* 2014.
- [Zhang et al. ()] 'Exogenous plant MIR168a specifically targets mammalian LDLRAP1: evidence of cross-kingdom regulation by microRNA'. L Zhang , D X Hou , X Chen , D H Li , L Y Zhu , Y J Zhang . *Cell Res* 2012. 22 p. .
- [Park et al. ()] 'Extracellular microRNAs activate nociceptor neurons to elicit pain via TLR7 and TRPA1'. C K Park , Z Z Xu , T Berta , Q Han , G Chen , X J Liu . *Neuron* 2014. 82 p. .
- [Albert et al. ()] 'Fish oil supplements in New Zealand are highly oxidised and do not meet label content of n-3 PUFA'. B B Albert , Jgb Derraik , D Cameron-Smith , P L Hofman , S Tumanov , S G Villas-Boas . *Sci Rep* 2015. 5 (7928) .
- [O'carroll and Schaefer ()] 'General principals of miRNA biogenesis and regulation in the brain'. D O'carroll , A Schaefer . *Neuropsychopharmacology* 2013. 38 p. .
- [Mandal et al. ()] 'Ghosh-Choudhury N. miR-21 is targeted by omega-3 polyunsaturated fatty acid to regulate breast tumor CSF-1 expression'. C C Mandal , T Ghosh-Choudhury , N Dey , G G Choudhury . *Carcinogenesis* 2012. 33 (10) p. .
- [Ferri et al. ()] 'Global prevalence of dementia: a Delphi consensus study'. C P Ferri , M Prince , C Brayne , H Brodaty , L Fratiglioni , M Ganguli . *Lancet* 2005. 366 p. .
- [Morozova et al. ()] 'Harel-Bellan A. Kinetic signatures of microRNA modes of action'. N Morozova , A Zinovyev , N Nonne , L L Pritchard , A N Gorban . *RNA* 2012. 18 p. .
- [Knight et al. ()] 'High-fat diet-induced memory impairment in triple-transgenic Alzheimer's disease (3xTgAD) mice is independent of changes in amyloid and tau pathology'. E M Knight , Iva Martins , S Gumusgoz , S M Allan , C B Lawrence . *Neurobiol Aging* 2014. 35 p. .
- [Ishii and Haga ()] 'Identification of components of immunoglobulins in senile plaques by means of fluorescent antibody technique'. T Ishii , S Haga . *Acta neuropath* 1975. 32 p. .
- [Lukasik and Zielenkiewicz ()] 'In silico identification of plant miRNAs in mammalian breast milk exosomes -a small step forward?'. A Lukasik , P Zielenkiewicz . *PLoS ONE* 2014.
- [Snow et al. ()] 'Ineffective delivery of diet-derived microRNAs to recipient animal organisms'. J W Snow , A E Hale , S K Isaacs , A L Baggish , Chan Sy . *RNA Biol* 2013. 10 (6) p. .
- [Macchi et al. ()] 'Inflammation and programmed cell death in Alzheimer's disease: comparison of the central nervous system and peripheral blood'. B Macchi , F Marino-Merlo , C Frezza , S Cuzzocrea , A Mastino . *Mol Neurobiol* 2014. 50 p. .
- [Ferreira et al.] *Inflammation, defective insulin signaling, and neuronal dysfunction in Alzheimer's disease*, S T Ferreira , J R Clarke , T R Bomfim , F G De Felice .
- [Shah et al. ()] 'Integrated microRNA and mRNA expression profiling in a rat colon carcinogenesis model: effect of a chemo-protective diet'. M S Shah , S L Schwartz , C Zhao , L A Davidson , B Zhou , J R Lupton . *Physiol Genomics* 2011. 43 p. .
- [Boon and Vickers ()] 'Intercellular transport of microRNAs'. R A Boon , K C Vickers . *Arterioscler Thromb Vasc Biol* 2013. 33 p. .
- [Nunez-Iglesias et al. ()] 'Joint genome-wide profiling of miRNA and mRNA expression in Alzheimer's disease cortex reveals altered miRNA regulation'. J Nunez-Iglesias , C C Liu , T E Morgan , C E Finch , X J Zhou . *PLoS ONE* 2010. 5 (2) p. e8898.
- [Hebert et al. ()] 'Loss of microRNA cluster miR-29a/b-1 in sporadic Alzheimer's disease correlates with increased BACE1/beta-secretase expression'. S S Hebert , K Horre , L Nicolai , A S Papadopoulou , W Mandemakers , A S Silahdaroglu . *Proc Natl Acad Sci* 2008. 105 (17) p. .
- [Ben-Zvi et al. ()] 'Mfsd2a is critical for the formation and function of the blood-brain barrier'. A Ben-Zvi , B Lacoste , E Kur , B J Andreone , Y Mayshar , H Yan . *Nature* 2014. 509 p. .
- [Sethi and Lukiw ()] 'Micro-RNA abundance and stability in human brain: Specific alterations in Alzheimer's disease temporal lobe neocortex'. P Sethi , W J Lukiw . *Neurosci Lett* 2009. 459 p. .

393 [Almeida et al. ()] 'MicroRNA history: discovery, recent applications, and next frontiers'. M I Almeida , R M
394 Reis , G A Calin . *Mutat Res Fundam Mol Mech Mutagen* 2011. 717 p. .

395 [Bekris et al. ()] 'MicroRNA in Alzheimer's disease: an exploratory study in brain, cerebrospinal fluid and
396 plasma'. L M Bekris , F Lutz , T J Montine , C E Yu , D Tsuang , E R Peskind . *Biomarkers* 2013. 18
397 (5) p. .

398 [Farago et al. ()] 'MicroRNA profile of polyunsaturated fatty acid treated glioma cells reveal apoptosis-specific
399 expression changes'. N Farago , L Z Feher , K Kitajka , U N Das , L G Puskas . *Lipids Health Dis* 2011. 10
400 p. .

401 [Bredy et al. ()] 'MicroRNA regulation of neural plasticity and memory'. T W Bredy , Q Lin , W Wei , D
402 Baker-Andresen , J S Mattick . *Neurobiol Learn Mem* 2011. 96 p. .

403 [Long and Lahiri ()] 'MicroRNA-101 downregulates Alzheimer's amyloid-beta precursor protein levels in human
404 cell cultures and is differentially expressed'. J M Long , D K Lahiri . *Biochem Biophys Res Commun* 2011.
405 404 p. .

406 [Bicker et al. ()] 'MicroRNA-132, -134, and -138: a microRNA troika rules in neuronal dendrites'. S Bicker , M
407 Lackinger , K Weib , G Schratt . *Cell Mol Life Sci* 2014. 71 p. .

408 [Geekiyange and Chan ()] 'MicroRNA-137/181c regulates serine palmitoyltransferase and in turn amyloid beta,
409 novel targets in sporadic Alzheimer's disease'. H Geekiyange , C Chan . *J Neurosci* 2011. 31 (41) p. .

410 [Schroder et al. ()] 'MicroRNA-138 is a potential regulator of memory performance in humans'. J Schroder , S
411 Ansaloni , M Schilling , T Liu , J Radke , M Jaedicke . *Front Hum Neurosci* 2014. 8 (501) p. .

412 [Thulin et al. ()] 'MicroRNA-9 regulates the expression of peroxisome proliferator-activated receptor ? in human
413 monocytes during the inflammatory response'. P Thulin , T Wei , O Werngren , L Cheung , R M Fisher , D
414 Grander . *Int J Mol Cell Med* 2013. 31 (5) p. .

415 [Baier et al. ()] 'MicroRNAs are absorbed in biologically meaningful amounts from nutritionally relevant doses
416 of cow milk and affect gene expression in peripheral blood mononuclear cells, HEK-293 kidney cell cultures,
417 and mouse livers'. S R Baier , C Nguyen , F Xie , J R Wood , J Zempleni . *J Nutr* 2014. 144 p. .

418 [Ponomarev et al. ()] 'MicroRNAs are universal regulators of differentiation, activation, and polarization of
419 microglia and macrophages in normal and diseased CNS'. E D Ponomarev , T Veremeyko , H L Weiner
420 . *Glia* 2013. 61 p. .

421 [Delay et al. ()] 'MicroRNAs in Alzheimer's disease'. C Delay , W Mandemakers , S S Hebert . *Neurobiology of*
422 *Disease* 2012. 46 p. .

423 [Muller et al. ()] 'MicroRNAs in Alzheimer's disease: differential expression in hippocampus and cell-free
424 cerebrospinal fluid'. M Muller , H B Kuiperij , J A Claassen , B Kusters , M M Verbeek . *Neurobiol Aging*
425 2014. 35 p. .

426 [Thounaojam et al. ()] 'MicroRNAs in the brain: it's regulatory role in neuroinflammation'. M C Thounaojam ,
427 D K Kaushik , A Basu . *Mol Neurobiol* 2013. 47 p. .

428 [Tosar et al. ()] 'Mining of public sequencing databases supports a nondietary origin for putative foreign miRNAs:
429 underestimated effects of contamination in NGS'. J P Tosar , C Rovira , H Naya , A Cayota . *RNA* 2014. 20
430 (6) p. .

431 [Absalon et al. ()] 'MiR-26b, upregulated in Alzheimer's disease, activates cell cycle entry, tau-phosphorylation,
432 and apoptosis in postmitotic neurons'. S Absalon , D M Kochanek , V Raghavan , A M Krichevsky . *J*
433 *Neurosci* 2013. 33 (37) p. .

434 [Wang et al. ()] 'MiR-27a is up regulated and promotes inflammatory response in sepsis'. Z Wang , Z Ruan , Y
435 Mao , W Dong , Y Zhang , N Yin . *Cell Immunol* 2014. 290 p. .

436 [Misso et al. ()] 'Mir-34: a new weapon against cancer?'. G Misso , M T Di Martino , G De Rosa , A A Farooqi
437 , A Lombardi , V Campani . *Mol Ther Nucleic Acids* 2014. 3 p. e194.

438 [Yao et al. ()] 'MiR-9 promotes microglial activation by targeting MCP1'. H Yao , R Ma , L Yang , G Hu , X
439 Chen , M Duan . *Nat Commun* 2014. 5 (4386) p. .

440 [Tetreault and De Guire ()] 'MiRNAs: their discovery, biogenesis and mechanism of action'. N Tetreault , V De
441 Guire . *Clin Biochem* 2013. 46 (10) p. .

442 [Rippo et al. ()] 'MitomiRs in human inflamm-aging: a hypothesis involving miR-181a, miR-34a and miR-146a'.
443 M R Rippo , F Olivieri , V Monsurro , F Prattichizzo , M C Alberii , A D Procopio . *Exp Gerontol* 2014. 56
444 p. .

445 [Visioli et al. ()] 'Molecular targets of omega 3 and conjugated linoleic fatty acids -"micromanaging" cellular
446 response'. F Visioli , E Giordano , N M Nicod , A Davalos . *Front Physiol* 2012. 3 (42) .

447 [Obulesu and Jhansilakshmi ()] 'Neuroinflammation in Alzheimer's disease: an understanding of physiology and
448 pathology'. M Obulesu , M Jhansilakshmi . *Int J Neurosci* 2014. 124 (4) p. .

- [Pimplikar ()] 'Neuroinflammation in Alzheimer's disease: from pathogenesis to a therapeutic target'. S W Pimplikar . *J Clin Immunol* 2014. 34 p. .
- [Morel et al. ()] 'Neuronal exosomal miRNA dependent translational regulation of astroglial glutamate transporter GLT1'. L Morel , M Regan , H Higashimori , S K Ng , C Esau , S Vidensky . *J Biol Chem* 2013. 288 (10) p. .
- [Yin et al. ()] 'Peroxisome proliferator-activated receptor delta regulation of miR-15a in Ischemia induced cerebral vascular endothelial injury'. K J Yin , Z Deng , M Hamblin , Y Xiang , H Huang , J Zhang . *J Neurosci* 2010. 30 (18) p. .
- [Geldenhuys and Darvesh ()] 'Pharmacotherapy of Alzheimer's disease: current and future trends'. W J Geldenhuys , A S Darvesh . *Expert Rev Neurother* 2015. 15 (1) p. .
- [Matthews et al. ()] 'Physical activity, Mediterranean diet and biomarkers-assessed risk of Alzheimer's: a multi-modality brain imaging study'. D C Matthews , M Davies , J Murray , S Williams , W H Tsui , Y Li . *Adv J Mol Imaging* 2014. 4 p. .
- [Da Silva et al. ()] 'Plasma nutrient status of patients with Alzheimer's disease: systematic review and meta-analysis'. S L Da Silva , B Vellas , S Elemans , J Luchsinger , P Kamphuis , K Yaffe . *Alzheimers Dement* 2014. 10 p. .
- [Bowman et al. ()] 'Plasma omega-3 PUFA and white matter mediated executive decline in older adults'. G L Bowman , H H Dodge , N Mattek , A K Barbey , L C Silbert , L Shinton . *Front Aging Neurosci* 2013. 5 (92) p. .
- [Davidson et al. ()] 'Polyunsaturated fatty acids modulate carcinogen-directed non-coding microRNA signatures in rat colon'. L A Davidson , N Wang , M Shah , J R Lupton , I Ivanov , R S Chapkin . *Carcinogenesis* 2009. 30 (12) p. .
- [Banerjee et al. ()] 'Pomegranate polyphenolics suppressed azoxymethane-induced colorectal aberrant crypt foci and inflammation: possible role of miR-126/VCAM-1 and miR-126/PI3K/AKT/mTOR'. N Banerjee , H Kim , S Talcott , S Mertens-Talcott . *Carcinogenesis* 2013. 34 (12) p. .
- [Chiang ()] 'Post-harvest consideration factors for microRNA research in cellular, tissue, serum and plasma samples'. Vsc Chiang . *Cell Biol Int* 2014. 38 (12) p. .
- [Serhan ()] 'Pro-resolving lipid mediators are leads for resolution physiology'. C N Serhan . *Nature* 2014. 510 p. .
- [Joilin et al. ()] 'Rapid regulation of microRNA following induction of long-term potentiation in vivo'. G Joilin , D Guevremont , B Ryan , C Claudianos , A S Cristino , W C Abraham . *Front Mol Neurosci* 2014. 7 (98) p. .
- [Witwer et al. ()] 'Real-time quantitative PCR and droplet digital PCR for plant miRNAs in mammalian blood provide little evidence for general uptake of dietary miRNAs'. K W Witwer , M A McAlexander , S E Queen , R J Adams . *RNA Biol* 2013. 10 (7) p. .
- [Wang et al. ()] 'Regulation of retinal inflammation by rhythmic expression of miR-146a in diabetic retina'. Q Wang , S Bozack , Y Yan , M E Boulton , M B Grant , J Busik . *Invest Ophthalmol Vis Sci* 2014. 13.
- [Sun et al. ()] 'Role of miR-181 family in regulating vascular inflammation and immunity'. X Sun , A Sit , M W Feinberg . *Trends Cardiovasc Med* 2014. 24 p. .
- [Kosaka et al. ()] 'Secretory mechanisms and intercellular transfer of microRNAs in living cells'. N Kosaka , H Iguchi , Y Yoshioka , F Takeshita , Y Matsuki , T Ochiya . *J Biol Chem* 2010. 285 (23) p. .
- [Corsi et al. ()] 'Supplementation of omega 3 fatty acids improves oxidative stress in activated BV2 microglial cell line'. L Corsi , B M Dongmo , R Avallone . *Int J Food Sci Nutr* 2015.
- [Swerdlow et al. ()] 'The Alzheimer's disease mitochondrial cascade hypothesis: Progress and perspectives'. R H Swerdlow , J M Burns , S M Khan . *Biochim Biophys Acta* 2014. 1842 p. .
- [Karran et al. ()] 'The amyloid cascade hypothesis for Alzheimer's disease: an appraisal for the development of therapeutics'. E Karran , M Mercken , Strooper De . *Nat Rev Drug Discov* 2011. 10 p. .
- [Li et al. ()] 'The association between single nucleotide polymorphisms of GSK-3beta gene and sporadic Alzheimer's disease in a cohort of southern Chinese Han population'. J Li , F Zeng , J Deng , J Zhu , Z Li , T Liu , J . *Neurotox Res* 2014. 26 p. .
- [De Waal et al. ()] 'The effect of Souvenaid on functional brain network organisation in patients with mild Alzheimer's disease: a randomised controlled study'. H De Waal , C J Stam , M M Lansbergen , R L Wieggers , Pjgh Kamphuis , P Scheltens . *PLoS ONE* 2014. 9 (1) p. e86558.
- [Hooijmans et al.] *The effects of long-term omega-3 fatty acid supplementation on cognition and Alzheimer's pathology in animal models of*, C R Hooijmans , -De Pasker , Pcm Jong , Rbm De Vries , M Ritskes-Hottinga .

504 [Wang et al. ()] ‘The expression of microRNA miR-107 decreases early in Alzheimer’s disease and may accelerate
505 disease progression through regulation of beta-site amyloid precursor proteincleaving enzyme 1’. W X Wang
506 , B W Rajeev , A J Stromberg , N Ren , G Tang , Q Huang . *J Neurosci* 2008. 28 (5) p. .

507 [Piantadosi and Suliman ()] ‘Transcriptional control of mitochondrial biogenesis and its interface with inflam-
508 matory processes’. C A Piantadosi , H B Suliman . *Biochim Biophys Acta* 2012. 1820 p. .

509 [Taghizadeh et al. ()] ‘Vitamin D supplementation restores suppressed synaptic plasticity in Alzheimer’s disease’.
510 M Taghizadeh , S A Talaei , A Djazayeri , M Salami . *Nutr Neurosci* 2014. 17 (4) p. .