



GLOBAL JOURNAL OF MEDICAL RESEARCH: K
INTERDISCIPLINARY
Volume 21 Issue 6 Version 1.0 Year 2021
Type: Double Blind Peer Reviewed International Research Journal
Publisher: Global Journals
Online ISSN: 2249-4618 & Print ISSN: 0975-5888

Monitoring and Mapping of Insecticide Resistance in Vector of Cutaneous Leishmaniasis, *Phlebotomus Papatasi* (Diptera: Psychodidae) in Iran

By Maryam Molaezadeh, Amrollah Azarm, Mohammad Nasrabadi,
Fateme Shahidi, Faramarz Bozorgomid & Hassan Vatandoost

Tehran University of Medical Sciences

Abstract- Sandfly, *Phlebotomus papatasi* is an important vector of zoonotic cutaneous leishmaniasis (ZCL) that plays the main role in the transmission of leishmaniasis in Iran. ZCL is one of the most common endemic diseases in Iran. The prevalence of resistance to insecticides in vector species around worldwide is a serious threat to the fight against vector-borne diseases. To provide authentic information about this novel, the reliable data on academic resources such as Google Scholar, Scopus, Web of Science, Springer, Pro-Quest, Wiley Online, Science Direct, Research Gate, PubMed, Sage, and SID were used. There are some levels of resistance in some parts of Iran like, Lorestan and Isfahan province. Resistance to DDT 4% was observed in some parts of Iran. In Lorestan province -Pol-e Dokh-tar, Rumeshgan, and Kuhdasht districts- samples collected in the form of hand catch and indoors showed resistance to DDT 4%. Studies in the rural district of Badrood, Natanz County, Esfahan province showed resistance to DDT 4%.

Keywords: *phlebotomus papatasi, insecticide, resistance, leishmaniasis, Iran.*

GJMR-K Classification: NLMC Code: WR 345



Strictly as per the compliance and regulations of:



Monitoring and Mapping of Insecticide Resistance in Vector of Cutaneous Leishmaniasis, *Phlebotomus Papatasi* (Diptera: Psychodidae) in Iran

Maryam Molaezadeh ^α, Amrollah Azarm ^σ, Mohammad Nasrabadi ^ρ, Fatemeh Shahidi ^ω,
Faramarz Bozorgomid [¥] & Hassan Vatandoost [§]

Abstract- Sandfly, *Phlebotomus papatasi* is an important vector of zoonotic cutaneous leishmaniasis (ZCL) that plays the main role in the transmission of leishmaniasis in Iran. ZCL is one of the most common endemic diseases in Iran. The prevalence of resistance to insecticides in vector species around worldwide is a serious threat to the fight against vector-borne diseases. To provide authentic information about this novel, the reliable data on academic resources such as Google Scholar, Scopus, Web of Science, Springer, Pro-Quest, Wiley Online, Science Direct, Research Gate, PubMed, Sage, and SID were used. There are some levels of resistance in some parts of Iran like, Lorestan and Isfahan province. Resistance to DDT 4% was observed in some parts of Iran. In Lorestan province -Pol-e Dokh-tar, Rumesghar, and Kuhdasht districts-samples collected in the form of hand catch and indoors showed resistance to DDT 4%. Studies in the rural district of Badrood, Natanz County, Esfahan province showed resistance to DDT 4%. The *Phlebotomus papatasi* is susceptible to other insecticides like permethrin 0.75%, deltamethrin 0.1%, cyfluthrin 0.15% and Lambda-cyhalothrin 0.05%. According to a recent study, *Ph. papatasi* showed resistance to some kind of pesticides. Constant monitoring, having a map of insecticide resistance can be an alert for the health system and is a good guide for vector disease control. Furthermore, guidelines are needed for monitoring and evaluation of insecticide susceptibility tests against sand flies.

Keywords: *phlebotomus papatasi*, insecticide, resistance, leishmaniasis, Iran.

1. INTRODUCTION

Leishmaniasis is one of the most important communicable diseases between humans and animals transmitted to humans by sand fly species. The prevalence of ZCL in Iran has always been increasing, so that between 2001 and 2005 shows about a 105% increase. The known rural foci of Leishmaniasis have been reported from the villages of East Isfahan, Turkmen Sahara, Natanz, Sarakhs, Lotfabad, Khuzestan, Ilam, Khorasan, Shiraz, and Kashan¹⁻⁵ (Fig.1). Due to the widespread prevalence of ZCL in Iran

and the world, to break the disease transmission chain, appropriate practical approaches are needed, such as the use of various personal protection methods like long-lasting bed nets and insecticide-impregnated curtains, using insect repellents at work, and outdoors, indoor spraying is limited in scale⁶. As part of control programs, sand flies have been exposed to four major classes of synthetic insecticides: Organochlorine, pyrethroids, Organophosphates, and Carbamates. These exposures have been either intentional in directed vector control efforts or have been inadvertent as part of malaria control efforts against *Anopheles*⁷. The prevalence of insecticide resistance in vector species worldwide is a continuous threat for any success at mitigating the spread of vector-borne diseases. Most species of phlebotomine sandflies remain susceptible to insecticides. However, around the world, there is increasing evidence of insecticide resistance.

Author α σ ρ ω ¥ §: Department of Medical Entomology and Vector Control, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran.

Corresponding Author §: Department of Environmental Chemical Pollutants and Pesticides, Institute for Environmental Research, Tehran University of Medical Sciences, Tehran, Iran.

e-mails: hvatandoost1@yahoo.com, vatando@tums.ac.ir



Figure 1: Map of Iran providing the province outlines, in brown the provinces that are endemic for zoonotic cutaneous leishmaniasis(1)

a) Detecting Insecticide Resistance

Managing insecticide resistance requires timely, accurate data through resistance monitoring and insecticide evaluation to assess a vector species' susceptibility to insecticides. The primary way to assess insecticide resistance in many vectors, including sand flies, is to use insecticide susceptibility bioassays. The two most commonly used bioassays worldwide are the WHO exposure kit bioassay and the Centers for Disease Control (CDC) bottle bioassay⁸. The WHO exposure kit bioassay is a standardized protocol that consists of an exposure kit containing tubes lined with filter papers that are impregnated with a specific concentration of an insecticide. The CDC bottle bioassay protocol consists of exposing insects to concentrations of insecticide that are coated on the interior of glass bottles. Both bioassays have been used to assess insecticide resistance in sand flies, but the WHO bioassay is used more frequently⁷.

b) Resistance Mechanisms

Insecticide resistance to synthetic insecticides have been reported in many important insect vectors such as mosquitoes, black flies, Triatomine bugs, lice, fleas, and sand flies. Four mechanisms of resistance are known to exist in insects: reduced penetration, behavior avoidance, target-site insensitivity, and metabolic detoxification. Of the four, target-site insensitivity and metabolic detoxification are the two most geographically and entomologically widespread. Today, there is evidence of target-site insensitivity and metabolic detoxification resistance to the four main classes of synthetic insecticides in all major vector species⁷. The insecticide resistance mechanisms in *Ph. papatasi* have

not been identified, unlike the mechanisms of more intensely studied insects such as mosquitoes and house flies. Numerous susceptibility tests have been carried out in the foci of ZCL in Iran in against *Ph. papatasi*. This results are the reviews on the monitoring and mapping of insecticide resistance in *Ph. papatasi* in Iran.

c) Characteristics of *Ph.papatasi*

Sandflies are tiny insects, 1.5-3.5 mm in length, with a hairy appearance, large black eyes, and long, stilt-like legs. Sandflies can be distinguished from other Diptera, especially members of the Psychodidae family to which these insects belong, by the way, they rest their wings, which look like a V. The Sand-fly *Ph. papatasi* is the well-known vector of zoonotic cutaneous leishmaniasis and sand-fly fever³(2). This species is endemic to most parts of Iran³. The *Ph.papatasi* prefers human habitats rather than other species even though it found in human habitats in mountainous areas⁹(3). Their Resting places are animal and human habitats also that caught from the plains place much more than the mountains. It is found in rodents' nests, rooms, stables, and wall crevices, and in all biotopes. It is interested in heat and humidity as well as this grows well where the groundwater level is high. This species is sensitive to heat but is resistant to rain. In terms of blood-feeding, it is more interested in human, rodent blood and bites several times during feeding to supplement its food¹⁰⁻¹².

d) Distribution of *Ph.papatasi* in Iran

The sand fly *Ph. Papatasi* is widely distributed in Iran. There are in East Azerbaijan, West Azerbaijan, Ardabil, Isfahan, Ilam, Bushehr, Tehran, Chaharmahal Bakhtiari, Khorasan, Khuzestan, Zanjan, Semnan, Sistan

and Baluchestan, Fars, Qom, Kurdistan, Kerman, Markazi, Hormozgan, Hamedan and Yazd^{1,3,14} (Fig.2).
Kermanshah, Golestan, Gilan, Lorestan, Mazandaran, Figure 3 shows the symptoms of ZCL.

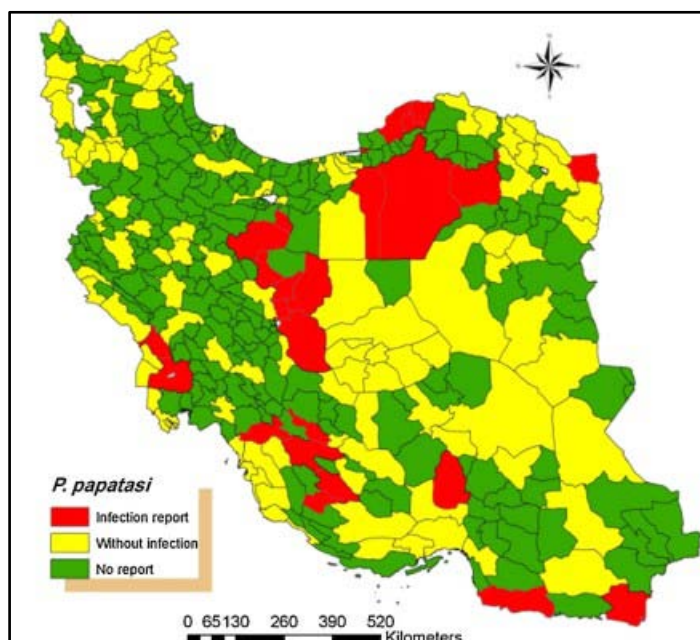


Figure 2: Distribution of *Ph. papatasi* infection to Leishmania parasites in Iran



Figure 3: Symptoms of Zoonotic Cutaneous Leishmaniasis (ZCL)

e) The control management *Ph.papatasi* in Iran

The main vector of the rural type of leishmaniasis is *Ph. papatasi*, which is semi-wild. The most of transmission takes place in an outdoor place, so spraying does not have a significant effect on reducing cases except in the event of epidemics, which may be effective. Although the make use of bed nets, curtains treated by the deltamethrin insecticide with a shelf life of more than five years may lead to the severance of the transmission chain in rural seekers, the covered population must have been properly trained beforehand. Control of sandflies was started using residual insecticides such as DDT, lindane, and aerosols. DDT and lindane were used as emulsions, aqueous suspensions, soapy water suspensions, solutions, and powders. DDT in the form of aerosol was also very effective. DDT and lindane aerosols were mainly used for surface spraying. These compounds were characterized by their lasting effect. Sprayed surfaces retained their insecticidal effect for several weeks and even months after application.

II. METHOD

To provide authentic information about these novel results, we used reliable data on academic

resources such as Google Scholar, Scopus, Web of Science, Springer, Pro-Quest, Wiley Online, Science Direct, Research Gate, PubMed, Sage, and SID.

III. RESULT AND DISCUSSION

A glance at the table number 1 and Fig.4 provided reveals the susceptibility status of *Ph. papatasi* to DDT (4%), permethrin (0.75%), deltamethrin (0.1%), cyfluthrin (0.15%) and lambda-cyhalothrin (0.05%), In four different years 2011, 2013, 2017 and 2020. It has been estimated in the rural district of Badrood, Natanz County, Esfahan province. The results revealed that this species was resistant candidate to DDT but susceptible to other insecticides^{13,15-17}. In a similar study in Lorestan Province-Pol-e Dokh-tar, Rumeshgan, and Kuhdasht districts the results showed that this species was resistant to DDT 4% but susceptible to bendiocarb 0.1%, permethrin 0.75%, deltamethrin 0.05%, and cyfluthrin 0.15%¹⁸⁻²³.

Table 1: Status of insecticide resistance in *Ph. papatasi* in different parts of Iran

Area	Method	Insecticide	Susceptibility status	Ref.	year
Dehbakri County, Esfahan	Aspirator	DDT4%	100	(4)	2011
		Deltamethrin 0.05%	100		
Lorestan Province	Hand catch/indoor/baited traps/outdoor	DDT 4%	R(87.7)	(5)	2020
			S(92.0)		
		Bendiocarb 0.1%	S(93.4)		
			S(94)		
		Permethrin 0.75%	S(92.4)		
			S(97.9)		
		Deltamethrin 0.05%	S (96.8)		
			S (97.8)		
		Cyfluthrin 0.15%	100		
			100		
Arsanjan - Fars province	Aspirator	DDT4%	S (96.7)	(6)	2000
Natanz county, Esfahan	Aspirator	Deltamethrin 0.05%	S (97.86)	(7)	2017
		λ-cyhalothrin 0.05%	S (97.78)		
		Cyfluthrin 0.15%	S (100)		
		Permethrin 0.75%	S (98.7)		
		DDT4%	RC (96.)		
Natanz county, Esfahan	Aspirator	DDT 4%	Female, LT50:1312.66	(8)	2013
			Male, LT50:1200.97		
		Permethrin 0.75%	Female, LT50:2		
			53.66		
			Male, LT50:310.10		
		Deltamethrin 0.1%	Female, LT50:36.47		
			Male, LT50:18.63		
		Cyfluthrin 0.15%	Female, LT50:9.36		
			Male, LT50:6.08		
		λ-cyhalothrin 0.05%	Female, LT50:6		
			Male, LT50 : 0.77		
Natanz county, Esfahan	Aspirator	DDT 4%	Female, LT50:1104.97	(9)	2011

			Male, LT50:973.51		
		Deltamethrin 0.1%	Female, LT50:26.79		
			Male, LT50:4.4,		
		Permethrin 0.75%	Female, LT50:182.35		
			Male, LT50:59.5		
			Female, LT50:1.48		
		λ-cyhalothrin 0.05%	Male, LT50:1.5 s		
		Cyfluthrin 0.15%	Female, LT50:15.42		
			Male, LT50:2.65		
Natanz county, Esfahan	Aspirator	DDT 4%	Female, LT50:641.62	(10)	2020
			Male, LT50:439.28		
		Deltamethrin 0.1%	Female, LT50:146.44		
			Male, LT50:97.75		
		Permethrin 0.75%	Female, LT50:136.15		
			Male, LT50:108.90		
		λ-cyhalothrin 0.05%	Female, LT50:72.69		
			Male, LT50:57.84		
		Cyfluthrin 0.15%	Female, LT50:8.71		
			Male, LT50:5.00		
KoohSangi, Mashhad	Aspirator	DDT 4%	100	(11)	1985
Khajeh Rabi, Mashhad	Aspirator	DDT 4%	100	(11)	1986
Shahporabad, Isfahan	Aspirator	DDT 4%	75	(11)	1985
Komshechreh, Isfahan	Aspirator	DDT 4%	76	(11)	1985
Komshechreh, Isfahan	Aspirator	DDT 4%	100	(11)	1986
Omkolsum, Khuzestan	Aspirator	DDT 4%	100	(11)	1986
Susangerd, Khuzestan	Aspirator	DDT 4%	100	(11)	1986
Natanz county, Esfahan	bed nets	Deltamethrin 0.1%	100	(12)	2006

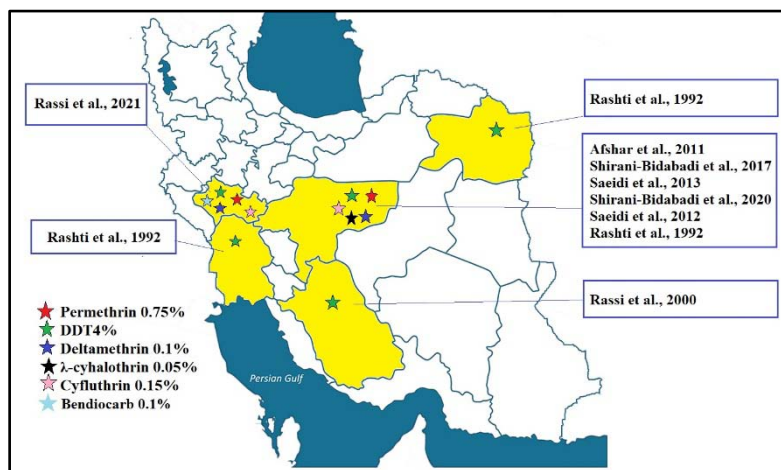


Figure 4: Status of insecticide resistance in *Ph. papatasi* in different parts of Iran

IV. CONCLUSION

Constant monitoring, having a map of insecticide resistance in Iran can be alert for the health system and is a good guide for vector disease control. Furthermore, guidelines is needed for monitoring and evaluation of insecticide susceptibility tests against sand flies.

Declarations: All the author declare that there is no conflict of interest.

Statements on the authors' contributions: All the authors were involved.

Funding: This research is financially supported by by Ministry of Health and Medical Education under code number of NIMAD 995633

Conflict of Interest: All authors declare that there is no conflict of interest.

Ethical approval: Not applicable

Acknowledgments: Ministry of Health and Medical Education of Iran for support.

REFERENCES RÉFÉRENCES REFERENCIAS

1. Sofizadeh A, Cherabin M, Mehravaran A. 2013. Cutaneous leishmaniasis in GonbadKavoos, North

- of Iran (2009-11): an epidemiological study. *Journal of Gorgan University of Medical Sciences*, 14(4), pp. 100-106.
2. Sabzevari S, Teshnizi SH, Shokri A. 2021. Cutaneous leishmaniasis in Iran: A systematic review and meta-analysis. *Microbiology and Pathology*, 104721.
3. Yaghoobi-Ershadi MR. 2016. Control of phlebotomine sand flies in Iran: a review article. *Journal of Arthropod-borne Diseases*, 10(4), pp. 429-436.
4. Yaghoobi-Ershadi M, Marvi-Moghadam N, Jafari R, et al. (2015) Some epidemiological aspects of cutaneous leishmaniasis in a new focus, central Iran. *Dermatology Research and Practice*. <https://doi.org/10.1155/2015/286408>
5. Yaghoobi-Ershadi M 2012. Phlebotomine sand flies (Diptera: Psychodidae) in Iran and their role on Leishmania transmission. *Journal of Arthropod-borne Diseases*, 6(1), pp.1-10.
6. Pener H, Wilamovsky A. 1987. Base-line susceptibility of *Phlebotomus papatasi* to insecticides. *Medical and Veterinary Entomology*, 1(2), pp. 147-149.
7. Denlinger DS. 2017. Understanding the mechanisms of insecticide resistance in *Phlebotomus papatasi* and *Lutzomyia longipalpis* sand flies (Diptera: Psychodidae: Phlebotominae): *Utah State University*, pp.258.
8. Berríos-Torres SI, Umscheid CA, Bratzler DW, et al. (2017). Centers for disease control and prevention guideline for the prevention of surgical site infection, *JAMA Surgery*, 152(8), pp.784-91.
9. Karimi A, Hanafi-Bojd AA, Yaghoobi-ErshadiMR, et al. 2014. Spatial and temporal distributions of phlebotomine sand flies (Diptera: Psychodidae), vectors of leishmaniasis, in Iran. *ActaTropica*, 132, pp.131-139.
10. Abedi-Astaneh F, Hajjaran H, Yaghoobi-ErshadiMR, et al. Risk mapping and situational analysis of cutaneous leishmaniasis in an endemic area of Central Iran: a GIS-based survey. *PLoS One*. 2016, 11(8), e0161317.
11. Yaghoobi-ErshadiMR, Zahraei-Ramazani AR, Akhavan AA, et al. Rodent control operations against zoonotic cutaneous leishmaniasis in rural Iran. *AnnSaudi Med*.2005, 25(4), 309-12.
12. Abedi-Astaneh F, Akhavan AA, ShirzadiMR, et al. 2015. Species diversity of sand flies and ecological niche model of *Phlebotomus papatasi* in central Iran. *ActaTropica*, 149, pp.246-453.
13. Azizi K, Rassi Y, Javadian E, et al. 2006.*Phlebotomus (Paraphlebotomus) alexandri*: a probable vector of *Leishmania infantum* in Iran. *Annals of Tropical Medicine and Parasitology*, 100(1), pp.63-68.
14. Hanafi-Bojd AA, Yaghoobi-Ershadi MR, Haghdoost AA, et al. 2015. Modeling the distribution of cutaneous leishmaniasis vectors (Psychodidae: Phlebotominae) in Iran: a potential transmission in disease prone areas. *Journal of Medical Entomology*, 52(4), pp.557-65.
15. Shirani-Bidabadi L, Zahraei-Ramazani A, Yaghoobi-ErshadiMR, et al. 2017. Assessing the insecticide susceptibility status of field population of *Phlebotomus papatasi* (Diptera: Psychodidae) in a hyperendemic area of zoonotic cutaneous leishmaniasis in Esfahan Province, Central Iran. *ActaTropica*, 176, pp.316-322.
16. Saeidi Z, Vatandoost H, Akhavan A, et al. 2013. Baseline insecticide susceptibility data of *Phlebotomus papatasi* in Iran. *Journal of Vector borne Diseases*, 50(1), pp.57-65.
17. Shirani-Bidabadi L, Zahraei-Ramazani AR, Yaghoobi-ErshadiMR, et al. 2020. Monitoring of laboratory reared of *Phlebotomus papatasi* (Diptera: Psychodidae), main vector of zoonotic cutaneous leishmaniasis to different imagicides in hyper endemic areas, Esfahan province, Iran. *Journal of Arthropod-borne Diseases*, 14(1), pp.116-126.
18. Rassi Y, Kayedi MH, AbaiMR, et al. 2021. Efficiency of two capture methods providing live sand flies and assessment the susceptibility status of *Phlebotomus papatasi* (Diptera: psychodidae) in the foci of cutaneous leishmaniasis, Lorestan Province, western Iran. *Journal of Arthropod-borne Diseases*. <https://doi.org/10.18502/jad.v14i4.5278>
19. Afshar AA, Rassi Y, Sharifi I, et al. 2011. Susceptibility status of *Phlebotomus papatasi* and *Ph. sergenti* (Diptera: Psychodidae) to DDT and deltamethrin in a focus of cutaneous leishmaniasis after earthquake strike in Bam, Iran. *Iranian Journal of Arthropod-borne Diseases*, 5(2),pp.32-40.
20. Rassi Y, Jalali M, VatandoostH.2000.Susceptibility status of *Phlebotomus papatasi* to DDT in Arsanjan country in Fars province, Iran. *Iranian Journal of Public Health*, 21-6.
21. Saeidi Z, Vatandoost H, Akhavan AA, et al. 2021. Baseline susceptibility of a wild strain of *Phlebotomus papatasi* (Diptera: Psychodidae) to DDT and pyrethroids in an endemic focus of zoonotic cutaneous leishmaniasis in Iran. *Pest Management Science*, 68(5), pp.669-675.
22. Rashti MS, Panah HY, Mohamadi HS, et al.1992. Susceptibility of *Phlebotomus papatasi* (Diptera: Psychodidae) to DDT in some foci of cutaneous leishmaniasis in Iran. *Journal of American Mosquito Control Association*, 2,8(1),pp.99-100.
23. Yaghoobi-Ershadi M, Moosa-Kazemi S, Zahraei-Ramazani A, et al.2006. Evaluation of deltamethrin-impregnated bed nets and curtains for control of zoonotic cutaneous leishmaniasis. *Bulletin of Society of Pathology and Exotic*, 99(1), pp.43-48.