

The Impact of Senna Siamea (Lam) Leaves Extracts on 2, 2-Dichlorovinyl dimethyl Phosphate Induced Brain Oxidative Stress in Wistar Albino Rats

Oni, Olaiya Peter¹, Oseni, Olatunde Abass² and Okoh, Olayinka Sunday³

¹ Ekiti-State University

Received: 12 June 2021 Accepted: 30 June 2021 Published: 15 July 2021

Abstract

Organophosphate compounds have been a common source of mortality in recent times. A typical example is dichlorvos [2, 2-Dichlorovinyl dimethyl phosphate (DDVP)]; an important agricultural pesticide. This study sought to investigate the phytochemical screening, in-vitro antioxidant property of the leaf solvents extracts and ameliorating effects of Senna siamea in DDVP-induced brain oxidative stress in Wistar rats. The aqueous, methanol and ethanol extracts of the leaves were obtained and analyzed for their in-vitro antioxidant parameters. Thirty-two healthy Wistar albino rats were grouped into eight of 4 rats each weighing between 140-150g. The animals were orally administered with 6.6mg/kg body weight of DDVP except groups 1 and 2 followed by treatments with the three solvents extracts respectively that lasted for four weeks.

Index terms— Senna siamea; brain toxicity; oxidative stress; antioxidant properties; ?-glutamyl transferase.

1 Introduction

Exposure of human beings to poisons and some other toxic materials has been responsible for many mortality cases in our generation most of which are accidental. Reports indicate that nothing less than 200,000 were dead as a result of organophosphate compound, one of which is 2, 2-Dichlorovinyl dimethyl phosphate (DDVP) [1]. The organophosphate compounds is unarguably one of the toxic and adequately chronic organophosphates that is of detrimental effect to the health of humans and animal [2]. It was said that the continuous exposure to humans and animals to DDVP has been identified as one of the leading causes of acetyl cholinesterase (AChE) inhibition especially at the presynaptic cleft, thus leading to the accumulation of acetylcholine as well as the triggering of postsynaptic neurons in animals, and ultimately to death [3]. Notably, the exposure of humans and animals have been fingered to be a key player in respiratory problems including that of discomfort in the chest, bloody or running nose, severe and sometimes dry coughing, difficulty in breathing and increased fluid in the bronchial tubes [4]. Oxidative stress arising from free radicals like reactive oxygen species (ROS) now appears to be a fundamental mechanism underlying many degenerative diseases such as diabetes, viral infection, auto-immune pathologies and probably aging. Evidence suggests that ROS can be scavenged through chemoprevention utilizing antioxidant compounds present in foods and medicinal plants [5]. Plants play a significant role in maintaining human health and improving the quality of human life. They serve humans as valuable components of food, cosmetics, dyes, and medicines. The World Health Organization estimated that 80% of the earth's inhabitants rely on traditional medicine for their primary healthcare needs, and most of this therapy involves the use of plant extracts and their active components [6]. Senna siamea has a long history of use as a folk-medicine and its therapeutic efficacy is well recognized. Different parts of *S. siamea* can be used for various medical purposes [7;8;9]. The fruit is used to charm away intestinal worms and to prevent convulsions in children. The heartwood is said to be a laxative, and a decoction is used against scabies [10]. Senna siamea also known as Siamese cassia,

43 cassod tree, cassod tree and cassia tree is a legume in the subfamily Caesalpinioideae. It is native to South and
44 Southeast Asia, widespread in Africa, although its exact origin is unknown¹¹. This plant has proven to contain
45 some important biochemical components like alkaloids, volatile essential oils, phenols and phenolic glycosides,
46 resins, oleosins, steroids, tannins and terpenes. Senna siamea is a medicinal plant acknowledged to be rich in
47 phenolics, consisting of condensed tannin and phlobatannin, Gallic acid, protocatechuic acid, pyrocatechol, (+)-
48 catechin, (-) epi-gallocatechin-7gallate and (-) epigallocatechin-5, 7-digallate^{9,12}. Its antioxidative property
49 and ameliorating effects on organophosphate toxicity has not been done. This current study however tends to
50 investigate the effects of Senna siamea leaf extracts on some biochemical indices in 2, 2-dichlorovinyl dimethyl
51 phosphate (DDVP) induced brain toxicity using Wistar albino rats.

52 2 II.

53 3 Materials and Methods

54 4 a) Collection and Extraction of the Senna siamea leaf

55 The fresh leaves of Senna siamea (Lam) Irwin & Barneby (Fabaceae) were obtained from Ifaki-Ekiti community,
56 Ekiti State and was authenticated at Department of Plant Science, Ekiti State University, Ado-Ekiti and the
57 plant specimen was preserved with Herbarium numbers (UHAE 2020055). The leaves were rinsed with water and
58 then air-dried by spreading them on a clean surface at room temperature in the laboratory. The air-dried leaves
59 were then pulverized and three major separate extractions were carried out with two hundred grams portions
60 each of the dried powdered leaves soaked in 500mL each of water, ethanol and methanol as solvents to obtain
61 three different extracts. The extracts were then concentrated by increased surface area evaporation to obtain
62 dried extracts for analyses.

63 5 b) Phytochemical screening and in-vitro anti-oxidant param- 64 eters determination of the leaf's extracts

65 The qualitative phytochemical screening [flavonoids, saponin, phlobatannins, terpenoids, Salkowski test for car-
66 diac glycosides (steroidal ring or terpenoids), Keller-Killani test for cardiac glycosides (deoxysugar), Lieberman's
67 test for steroidal nucleus and test for tannins] of aqueous, methanol and ethanol extracts of the leaves were
68 carried out according to the methods of^{13,14} to identify the active constituents while the in-vitro antioxidants
69 properties were determined by the following methods: a. Hydrogen Peroxide Scavenging Effects: The ability of
70 the leaf extracts to scavenge hydrogen peroxide was assessed by the method of Ruch et al¹⁵.

71 b. ABTS Scavenging Effects: The antioxidant effect of the leaf extracts was studied using ABTS (2,2'-
72 azinobis-3-ethyl benzthiazoline-6-sulphonic acid) radical cationide colourisation assay according to the method of
73 Shirwaikar et al¹⁶.

74 c. Measurement of Nitric Oxide Scavenging Activity:

75 The extent of inhibition of nitric oxide radical generation in vitro was followed as per the method reported by
76 Green et al¹⁷.

77 d. DPPH spectrophotometric assay: The free radical scavenging activities of the samples by DPPH method
78 were determined according to the method reported by Brand-Williams et al¹⁸.

79 e. Measurement of Superoxide Scavenging Activity:

80 The superoxide scavenging ability of the extracts was assessed by the method of Winter bourn et al¹⁹.

81 f. Estimation of Total Phenols: The total phenolic content was determined according to a well-cited protocol
82²⁰.

83 g. Estimation of Flavonoids: The total flavonoid contents in the samples were determined following the method
84 reported by Zhishen et al²¹.

85 6 c) Animal management

86 Thirty-two (32) healthy albino Wistar rats were obtained and housed in the animal house of the College of
87 Medicine, Ekiti State University, Ado-Ekiti, Nigeria. The animals were acclimatized for two weeks before
88 administration of DDVP. The acclimatization was done under standard environmental conditions of good lighting,
89 moderate temperature and adequate ventilation. They were also fed on standard rat feed containing adequate
90 proteins, carbohydrate, fats, vitamins, minerals back up with clean and adequate water. The animals were
91 handled under standard laboratory protocols as stipulated by the Institutional Animal Care and Use Committee
92²².

93 7 d) Experimental design

94 The animals were divided into six groups according to their weights with Groups 2 having 3 subgroups, one for
95 each of the three extracts. Each group had four animals. The animals were orally administered with 0.5mL of
96 6.6mg/kg body weight of 500 folds dilution of DDVP solution for two weeks except for Groups 1 and 2 followed

97 by treatments with 0.5mL of 3.3mg/kg body weight of each extract of the plant for another two weeks of the
98 four weeks study.

99 **8 Group 1 Normal Control**

100 Group 2 Extract control (Each subgroup animal was given 0.5mL of 3.3mg/kg body weight of 0.5g/100mL of
101 each solvent (aqueous, methanolic and ethanolic) extract of the plant) Group 3 DDVP control (animals were
102 administered orally with 0.5mL of 6.6mg/kg body weight of 500 folds dilution of DDVP solution to induce brain
103 toxicity) Group 4 DDVP induced animals + 0.5mL of 3.3mg/kg body weight of 0.5g/100mL aqueous extract of
104 the plant.

105 Group 5 DDVP induced animals + 0.5mL of 3.3mg/kg body weight of 0.5g/100mL methanolic extract of the
106 plant.

107 Group 6 DDVP induced animals + 0.5mL of 3.3mg/kg body weight of 0.5g/100mL ethanolic extract of the
108 plant.

109 All animals in the groups were also given rats feed and drinking water ad libitum.

110 **9 e) Preparation of serum and brain homogenate**

111 At the end of the experiment, the rats were chloroform anesthetized and quickly dissected with their blood samples
112 and brain removed. 10% of the brain homogenate was prepared in 6.7nM potassium phosphate buffer (pH 7.4)
113 using the Top driven electric homogenizer. The homogenate was centrifuged at 3,000rpm for 10 minutes at 4 0 C to
114 obtain a clear supernatant while serum sample was prepared from the whole blood collected from the heart into the
115 plain sample bottle and centrifuged at 3,000 rpm after coagulation. The individual serum and brain homogenate
116 were used for measurement of the studied biochemical parameters. The lipid peroxidation was done by measuring
117 the TBARS in accordance with the modified method of Uteley et al 23 ; GGT activity was determined using
118 standard Sigma-Aldrich 24 kit from USA while the antioxidants enzymes activities [Catalase (CAT), superoxide
119 dismutase (SOD), Glutathione-S-transferase (GST), Glutathione reductase (GR) and Glutathione peroxidase
120 (GPx)]; reduced glutathione (GSH) were determined by the methods described by Chance and Maehly 25

121 **10 f) Statistical analyses**

122 The results obtained were evaluated using the statistical test of Means triplicates results of four animals per
123 group.

124 **11 III.**

125 Results IV.

126 **12 Discussion**

127 The Phytochemical assessment of the three solvents extracts of the leaves of *Senna siamea* contained some plant
128 chemicals of interest and important medicinal potentials as shown in Figure 1 The three extracts of the leaves
129 showed appreciable scavenging potentials for DPPH radicals in a concentration dependent manner. indicating
130 that the higher the concentration used, the higher the scavenging activity with the highest scavenging activity
131 found in methanol extract. The ability of the plant to freely scavenge DPPH radicals may be due to the presence of
132 flavonoids 35 . The scavenging of DPPH radical by antioxidants agents is due to the reaction between antioxidant
133 molecules and radical progress which results in the scavenging of the radicals by hydrogen donation. It is visually
134 noticeable as a change in colour from purple to yellow. Hence, DPPH is usually used as a substance to evaluate
135 the antioxidant potential of medicinal plants 36 . The Ethanolic extract of *Senna siamea* displayed a more
136 superior superoxide scavenging activity at the 1mg/mL concentration which is far greater than the results for
137 both aqueous and methanolic extracts at the same concentration. The super oxide scavenging activity of *Senna*
138 *siamea* can be seen in its ability to scavenge super oxide radical ions to form stable radicals and by such can
139 help in the termination of radical chain reaction 37 .In the hydrogen peroxide scavenging activity, the ethanolic
140 extracts possess the highest activity than those aqueous and methanol. However, Shaluetal 38 reported a similar
141 observation for the plant in their study. Total phenols and total flavonoids were also observed to be considerably
142 present in all the plant extracts. The total phenol aqueous extract had the highest value followed closely by the
143 ethanolic and methanolic extracts concentrations. These results obtained for *Senna siamea* showed resemblance
144 to the work of Jyotietal 39 on *Acacia nilotica*. Phenols are said to contribute to the quality and nutritional value
145 in terms of modifying aroma, color, taste and flavor 39 . Phenolic compounds could be a major determinant of
146 antioxidant potentials of food plants and could therefore be a natural source of antioxidants 40 . In this study, the
147 results obtained for total flavonoids showed that ethanolic extract contained the highest level of flavonoids than
148 aqueous and methanolic extracts. Flavonoids are part of the secondary metabolites present in plants as part of
149 its arsenal and has been reported by Choudharyetal 40 that flavonoids show some antioxidant activity and that
150 it has considerable effects on both human's health and nutrition. He described its mechanisms as the one with
151 either scavenging or chelating process. They are said to possess hydroxyl groups which prompted their radical
152 scavenging effects in the plant. Figure ??0 showed the effects of the leaf extracts on the level of lipid peroxidation

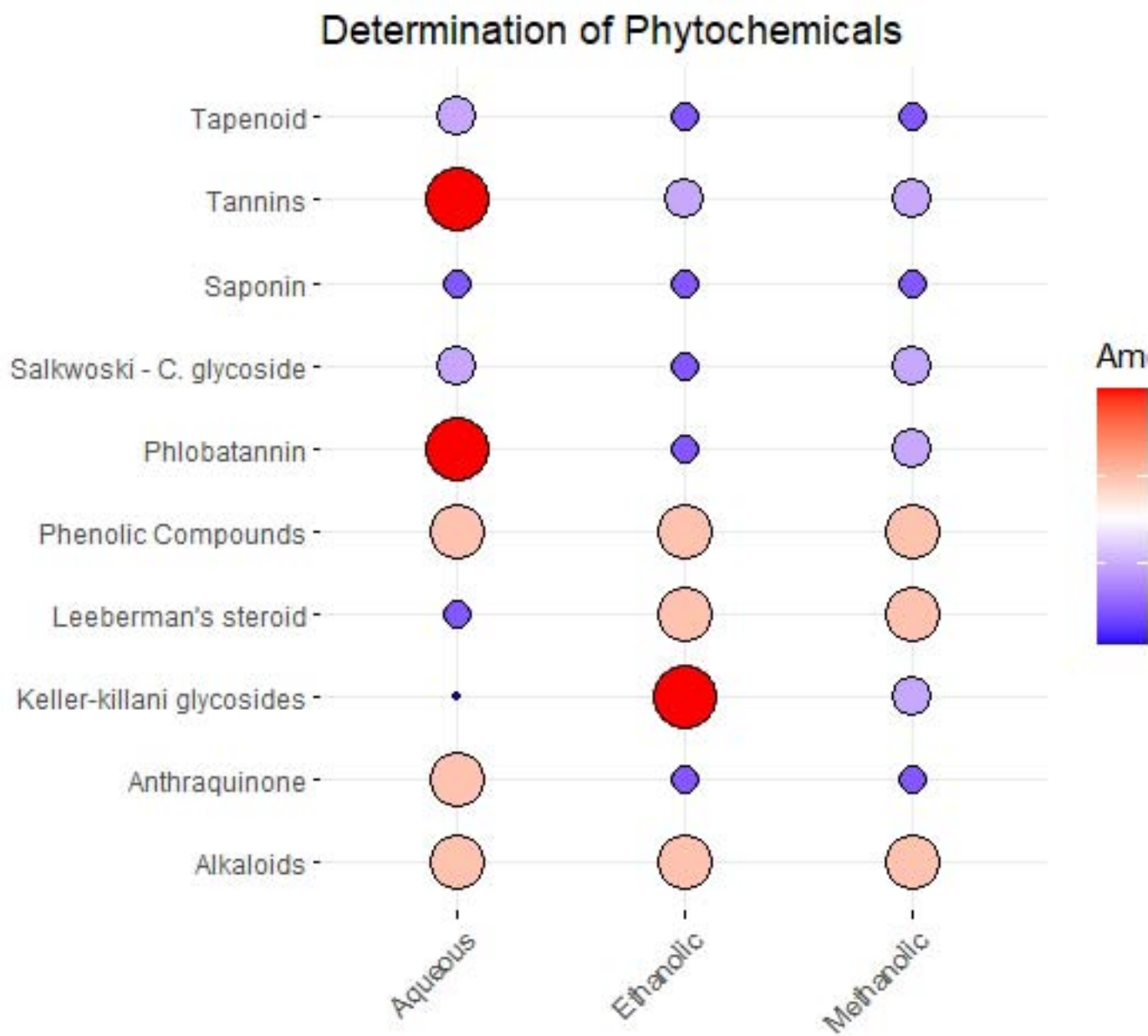
153 both in the serum and brain of the studied animals which revealed that the DDVP induced control group 3 caused
154 almost tenfold elevation in the malondialdehyde concentration of both serum and brain tissue. The treatment
155 of the rats with the three solvents extracts showed that both extracts produced ten-fold reduction effects in
156 the concentration of malondialdehyde in both serum and brain tissues when compared with the DDVP-induced
157 group. It has been reported that enhanced lipid peroxidation leads to tissue damage and failure of antioxidant
158 defense mechanism to prevent formation of excessive free radicals 41 . It is clearly evidenced in this study that
159 treatments of the induced animals with the aqueous, methanolic and ethanolic extracts of the Senna siamea leaf
160 respectively caused a substantive decrease in the level of lipid peroxidation of both serum and brain tissues. This
161 result however corroborated the observation of Ojo et al 32 who postulated that DDVP induction would lead to
162 an increased MDA value and the treatment with antioxidant containing extracts would lead to the reduction of
163 the MDA value. The induction of DDVP in group 3 from Figure 4.0 showed a significantly increase in the value
164 of GGT which thus implies that the exposure caused a damage to the brain including increased permeability,
165 possible neurosis in the brain. This result also showed that the treatment with the various solvents extracts caused
166 a significant decrease in the level of the GGT indicating that the extracts of Senna siamea caused reversal to the
167 detrimental DDVP induction in the animals. This is in line with the results of Ojo et. al 32 who also postulated
168 that DDVP induction will lead to an increased GGT value and the treatment with a good antioxidant plant like
169 Senna siamea will reverse back the damage caused by the exposure to DDVP; it should also be noted in this study
170 that all the solvents extracts of the plant produced tremendous attenuating effect of the brain toxicity. Figures
171 ??a and 25b B glutathione transferase, glutathione reductase) and reduced glutathione. Oxidative stress plays
172 a major role in the pathogenic of many disorders including aging, cancer, diabetes, Alzheimer's, strokes, viral
173 infections (that cause airway epithelial inflammation), neurodegenerative processes (including cell death, motor
174 neuron diseases and axonal injury) and infraction, and brain edema. Antioxidant enzyme plays an important
175 role in protecting oxidative injury to the body. One of the therapeutic approaches by which these disorders can
176 be prevented is to increase the levels of these antioxidant enzymes 42 . The catalase activity was significantly
177 increased at ($p < 0.05$) in the aqueous, methanolic and ethanolic treatments groups towards the control and extract
178 treated groups when compared with the DDVP induced group. Other researchers have also reported an upward
179 trend in catalase level using various plants extract treatments in other complications as we observed in this study
180 43,44,45,46 . Catalase are hemecontaining enzymes that convert hydrogen peroxide (H_2O_2) to water and O_2
181 , and they are largely localized in subcellular organelles such as peroxisomes 47 . The %SOD activity observed in
182 this study showed a significant ($p < 0.05$) increase in the various treatment's groups after a gross reduction by the
183 DDVP induced group when compared with the normal control and extracts treated groups. Similar observations
184 were also reported in earlier studies of (Oseni et al 43 ; Uroko et al 44 ; Onoja et al 45 and Sani et al 46). SOD
185 is the antioxidant enzyme that catalysed the dismutation of the highly reactive superoxide anion to O_2 and to
186 the less reactive species H_2O_2 ; the peroxide can then be destroyed by Catalase or glutathione peroxidase
187 reactions as reported by 48,49,50 . Glutathione peroxidase, glutathione transferase and glutathione reductase in
188 addition with SOD are antioxidants enzymes that work in synergy to protect the organism from reactive oxygen
189 species (ROS). These enzymes were observed to be significantly ($p < 0.05$) increased in all the treatment groups
190 when compared with the induced group to reverse the effects of induction towards the normal control group.
191 Our observation is in consonance with what was reported by 51,52 in their various studies on amelioration of
192 thioacetamide induced oxidative stress and hepatic damage in albino rats by Solanum trilobatum and antioxidant
193 effect of grapevine leaf extract on the oxidative stress induced by a high-fat diet in rats respectively. Reduced
194 glutathione (GSH) is another compound that play a vital role as an antioxidant. Senna siamea solvents extracts
195 were found to significantly ($p < 0.05$) reverse the reduced GSH in DDVP induced rats to the normal control as
196 observed in this study. Reduced glutathione is found in high concentrations in cellular systems and plays a major
197 role in detoxication of various electrophilic compounds, deficiency of which puts the cell at risk for oxidative
198 damage. Previous works have also shown that medicinal plants extracts have abilities to enhance glutathione
199 concentration to reverse the effects of oxidative stress 51,53 .
200 V.

201 13 Conclusion

202 This study has reasonably showed that the oral exposure of the rats to DDVP (Dichloros) caused the brain
203 oxidative stress in the Wistar albino rats as indicated by the increased level of lipid peroxidation, increased GGT
204 activity, reduced antioxidant potentials both in the serum and brain while the aqueous, methanolic and ethanolic
205 extracts of Senna siamea showed a significant and protective effects against the action of DDVP induced oxidative
206 stress in the rats.

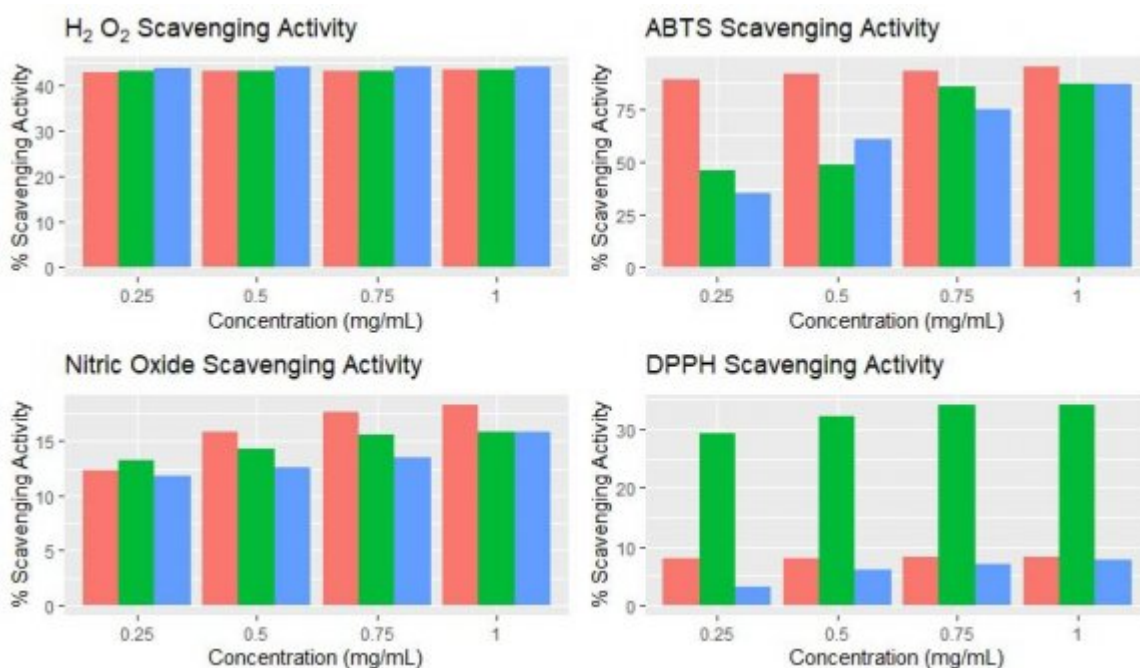
207 The plant studied here will in no doubt do well as a neurotoxicity protective agents and further researches
208 needed to be carried out to explore it for raw materials needed for the treatment of neurodegenerative diseases
209 like Alzheimer's diseases and other complications. ¹

¹© 2021 Global Journals The Impact of Senna Siamea (Lam) Leaves Extracts on 2, 2-Dichlorovinyl dimethyl Phosphate Induced Brain Oxidative Stress in Wistar Albino Rats



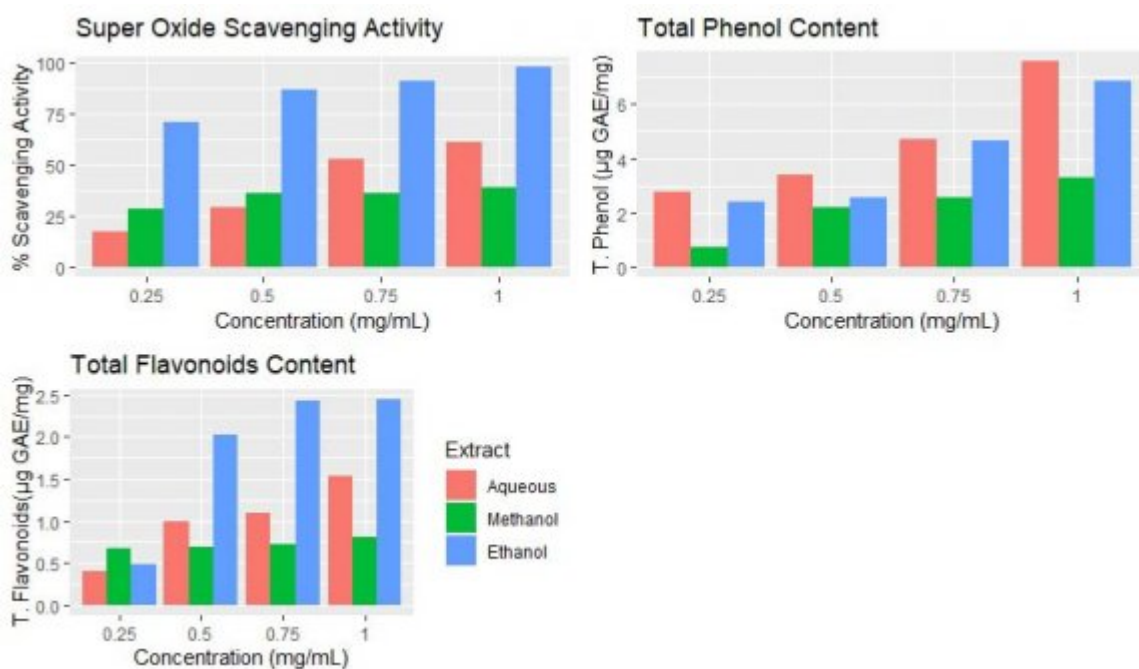
1

Figure 1: Figure 1 :



23

Figure 2: Figure 2 :Figure 3 :



45a5b

Figure 3: Figure 4 :Figure 5a :Figure 5b :

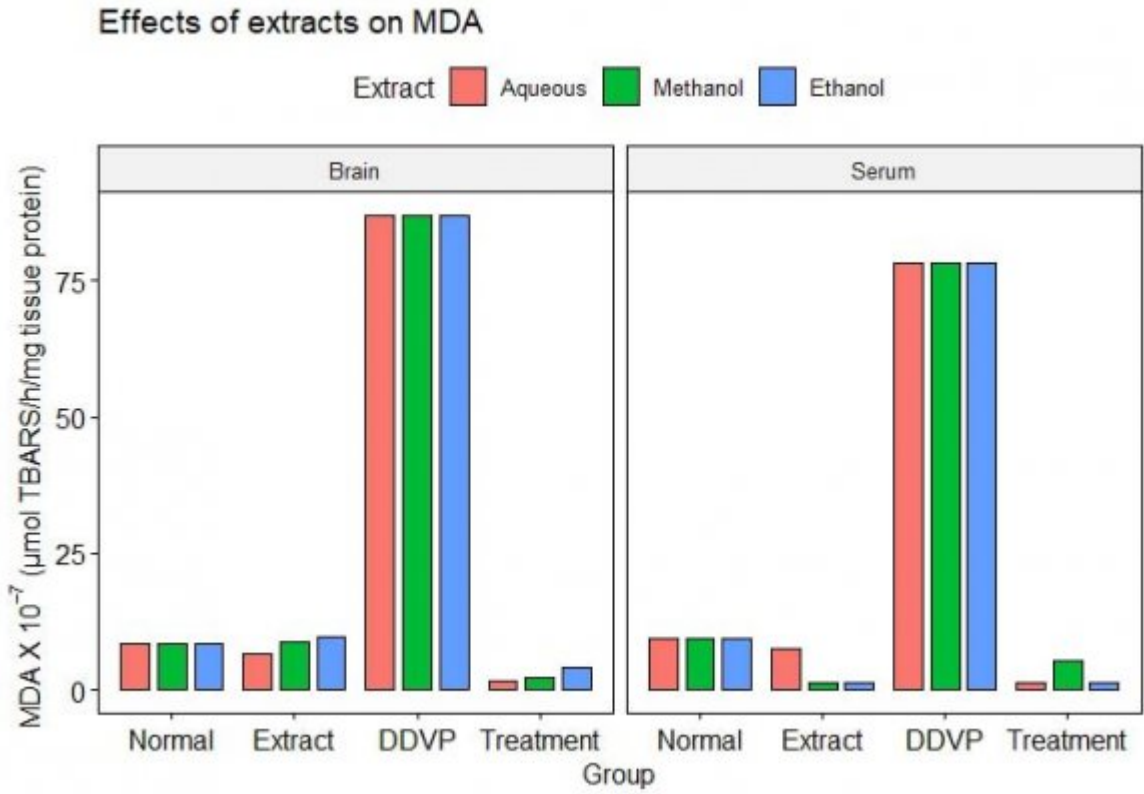


Figure 4:

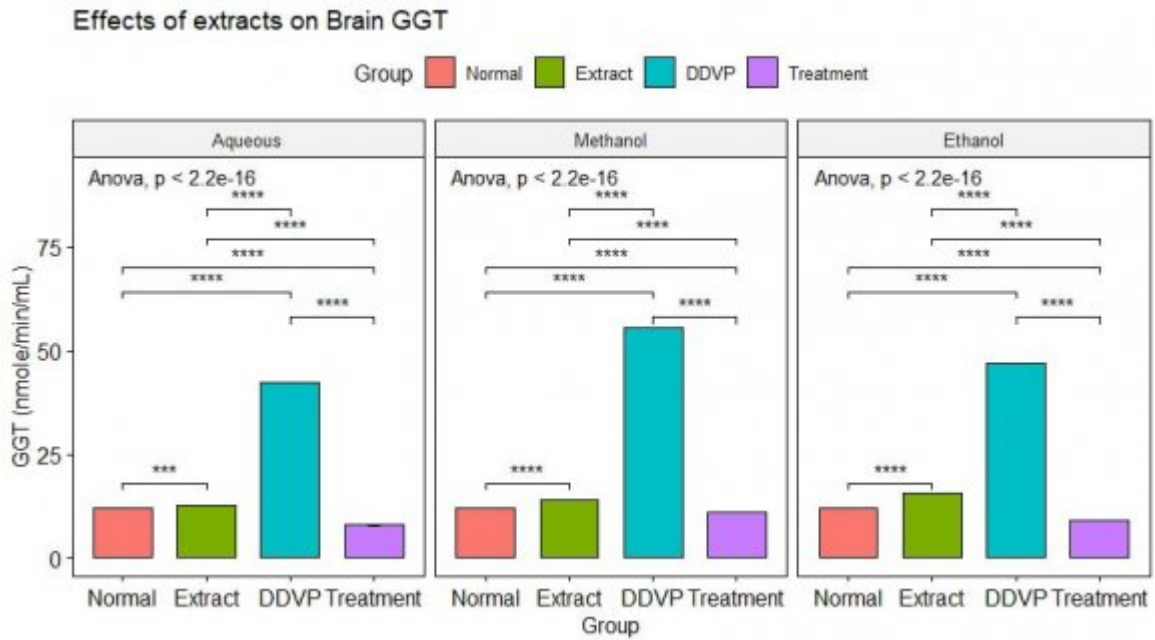


Figure 5:

Figure 6:

The Impact of Senna Siamea (Lam) Leaves Extracts on 2, 2-Dichlorovinyl dimethyl Phosphate Induced Oxidative Stress in Wistar Albino Rats
 therefore be responsible for the medicinal properties of these leaves extracts which was similar to many other Glutathione transferase reported studies by 32 0.08 ethanolic extract of Alstonia boonei stem bark and 33 in
 Year 0 0.01 0.02 0.03 0.04 0.05 0.06 in-vitro compositional investigations of antioxidants, 0.07 ($\mu\text{mol}/\text{min}/\text{m}$
 2021

34

Volume 5 10 15 20 25 30 35 Glutathione reductase Glutathione Extra Serum DDVP Induced
 XXI reductase (U/L) Normal Control Con-
 Is- trol

sue

III

Ver-

sion

I D

D

D

B

D)

(

Global 2 4 6 8 10 12 14 Reduced Glutathione Reduced (mM)

Jour- Glutathione

nal

of

Med-

ical

Re-

search

0

Normal Control

Extra DDVP Induced

Con-

trol

© 2021 Global Journals

Figure 7:

210 Ethics approval and consent to participate: All necessary National and International ethical considerations
211 were fully followed in handling the animals.

212 .1 Consent for publication:

213 The consent for publication was given by all the Authors.

214 Availability of data and material: All data and material regarding the manuscript are available and not under
215 any restriction elsewhere.

216 .2 Competing interests:

217 The Authors declare that no Competing interests exist in any-form Funding: The research was self-funding by the
218 authors Authors' contributions: The Authors OOA and OOP designed the work concept, involved in laboratory
219 Analyses, and write up of the manuscript. Author OOS was involved in the statistical analysis and proof-reading
220 of the manuscript.

221 [Euler and Josephson ()] , Von Euler , H V Josephson , K . *European Journal of Organic Chemistry -Eur J. Org*
222 *Chem* 1972. 452 (1) p. .

223 [Ruch et al. ()] , R J Ruch , S J Cheng , J E Klaunig . *Carcinogenesis, Rely. Chim. Acta* 1989. 10 p. .

224 [Trease and Evans (ed.) ()] , G E Trease , W C Evans . *Pharmacognosy*. 15th Ed. London (ed.) 2002. Saunders
225 Publishers. p. .

226 [Shalu et al. ()] 'A Comparative Study on the Antioxidant Activity of Methanol Extracts of *Acacia nilotica* and
227 *Berberis chitria*'. A Shalu , G T Kulkarni , V N Sharma . *Advances in Natural and Applied Sciences* 2010. 4
228 (1) p. .

229 [Atif et al. ()] 'Acacia nilotica: A plant of multipurpose medicinal uses'. A Atif , A Naveed , A K Barkat , S
230 K Muhammad , R Akhtar , U Z Shahiq-Zaman , K Nayab , W Khalid , M Tariq , A Liaqat . *Journal of*
231 *Medicinal Plants Research* 2012. 6 (9) p. .

232 [Green et al. ()] 'Analysis of nitrate, nitrite, and [15N] nitrate in biological fluids'. L C Green , D A Wagner , J
233 Glogowski , P L Skipper , J S Wishnok , S R Tannenbaum . *Anal Biochem* 1982. 126 (1) p. .

234 [Hossain et al. ()] 'Antidiarrhoea activity, nitric oxide scavenging and total tannin Content from the Bark of
235 *Ceriops decandra* (Griff.) Ding Hou'. M H Hossain , M M Hassan , I A Jahan , I Nimmi , A Islam .
236 *International Journal of Pharmaceutical Sciences and Research* 2012. 3 (5) p. .

237 [Rauha et al. ()] 'Antimicrobial effects of finished plant extract containing flavonoids and other phenolic
238 compounds'. J P Rauha , S Remes , W Herinonen , M Hopia , T Kgjala , K Pitinlaja , H Vaorela , P
239 Vaorela . *Int. Journal of Food Microbiology* 2000. 56 p. .

240 [Amoussa et al. ()] 'Antioxidant activity and total phenolic, flavonoid and flavonol contents of the bark extracts
241 of *Acaciaataxacantha*'. A O Amoussa , A Sanni , L Lagnika . *Journal of Pharmacognosy and Photochemistry*
242 2015. 4 (2) p. .

243 [Asha et al. ()] 'Antioxidant Activity of *Euphorbia hirta* Linn Leaves Extracts'. S Asha , P Thirunavukkarasu ,
244 V M Mani , A M Sadiq . *European Journal of Medicinal Plants* 2016. 14 (1) p. .

245 [Yu et al. ()] 'Antioxidant effect of grapevine leaf extract on the oxidative stress induced by a high-fat diet in
246 rats'. Q Yu , E Lim , S Choi , J Seo . *Food Sci Biotechnol* 2014. 23 p. .

247 [Chance and Maehly ()] 'Assay of catalase and peroxidase'. B Chance , A C Maehly . *Meth. Enzymol* 1955. 2 p.
248 .

249 [Jyoti and Saroj ()] 'Assessment of Antioxidative Potential of *Acacia Nilotica* (L.) Willd Ex Del. Via In Vitro
250 Models'. M Jyoti , A Saroj . *International Journal of Life Sciences Biotechnology and Pharma Research* 2013.
251 2 (4) p. .

252 [Jollow et al. ()] 'Bromoibenzene-induced Liver necrosis: Protective role of glutathione and evidence for 3,4-
253 Bromobenzene oxide as hepatotoxic metabolite'. D J Jollow , J R Michell , Zampaglionic , J R Gillete .
254 *Pharmacology* 1974. 11 p. .

255 [Singleton and Rossi (1965)] 'Colorimetry of Total Phenolics with Phosphomolybdc-Phosphotungstic Acid
256 Reagents'. V L Singleton , J A Rossi . *Am J EnolVitic* January 1965. 16 p. .

257 [Aminu et al. ()] 'Dichlorvos induced AChE inhibition in discrete bbrain regions and the neuro-cognitive
258 implications: Ameliorative effect of *Nigella sativa*'. I Aminu , A Muhammed , I A Wahab , A Abdulmusawir ,
259 A Abdulbasit , I Abdulmuwim , G Sadiya , N P Abdulgafar . *Iranian Journal of Toxicology* 2018. 12 (5) p. .

260 [Ojo et al. ()] 'Dichlorvos-induced oxidative stress in rat brain: Protective effects of the ethanolic extract of
261 *Alstonia boonei* stem bark'. O A Ojo , B E Oyinloye , B O Ajiboye , A B Ojo , H Musa , O I Olarewaju .
262 *Asian Journal of Pharmaceutical* 2014. 8 p. .

13 CONCLUSION

- 263 [Ojo et al. ()] 'Dichlorvos-induced oxidative stress in rat brain: Protective effects of the ethanolic extract of
264 Alstonia boonei stem bark'. O A Ojo , B E Oyinloye , B O Ajiboye , A B Ojo , H Musa , O I Olarewaju .
265 *Asian J Pharm* 2014. 8 p. .
- 266 [Chedi and Aliyu ()] 'Effect and management of Acute Dichloros poisoning in wistar rats'. Baz Chedi , M Aliyu
267 . *Bayero Journal of pure and applied sciences* 2010. 3 (2) p. .
- 268 [Utley et al. ()] 'Effect of sulfhydryl reagents on peroxidation in microsomes'. H G Utley , F Bernheim , P
269 Hochstein . *Arch. Biochem. Biophys* 1967. 118 p. 29.
- 270 [Sani et al. ()] 'Effects of Three Medicinal Plants Extracts in Experimental Diabetes: Antioxidant Enzymes
271 Activities and Plasma Lipids Profiles in Comparison with Metformin'. M F Sani , S M Kouhsari , L Moradabadi
272 . *Iran J Pharm Res* 2012. 11 (3) p. .
- 273 [Uroko et al. ()] 'Evaluation of Antioxidant Activity of Aqueous Extracts of Palm Fruits (*Elaeis guineensis*)'. R I
274 Uroko , A Agbafor , O N Uchenna , N K Achi , S I Egba , P C Nweje-Anyalowu , O R Ngwu . *Asian Journal
275 of Biochemistry* 2017. 12 p. .
- 276 [Onoja et al. ()] 'Evaluation of the In Vitro and In Vivo Antioxidant Potentials of Aframomum melegueta
277 Methanolic Seed Extract'. S O Onoja , Y N Omeh , M I Ezeja , M N Chukwu . 10.1155/2014/159343.
278 <https://doi.org/10.1155/2014/159343> *Journal of Tropical Medicine* 2014. p. .
- 279 [Halliwell Gutteridge (ed.) ()] *Free Radicals in Biology and Medicine*, B Halliwell, J Gutteridge (ed.) (New York)
280 1999. Oxford University Press. p. .
- 281 [Carlberg and Mannervik ()] 'Glutathione reductase'. I Carlberg , B Mannervik . *Meth. Enzymol* 1985. 113 p. .
- 282 [Habig et al. ()] 'Glutathione transferase: A first enzymatic step in mercapturic acid formation'. W H Habig ,
283 M S Pabst , W B Jekpoly . *J. Biol. Chem* 1974. 249 p. .
- 284 [Winston ()] 'Health-promoting properties of common herbs'. J C Winston . *Am J Clin Nutr* 1999. 70 p. .
- 285 [Sandalo et al. ()] 'Immunocytochemical localization of copper, zinc superoxide dismutase in peroxisomes from
286 watermelon'. L M Sandalo , E Lo'pez-Huertas , P Bueno , Del R? 'o , LA . *Citrullus vulgaris* Schrad.)
287 *cotyledons. Free Radic Res* 1997. 26 p. .
- 288 [Oseni and Williams ()] 'In-vitro compositional investigations of antioxidants, phytochemicals, nutritional and
289 minerals in the fruit of *Kigelia africana*'. O A Oseni , O D Williams . *Lam.) Benth. International Journal of
290 Contemporary Research and Review* 2018. 9 (8) p. .
- 291 [Krishnamurthy and Wadhvani ()] *InTech open access chapter distributed under the terms of the Creative
292 Commons Attribution License*, P Krishnamurthy , A Wadhvani . 2012. p. . (Antioxidant Enzymes and
293 Human Health)
- 294 [Teixeira et al. ()] 'Intracellular hydrogen peroxide levels in cells over expressing CuZn-superoxide dismutase'. H
295 D Teixeira , R I Schumacher , R Meneghini , Lower . *Proc Natl Acad Sci* 1998. 95 p. .
- 296 [Fiorino et al. ()] 'Isbarakol anxiolytic?'. D F Fiorino , D Treit , J Menard , L Lerner , A G Phillips . *Behav
297 Pharmacol* 1998. 9 p. .
- 298 [Mohandas et al. ()] 'Low activities of glutathione-related enzymes as factors in the genesis of urinary bladder
299 cancer'. J Mohandas , J J Marshal , G G Duggin , J S Horvath , D G Tiller . *Cancer. Res* 1984. 44 (11) p. .
- 300 [Michael et al. ()] 'Management of acute organophosphorus pesticide poisoning'. E Michael , A B Nick , E Peter
301 , H D Andrew . *Lancet* 2008. 371 (9612) p. .
- 302 [Sofowora ()] 'Medicinal Plants and Traditional Medicinal in Africa'. A Sofowora . *Spectrum Books Ltd; Screening
303 Plants for Bioactive Agents*, (Sunshine House, Ibadan, Nigeria) 1993. p. . (2nd Ed)
- 304 [Surh ()] 'Molecular mechanisms of chemopreventive effects of selected dietary and medicinal phenolic sub-
305 stances'. Y Surh . *Mutat Res* 1999. 428 p. .
- 306 [Oseni et al. ()] 'Nicotine Induced Liver Toxicity in Wistar Albino Rats: Protective effects of Aqueous Extract of
307 MoringaOlifera (Lam)'. O A Oseni , O Akindolire , A Musbau . *Global Journal of Medical Research: Pharma,
308 Drug Discovery* 2018a. 18 (4) p. . (Toxicology & Medicine)
- 309 [Oseni et al. ()] 'Partial Characterization of Chitosan-Iron Complex and its Effects on Alloxan Induced Diabetic
310 Mellitus in Wistar Albino Rats'. O A Oseni , S A Olagboye , O T Adams , Akintayo P Synthesis . *International
311 Journal of Contemporary Research and Review* 2019. 10 (4) p. .
- 312 [Choudhary et al. ()] 'Radical scavenging activity of phenolics and flavonoids in some medicinal plants of India'.
313 R K Choudhary , E S Ajaya , P L Swarnkar . *Journal of Pharmacy Research* 2011. 4 (3) p. .
- 314 [Subhadhirasakul and Khumfang ()] 'Screening of barakol from Cassia plants and some of its biological activities'.
315 S Subhadhirasakul , P Khumfang . *Songklanakarinn J Sci Technol* 2000. 22 p. .
- 316 [Senna siamea". Natural Resources Conservation Service PLANTS Database (2020)] *Senna siamea*". *Natural
317 Resources Conservation Service PLANTS Database*, May 09 2020. (USDA United State Department of
318 Agriculture)

- 319 [Shirwaikar A Shirwaikar et al. ()] A Shirwaikar A Shirwaikar , K Rajendran , Isj Punitha . *Vitro Antioxidant*
320 *Studies on the Benzyl Tetra Isoquinoline Alkaloid Berberine*, 2006. 29 p. .
- 321 [Ganesan et al. ()] ‘Solanum trilobatum L. Ameliorate Thioacetamide-Induced Oxidative Stress and Hepatic
322 Damage in Albino Rats’. K Ganesan , K Sukalingam , B Xu . *Antioxidants* 2017. 6 (68) p. .
- 323 [Fridovich ()] ‘Superoxide radical and superoxide dismutases’. I Fridovich . *Annu Rev Biochem* 1995. 64 p. .
- 324 [Sigma-Aldrich ()] *Technical Bulletin’ Sigma-Aldrich Co*, Sigma-Aldrich . LLC, USA. www.sigmaaldrich.com
325 2013.
- 326 [Zhishen et al. ()] ‘The determination of flavonoid contents in mulberry and their scavenging effects on superoxide
327 radicals’. J Zhishen , T Mengcheng , W Jianming . *IACUC. International Animal Care and Use Committee*
328 *Lab Animal* 1999. 2010. 64 (6) p. 39. (Food Chemistry)
- 329 [Winterbourne et al. ()] ‘The estimation of red cell superoxide dismutase activity’. C C Winterbourne , R E
330 Hawkins , M Brain , R W Carrel . *J. Lab.chem.Med* 1975. 85 p. .
- 331 [Oseni et al. ()] ‘The protective effects of aqueous extract of African nutmeg (*Myristicafragrans*) in bromatein-
332 duced spleen and cardiac tissue toxicities using male wistar albino rats’. O A Oseni , S A Olagboye , O T
333 Adams , B S Maikasawa . *Journal of Drug Delivery and Therapeutics* 2018b. 8 (5) p. .
- 334 [Misra and Fridovich ()] ‘The role of superoxide anion in the autoxidation of epinephrine and a simple assay for
335 superoxide dismutase’. H P Misra , I Fridovich . *J Biol Chem* 1972. 247 p. .
- 336 [Brand-Williams et al. ()] *Use of a free radical method to evaluate antioxidant activity. Lebensmittel-Wissenschaft*
337 *und-Technologie*, W Brand-Williams , M E Cuvelier , C Berset . 1995. 28 p. .