

Characterization and Antibiotic Sensitivity Testing of Clinical Bacteria Species Isolated from Kunu Drink Sold in Rumuolumeni, Rivers State

Dr. Austin A. Okwelle¹

¹ Ignatius Ajuru University of Education

Received: 7 June 2021 Accepted: 30 June 2021 Published: 15 July 2021

Abstract

The isolation, characterization and antibiotic sensitivity tests of some clinical bacteria species isolated from Kunu drink sold in Rumuolumeni, Rivers State was carried out. Samples of the Kunu drink was bought from vendors indifferent locations and their bacteriological counts enumerated using standard microbiological methods by the pour plate technique. The antibiotic sensitivity pattern of the pure bacteria isolates against some antibiotics was determined using the disc diffusion method. The total bacterial counts of the Kunu in the different locations ranged from 4.0×10^2 to 8.6×10^2 cfu/ml. Four species of bacteria including *Escherichia coli*, *Enterobacter aerogenes*, *Staphylococcus aureus* and *Streptococcus* spp were isolated and identified by grain staining and their biochemical reactions. The most prevalent isolate in terms of occurrence was *Escherichia coli* (50)

Index terms—

Samples of the Kunu drink was bought from vendors indifferent locations and their bacteriological counts enumerated using standard microbiological methods by the pour plate technique. The antibiotic sensitivity pattern of the pure bacteria isolates against some antibiotics was determined using the disc diffusion method. The total bacterial counts of the Kunu in the different locations ranged from 4.0×10^2 to 8.6×10^2 cfu/ml. Four species of bacteria including *Escherichia coli*, *Enterobacter aerogenes*, *Staphylococcus aureus* and *Streptococcus* spp were isolated and identified by grain staining and their biochemical reactions. The most prevalent isolate in terms of occurrence was *Escherichia coli* (50%) followed by *Enterobacter aerogenes* (30%), *Staphylococcus aureus* (10%) and *Streptococcus* spp (10%). The antibiotic tests showed that *Escherichia coli* had high resistance to Chloramphenicol (70%), followed by Septrin (62.7%) and Sparfloxacin (62.7%), while *Enterobacter aerogenes*, *Streptococcus* spp and *Staphylococcus aureus* had low rates of resistance to all the antibiotics tested. The results of this study demonstrated that Kunu drink sold in Rumuolumeni was contaminated with potentially pathogenic bacteria species, including antibiotic Introduction according to Maji et al., (2011), Kunu drink is a locally prepared indigenous non-alcoholic beverage normally prepared and consumed in large quantity in Nigeria, especially in the northern part of the country (Amusa and Aswaye, 2009). It can be consumed during the wet and dry seasons due to its thirst quenching properties. Umaru et al., (2014) reported that Kunu drink is sold in many public places such as markets, offices, schools, motor parks and as drinks during festival, weddings and naming ceremonies. It is an appetizer and food complement used to quench hunger (Adelekan et al., 2014). Kunu drinks are usually produced using maize, guinea corn, millet or sorghum in varying proportions (Maji et al, 2011) to which sweet potato sometimes is added to increase the taste of the Kunu, which is a major factor that attracts consumers to the product. This common drink is usually packaged and sold in 50ml to 1L plastic bottle and at times tied in some disposal polythene bags the drink is mostly consumed within 20-35 hours of production due to its poor keeping quality (Akoma et al, 2012). This drink is not expensive because the grains and other ingredients used for production are locally sourced. The packaging materials are also readily available, cheap and affordable within the communities.

8 G) ANTIBIOTIC SUSCEPTIBILITY TEST

45 Different workers have reported that Kunuis rich in vitamins, minerals, carbohydrates and proteins ??Adebayo
46 et

47 1 Materials and Methods

48 2 a) Study Area

49 The study was conducted in Rumuolumeni, Port Harcourt, Rivers state, Nigeria. Port Harcourt lies between
50 latitude 4°46'38.71"N and longitude 7°00'48.24"E. and located in the tropical rainforest in Nigeria.

51 3 b) Collection of Sample

52 Five bottles of hawked kunu samples were bought from vendors in different locations in Rumuolumeni, properly
53 labelled, placed in a sterile plastic container and transported to the Biology laboratory, Ignatius Ajuru University
54 of Education for microbiological analysis.

55 4 c) Processing of Sample

56 The samples were mixed gently and 10ml of each was added to 90 ml distilled water with a clean pipette.
57 The solution was mixed and diluted serially by transferring 1 ml of the stock sample into sterilized test tubes
58 containing 9ml of peptone water. The procedure was repeated for the third and fourth test tubes to make a
59 dilution of 10^3 and 10^7 .

60 5 d) Preparation of Media

61 All the glassware used such as petri-dishes, conical flasks, test tubes and pipettes were washed with detergent,
62 rinsed in water, dried and sterilized in the hot air oven at 60°C for 1 hour. Different culture media such as Nutrient
63 Agar, MacConkey Agar, Salmonella-Shigella Agar (SSA), Trisulphate Citrate Bile Salt Agar (TCBS) and Manitol
64 Salt Agar (MSA) were used for isolation. Each of the media was prepared by weighing out appropriate quantities
65 according to the manufacturers instruction and dissolved completely in the required volume of distilled water.
66 The media were autoclaved at 121°C for 15 minutes and allowed to cool at 45-50°C. The media was dispensed
67 aseptically into the petri-dish plates and left on the table to solidify at room temperature.

68 6 e) Isolation and Preservation

69 Using a sterile loop, discrete colonies were all sub-cultured onto another media to obtain pure colonies. This
70 was done by streaking a loopful of a particular isolate into freshly prepared culture media plates for bacteria.
71 The sub-cultured nutrient agar plates were incubated at 37°C. Bacteria pure cultures were accordingly stored in
72 sterile agar slants for preservation and further analysis at 4°C.

73 7 f) Identification of Isolates

74 The isolates were identified using gram staining and biochemical tests such as: motility test, urease test, citrate
75 utilization test, indole test, oxidase test, coagulase test, catalase test, vogues proskauer reaction and methyl red
76 test. Identification was based on comparison of the characteristics of the isolates described by (Chess brough,
77 2006).

78 8 g) Antibiotic Susceptibility Test

79 The isolates were screened for antimicrobial sensitivity using the Kirby-Bauer agar disk diffusion method (CLSI,
80 2009). A suspension of each isolate was prepared in peptone water to match 0.5 McFarland turbidity standards in
81 order to standardize the inoculum. The standardized inoculum of each isolate was inoculated onto the surface of
82 plain Mueller-Hinton agar plates and Septrin (30 µg), Chloramphenicol (30 µg), Sparfloxacin (5 µg), Amoxycillin
83 (30 µg), Ciprofloxacin (5 µg) Augmentin (30 µg), Gentamicin (10 µg), Pefloxacin (10 µg), Tarivid (30 µg)
84 and Streptomycin (10 µg) discs were placed and incubated at 37°C for 24 hours. The zones of inhibition
85 were measured and compared with the Clinical and Laboratory Standards Institute. 3 shows the Antibiotic
86 resistance pattern of the bacterial species isolated from Kunun drinks sold in Rumuolumeni, Port Harcourt
87 metropolis. E. coli exhibited very high percentage resistance to chloramphenicol (75.0%) followed by Septrin
88 (68.7%) and Sparfloxacin (68.7%) respectively, whereas there was no resistance to Perfloxacin (0.0%). The
89 highest percentage resistance for Enterobacter aerogenes was recorded with Augmentin (50.0%) and the least
90 resistance was recorded with Ciprofloxacin (7.1%). The percentage resistance Streptococcus spp. isolated was
91 relatively low which ranged from 38.3% to 33.3% for the antibiotics to which this species showed resistance (Trivid,
92 Sparfloxacin, Ciprofloxacin, Augmentin and Streptomycin). However, the isolates of the Streptococcus spp.
93 showed completely no resistance (0.0%) to Septrin, Chloramphenicol, Amoxicillin, Gentamicin and Perfloxacin.
94 Similarly, the percentage resistance of Staphylococcus aureus isolated was relatively low which ranged from
95 33.4% to 16.7% for the antibiotics to which these isolates showed resistance (Gentamicin, Perfloxacin, Septrin,
96 Sparfloxacin, Ciprofloxacin and Streptomycin).

9 III.

10 Results

IV.

11 Discussion

The relative high numbers of microbial counts obtained from the different samples of kunu in the study were indicative of high level of microbial contamination of the prpduct. The Kunu sold at Town Hall had the highest counts of 8.6×10^2 cfu/ml, while the one from St Johns location had the lowest counts of 4.0×10^2 cfu/ml. The high microbial counts experienced may be attributed to lack of effective precautions on hygiene practice in handling procedures during processing of the beverage. The practice of addition of some quantity of water to Kunu after fermentation may also be a source of microbial contaminants, which may have come from the water itself or from the utensils used for such purposes. In an earlier report, Amusa and Ashwaye (2009) had stated that the presence of coliforms such as *Esherichia coli* in hawked Kunu was as a result of contaminated water, containers, as well as dirty environment where the Kunu were being processed and hawked. The identification of *Escherichia coli*, *Staphylococcus aureus*, *Streptococcus spp* and *Enterobacter aerogenes* in the samples analyzed is a positive sign to the fact that the Kunu drink sold in the different locations in the community was contaminated with potentially pathogenic bacteria and this may have come from the water used for domestic purposes, or the human handlers during processing and sales of the product, respectively. This is in agreement with Amusa and Ashwaye (2009) and Akoma et al., (2013), who had noted that water used for production coupled with the crude method of production and packaging under improper sanitary conditions predisposes Kunudrink to microbial contamination of both gram negative and gram positive bacteria. The source of contamination may also have come from the spices used additives (Essien et al., 2009, Lawal, 2012). There is therefore need for surveillance by Public Health officials to ensure safety of the Kunu sold for to public. There is need to also ensure that the water used for production especially post-heating processing of the Kunu is safe and free from microbial contaminants.

Antibiogram of the isolates revealed varying levels of resistance to the antibiotics tested. *Escherichia coli* showed high resistance to chloramphenicol (75% is a reflection of the use and misuse of the antibiotics in the society. This is not surprising because outside the hospital environment, the general populace have access to various kinds of antibiotics at any drug store even without any prescription from a medical practitioner. The Public Health implication of this study is that antimicrobial resistant strains of pathogenic bacteria may colonize the human population through consumption of contaminated Kunun and this would lead to chemotherapeutic failures among the human consumers of this popular beverage in the Rumuolumeni, Port Harcourt metropolis.

V.

12 Conclusion

The presence of resistant strains of *E. coli*, *Enterobacter aerogenes*, *Staphylococcus aureus* and *Streptococcus spp* in Kunun sold in Rumuolumeni suggests that consumption of this beverage has potential health hazard to the consumers in Rumuolumeni, Port Harcourt, Nigeria. The consumers of this popular drink are therefore at health risk, which may culminate into failures of commonly used clinical antibiotics for the treatment of the infections.

¹

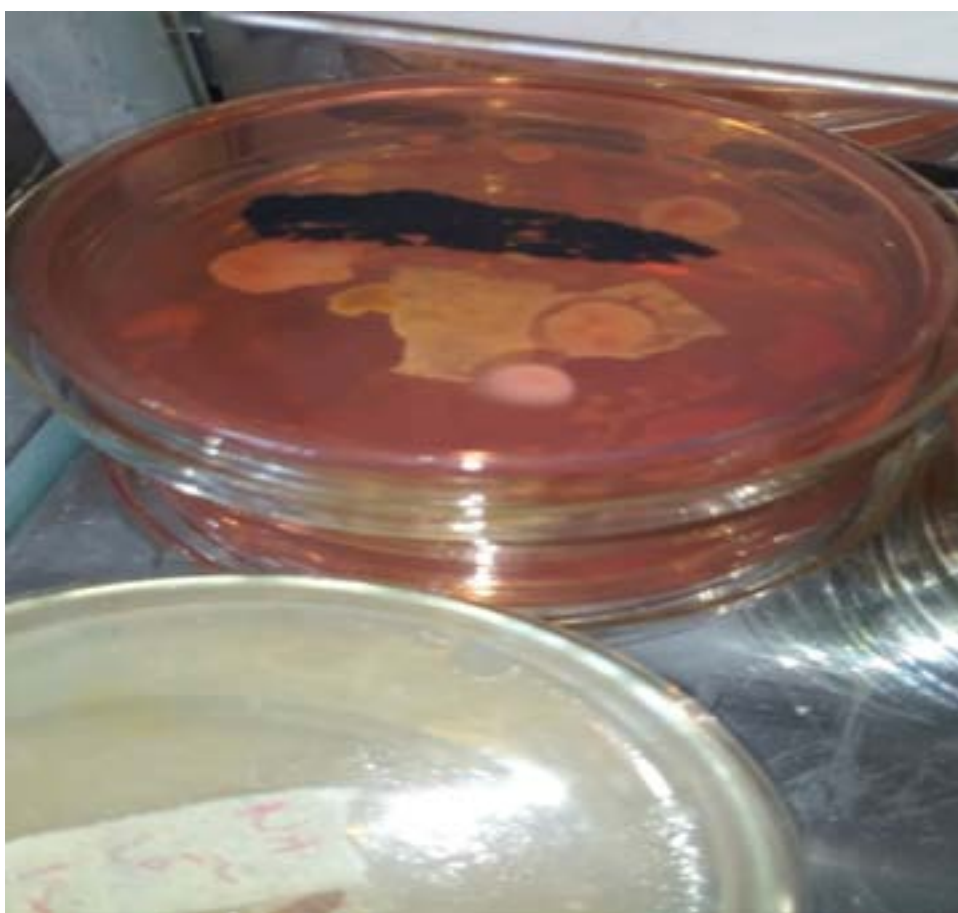


Figure 1:



1

Figure 2: Plate 1 :

Figure 3:

12 CONCLUSION

1

drink results from the processing activities and materials used such as water, handling and presentation techniques. The water content coupled with the crude method of production and packaging under poor sanitary conditions predisposes the drink to sudden microbial contamination (Akoma et al, 2012). Again, there is no public health regulatory agency that monitors the production processes in spite of the associated harm that kunu drink causes Bukar et al., (2010). The aim of this study is to characterize and determine the antibiotic sensitivity of some pathogenic bacteria species isolated from Kunu drink sold in Rumuolumeni, Rivers State.

Location	Kunu
A	5.1×10^2
B	6.2×10^2
C	4.6×10^2
D	7.1×10^2
E	4.1×10^2
F	

[Note: Keys: A = Waterside, B = Big tree market, C = Akar Junction, D = Iwofe school gate, E = St. John's, F = Town Hall]

Figure 4: Table 1 :

2

Locations					
Bacteria Isolates					
Escherichia coli	+	+	+	+	+
Staphylococcus aureus	+	-	-	+	-
Streptococcus spp	-	-	-	-	+
Enterobacteria aerogenes	+	+	+	-	+

Figure 5: Table 2 :

3

Figure 6: Table 3 :

the antibiotics tested. However, *E. coli* had very high sensitivity to Pefloxacin (100%), followed by Gentamicin (88%), Augmentin (75%), tarivid (68.7%) and Percentage Resistance of Isolates Streptomycin (68.7%). Streptococcus Antibiotics Conc. (μg) susceptible isolates which had very high sensitivity Escherichia coli Enterobacter Aer

Septrin30 Chloramphenicol, Chloramphenicol 30 Amoxicillin, Perfloxacin, respectively. Staphylococcus aureus

Sparfloxacin 5 Amoxicillin,	68.7	35.7
Ciprofloxacin 5	50.0	7.1
Amoxicillin 30	12.0	35.7
Augmentin30	25.0	50.0
Gentamicin10	12.0	14.3
Pefloxacin10	0.0	42.9
Tarvid 30	31.3	28.8
Streptomycin 10	31.3	14.3

Figure 7:

-
- 135 [Adebayo] , G B Adebayo .
136 [Eur. J. Food Res. Rev] , *Eur. J. Food Res. Rev* 1 p. .
- 137 [Omeke et al. ()] ‘Antibacterial activity of leaf extract of *Chromolaena odorata* and the effect of its combination
138 with some conventional antibiotics on *Pseudomonas aeruginosa* isolated from wounds’. P O Omeke , J O Obi
139 , N A Orabueze , A Ike . *Journal of Applied Biology Biotechnology* 2019. 7 (03) p. .
- 140 [Mcgeer et al. ()] *Antimicrobial resistance in Ontario: Are we making progress? Laboratory Proficiency Testing*
141 *Program Newsletter*, A Mcgeer , C A Fleming , K Gree , D E Low . 2010. 293 p. .
- 142 [Chesebrough ()] *District laboratory practice in tropical countries: Part 2*, M Chesebrough . 2006. New York:
143 Cambridge University Press.
- 144 [Adedokun and Onyeneke ()] ‘Effect of *Aframomum danellians* black pepper crude extracts on physio-chemical
145 and sensory properties of Kunun-zaki during storage’. I I Adedokun , E Onyeneke . *J. Food Techn* 2012. 10
146 p. .
- 147 [Maji et al. ()] *Effect of chemical treatment and pasteurization on the shelf life of Kunun zaki (sorghum and*
148 *maize gruel)*, A A Maji , J & O E Omale , Chigozie . 2011.
- 149 [Folasade and Oyenike ()] ‘Effect of sesame seed addition on the chemical and sensory qualities of sorghum based
150 Kunun-zaki drink’. M & O Folasade , Oyenike . *Afr. J. Food Sci. Techn* 2012. 3 p. .
- 151 [Essien et al. ()] ‘Evaluation of the nutritional and microbiological quality of Kunu (A Cereal Based Non-alcoholic
152 Beverage) in Rivers State’. E Essien , C Monago , E Edor . *Nigeria. The Internet Journal of Nutrition and*
153 *Wellness* 2009. 10 p. .
- 154 [Lawal ()] ‘Microbial quality of Kunun-zaki beverage sold in Ile-Ife, Osun State’. O A Lawal . *J. Food Techn* 2012.
155 10 p. .
- 156 [Amusa and R Aswaye ()] *Microbiological and Nutritional Quality of Hawked Kunun (A Sorghum Based Non-*
157 *Alcoholic Beverage) Widely Consumed in Nigeria*, O Amusa , B R & Aswaye . 2009.
- 158 [Oluwajoba and Adedeji ()] ‘Nutritional composition of a non-alcoholic beverage spiced with *Zingiber officinale*
159 extract produced from *Sorghum bicolor* stem sheath’. I B T O Oluwajoba , Adedeji . *Int. J. Food Sci. Nutr.*
160 *Eng* 2013. 3 p. .
- 161 [Bukar et al. ()] ‘Occurrence of some enteropathogenic bacteria in some minimally and fully processed ready-
162 to-eat foods in Kano metropolis’. A Bukar , A & T I Uba , Oyeyi . *Nigeria. Afr. J. Food Sci* 2010. 4 p.
163 .
- 164 [Csli ()] ‘Performance standards for antimicrobial susceptibility testing’. Csli . *Seventeenth Informational*
165 *Supplements* 2007. p. . Cinical and Laboratory Standard Institute
- 166 [Otunola and Ajao ()] ‘Physiochemical, microbiological and sensory characteristics of Kunu prepared from millet,
167 maize and guinea corn and stored at selected temperatures’. G Otunola , T A Ajao . *Adv. J. Food Sci. Techn*
168 2010. 2 p. .
- 169 [Falagas et al. ()] ‘Resistance to polymyxins: Mechanisms, frequency and treatment options’. M E Falagas , P I
170 Rafailidis , D K Matthaiou . *Drug Resist. Update* 2010. 13 p. .