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Relationship between Oxidative Stress and Diabetes Mellitus

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I. INTRODUCTION

DM is a metabolic disorder characterized by high levels of free glucose in the blood. Diabetes is a disorder in the human body that causes blood glucose (sugar) levels to rise above normal due to insulin secretion or insulin resistance. This is also called hyperglycaemia (1). DM is an incurable disease that has a detrimental effect on many metabolic pathways and contributes to the pathophysiology of diabetic complications (2, 3). Oxidative stress is a very important factor in the development of diabetes (8). Oxidative stress is known to be associated with lifestyle-related diseases including atherosclerosis, high blood pressure, and diabetes. Free radical forms are important for body parts in biological homeostasis (11, 12), but where their production is excessive and greater than the body's antioxidant capacity, then the effects of oxidative stress (12). Oxidative stress is a major factor in the development of diabetes and insulin resistance (12-14), triggering pathophysiologic pathways and initiating a burst of malignant pathways leading to insulin resistance and DM (8, 15). In this review, we discussed the potential roles of oxidative stress in building insulin resistance and DM.

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II. CLASSIFICATION OF DIABETES MELLITUS

There are different types of DM, but the most common subtypes are 1 DM (T1DM) and 2 DM type (T2DM). T1DM occurs due to beta-cell dysfunction, reduced insulin secretion, and low levels of circulating insulin. T2DM is the most common type of DM that accounts for approximately 90–95% of patients with diabetes and is mainly associated with insufficient response to insulin (reduced insulin sensitivity) and insulin resistance in borderline tissues (16). With type 2 diabetes, the human body does not use insulin properly. This is called insulin resistance. Initially, pancreas produces extra insulin to make it. Over time the pancreas may not be able to cope and it may not produce enough insulin to maintain normal blood sugar levels. Type 2 can be controlled by improving lifestyle, oral hypoglycaemic therapy, and insulin. Pregnancy diabetes is another topic that occurs in women during pregnancy when the body is less sensitive to insulin. Pregnancy diabetes does not occur in all women and usually develops after childbirth (17). Other types of DM are adolescent diabetes which is diabetes mellitus, pre-existing diabetes in adults, and secondary diabetes from other diseases such as pancreatitis or secondary to the use of drugs such as corticosteroids (18, 19).

III. OXIDATIVE STRESS AND ANTIOXIDANT PROCESS

Various normal cells produce free radicals such as aerobic respiratory products and other metabolic processes (7) including reactive oxygen species (ROS). ROS is a highly active oxidant and can have adverse effects on cellular lipids, proteins, and DNA or reactive oxygen species (ROS) produced by organisms due to normal cell metabolism and environmental factors, such as air pollution or cigarette smoke. ROS are highly active molecules and can damage cell formation such as carbohydrates, nucleic acids, lipids, and proteins and alter their functions. Cells usually contain enzymes and coenzymes that act as antioxidants. This helps to reduce ROS and prevents it from causing damage (6). Oxidative stress is defined as an imbalance between the chemical processes responsible for the production of active oxygen (ROS) and those responsible for the removal of ROS (20). There are many enzymes in the cell that have internal mechanisms such as superoxide dismutase (SOD), catalase (CAT), and glutathione

(GLT), which protect cells from free radicals (25). Some heavy metal products have free properties such as iron (ferric) and copper (26) that can mix proteins, lipids, and nucleic acids and produce toxic products that lead to tissue dysfunction (27,28). They alter the structure of biologic molecules and break them down (28). DNA fragmentation is a well-known result of oxidative stress, which affects genetic expression and cell survival (23). Malondialdehyde (MDA), total cholesterol, and active hydroperoxides (ROOH) are oxidative stress biomarkers that occur in diabetic patients. (30). Oxidative stress plays important roles in diabetic complications through lipid peroxidation, DNA damage, and mitochondrial dysfunction (13, 23, 31, 32). Its involvement has been shown in other illnesses and age-related problems such as cardiovascular disease, chronic obstructive pulmonary disease, chronic kidney disease, neurological diseases, and cancer and more. Including high free varieties (33). Many scientists believe that the theory of oxidative evolution is a major cause of aging and related problems (33). Problems caused by oxidative stress and insulin resistance are prevented with the help of redox biology (34).

IV. NORMAL INSULIN SIGNALING PATHWAYS AND INSULIN RESISTANCE

Insulin is usually a 51 amino acid dipeptide containing a series A and B series linked to 2 disulfide bonds found in cysteine residues. The chain contains 21 amino acids and the B chain contains 30 amino acids. Insulin is encoded by a short arm of chromosome 11 in pancreatic β -cells containing signal peptide, chain B, connective peptide (and A) and A chain (4,5). In proinsulin, C-peptide binding is enclosed at each end by the dibasic residues (Arg-Arg and Lys-Arg) that link the N-terminus of the A series and the C-terminus of the B chain (124,125). Proinsulin made from Golgi substances is converted to insulin by removing dibasic residues by trypsin-like endoprotease enzymes such as insulin and C-peptide (5). Insulin resistance is a key factor in T2DM where cells are unable to respond to insulin effectively (8, 35) There are different enzymes and mediators, which facilitate the entry of glucose into adipocytes, muscles, and myocardial cells via GLUT- 4 (glucose transporter- 4) transporters [8, 34]. The feature is triggered by binding insulin to α chain of insulin receptors (IRs), which are members of transmembrane tyrosine kinases that are made up of α and-chains and activated by insulin and IGF- (insulin-like growth -) 1 and IGF-2 (36). As a result, binding induces structural changes in the chain by autophosphorylation in tyrosine residues through different adapter proteins, namely, insulin receptor substrates (IRSs), Shc proteins (SHC-transforming), and APS protein (protein adapter) (37,38)). These processes provide a suitable site for binding IRS-1 (insulin receptor substrate-1) (38). Many

types of insulin-dependent kinases such as extracellular protein kinase C, S6K1 (ribosomal protein S6 kinase beta-1), serine / - threonine-protein kinase 2, protein kinase B etc and other types of kinases such as AMP-activated protein kinase and glycogen synthase kinase 3 can activate and activate phosphorylate IRS (38, 39). The activated IRS-1 binds to PI3K (phosphoinositide 3-kinase) and activates it, which, in turn, promotes the conversion of PIP2 (phosphatidylinositol 4,5-bisphosphate) to PIP3 (phosphatidylinositol 3,4,5-trisphosphate) (40). PIP3 itself is a potent activator of Akt, which also contributes to cell-induced glucose uptake through the production of GLUT-4 and inhibits glycogen synthase kinase leading to increased glycogen secretion (40, 41). Any disruption in the above-mentioned steps may cause insulin resistance and DM (34).

V. RELATIONSHIP BETWEEN OXIDATIVE STRESS AND INSULIN RESISTANCE

Oxidative stress increases the risk of insulin resistance and DM (26, 34). It should be noted that oxidative stress caused by DM has more complex interactions (42, 43). The following are potential molecular mechanisms by which free radicals disrupt the normal glucose Homeostasis contributes to the formation of DM.

a) β -Cell Dysfunction/Insulin Production and Secretion

The function of insulin is to maintain blood glucose levels by promoting glucose uptake into insulin-targeted tissues. Glucose is a key regulator of insulin secretion by pancreatic Beta cells, which triggers a burst of events called beta-stimulating cells - the ability to sense circulating blood sugar levels and release the right amount of insulin to keep blood glucose at a normal level (126- 129).

Normal glucose homeostasis is made up of healthy and active beta cells and DM is associated with various levels of beta-cell dysfunction (44, 45). DM occurs as a result of loss of beta-cell function and function (46). In these cases, insulin secretion produced by glucose from beta cells is reduced and decreased; therefore, glucose levels have risen above normal levels (47). In this process, insulin secretion occurs, which is interrupted by a decrease in the energy of the sugar to promote insulin secretion leading to severe unstable insulin release and followed by beta-cell failure (46). Beta-cell dysfunction occurs due to pathogenic mechanisms and oxidative stress (46, 48). Free radicals in pancreatic beta cells arise due to enzyme activity such as mitochondrial respiratory tract (MRC) and NADPH (nicotinamide adenine dinucleotide phosphate) oxidase or NOX enzyme (15, 49-51). Superoxide anion is a major free radical pathway produced by MRC and NOX enzyme in beta cells (52). Beta cells are affected by free radicals produced by phagocytic and immune

cells (53). Chronic hyperglycemia induces free radical production in the islands by increasing cytosolic calcium and protein kinase activation pathways (50, 54). Beta cells have a low dose of antioxidant protection system, so that oxidative stress on beta cells increases in DM and plays an important role in the loss of their function in both T1DM and T2DM (45, 48, 55). Oxidative stress disrupts beta-cell function through a number of molecular mechanisms (48, 55-62). Significantly reduces insulin production, disrupts the insertion of proinsulin vesicles into plasma membranes, and

reduces their exocytosis in response to glucose distribution (48, 55, 56). It can also cause apoptotic processes in pancreatic cells that lead to death and loss of beta cells (48, 55, 56). Therefore, depressive beta cell dysfunction caused by stress is a major potential target for experimental intervention in patients with DM. We suggest that pharmacologic agents protecting the islands from oxidative damage may provide Target therapeutics to promote beta-cell function that leads to improved glucose homeostasis.

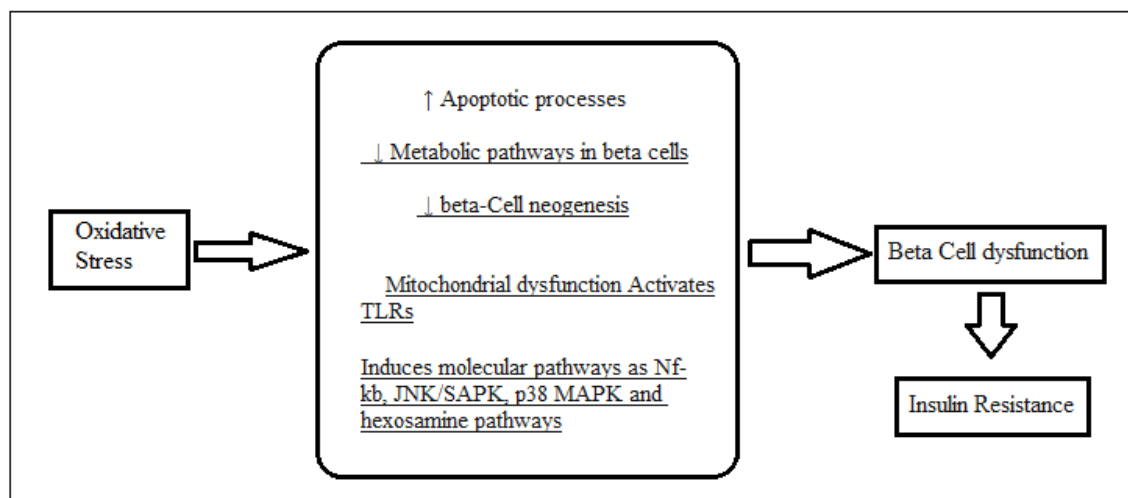


Figure 1: Possible molecular mechanisms between oxidative stress and beta-cell dysfunction

b) GLUT-4 Expression and/or Localization

GLUT4 is an insulin-regulated glucose Transporter found in adipose tissue and bound muscles such as skeletal and heart muscle and, therefore, to maintain insulin sensitivity in these tissues a normal body profile of GLUT-4 expression and / or localization is required (15,63). The reducing factor of the GLUT-4 antagonist has an effect on insulin sensitivity (63, 64) as a decrease in glucose entering target cells translates into lower insulin sensitivity in these tissues (65). According to Clinical Studies GLUT-4 expression and / or localization decreased insulin-resistant and T2DM-resistant patients (64, 66, 67). This patho physiologic condition is exacerbated by oxidative stress (68, 69). Oxidative stress can reduce the content of GLUT-4 by impairing gene expression by damaging the binding of the nuclear material to the GLUT-4-induced insulin receptor in 3T3-L1 adipocytes (70). 3T3-L1 adipocytes develop oxidative stress in these cells and receive Glut-4 expression from these tissues. They found that hydrogen peroxide produced by significant oxidative stress is regulated by GLUT-4 to 3T3-L1 adipocytes and, consequently, reduces cellular glucose (70). Other studies suggest that oxidative stress reduced GLUT-4 transport to cell membranes. They induced mitochondrial oxidative stress using mitochondria-targeted paraquat to adipocytes and found that

oxidative stress significantly reduced GLUT-4 transport and thus induced insulin resistance in these tissues (71). Long-term oxidative stress can suppress transcription factors involved in GLUT-4 expression such as peroxisome proliferator-activated receptor gamma, CCAAT-binding proteins, factor 1, MEF2 (myocyte enhancer factor 2), and Nf- κ b etc. (70, 72-74). It can also suppress small amounts of RNA involved in GLUT-4 expression such as miR-21a-5p, miR-222- 3p, miR-29c-3p, and miR-133a-3p etc (75-78). In addition, a variety of stress-inducing substances and products such as p38 MAPK, JNK / SAPK, PKC (protein kinase C), sorbitol, and hexosamine are all produced by oxidative damage and may suppress GLUT-4 (29). Therefore, reduced expression / activation of GLUT-4 is one of the major molecular mechanisms by which oxidative stress induces insulin resistance and contributes to the formation of DM (15).

c) Insulin Signaling Pathways

Insulin resistance with DM occurred due to any impairment in insulin signaling pathways (79, 80). In appreciation of insulin sensitivity was introduced novel therapeutic variables of the insulin signal (80). Oxidative stress can disrupt normal IST (insulin signal transduction) at various levels including IR, IRS-1 and IRS-2; PI3K enzyme and Akt signature methods (81-86). T2DM-induced oxidative stress and IST substances in

the brains of diabetic mice caused by Balbaa and colleagues in 2017 (87). They found that oxidative stress significantly reduced the expression of an IST substance such as p-IRS, p-AKT, and GSK-3 β in brain tissue with normal insulin signaling (87). Oxidative stress induced IRS-1 and IRS-2 serine phosphorylation, leading to disrupted IST (81, 82). Free radicals can induce serine phosphorylation of IRS-1 and suppress normal IST using JNK / SAPK signaling methods (85). Other types of serine / threonine kinases such as Akt (or PKB), GSK-3,

AMPK, and mTOR are also very sensitive to oxidative stress and may interfere with insulin signaling (100-102). Oxidative stress can also lead to IST impairment by down-regulating proteins involved in normal IST (87). IST substances such as Akt, IRS, IRS-1, and GSK-3 are under the influence of low free radicals regulated by oxidative stress thus interfering with insulin sensitivity leading to insulin resistance and DM (87). Therefore, IST disruption is another important link between oxidative stress and insulin resistance (81-86, 88-90).

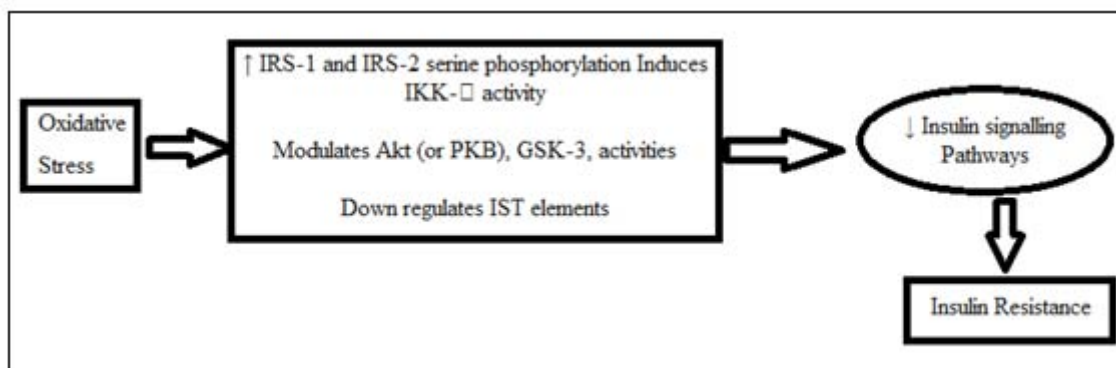


Figure 2: Oxidative stress impairs insulin signaling pathways by several molecular pathways.

d) Inflammatory Processes

The inflammatory response is one of the main molecular mechanisms involved in the pathophysiology of insulin resistance, DM, and related complications (53, 91, 92). A separate study suggests that chronic inflammation is involved in the pathophysiology of insulin resistance DM (92-98). This type of mechanism can also establish other pathophysiologic mechanisms of DM such as beta-cell dysfunction (53, 95). Animal experimental and pathological studies show that IR and inflammation are directly linked during T2DM development (24,42). Inflammatory mediators play an important role in improving IR and T2DM by activating various inflammatory responses such as IL- β . IL- α is an effective pro-inflammatory mediator that plays an important role in controlling various inflammatory mediators such as cytokines, adipokines and chemicals. It causes inflammation by binding to interleukin-1 receptor type I (IL-1RI) and reduces the expression of insulin receptor substrate-1 (IRS-1) at the ERK-dependent writing level and the post-transcriptional ERK level (43)). IL-1 β production is largely controlled by dietary stress caused by diet. Other experimental studies have been performed on a variety of experimental animals to investigate the presence of various inflammatory responses in β -cells that indicate that IL- β plays a key role in activating other inflammatory mediators such as cytokines and chemicals (21, 22). In cell-cells of pancreatic islands due to impaired insulin secretion occurs in β -cells of pancreatic islands. In this way IL- β also plays an important role in causing inflammation of the tissues of the body because it

reduces the ability of the insulin receptors to respond to glucose which ultimately leads to the formation of IR in borderline tissues. IL-6 is another mediator that can be positively linked to IR [19 - 22]. IL-6 not only inhibits the metabolism of non-oxidative glucose (120,121) but also suppresses the activity of lipoprotein lipase which increases plasma levels of triglycerides [23]. In addition, IL-6 also activates cytokine signaling proteins (SOCS) (101,108) that can inhibit cytokine transcriptional factor activation of the insulin receptor [26] causing IR development in borderline tissue.

TNF- α is another mediator in which TNF- α improved interactions between IR and T2DM (122,123). Experimental studies show that TNF- α expression increases in obese animals that modulate insulin action (135). From previous studies it has been found that serum levels of TNF- α are positively correlated with IR pathophysiology (135, 136) indicating that TNF- α is also a key factor contributing to IR development. There are many monocytes, macrophage activity and mediators such as CX3CL1 (fractalkine), CRP 4 Oxidative Medicine and Cellular Longevity, IL-18, MCP-1 (monocyte chemo attractant protein-1), resistin, PAI-1 (plasminogen activator inhibitor -1), E-selectin, and IFN- γ (interferon-gamma)-induced IR (91, 94-98).

Therefore, by making inflammatory processes a therapeutic approach for the management of diabetes (93,103,104). Several studies have reported the importance of anti-inflammatory agents in glucose homeostasis. For example, Goldfine and colleagues in 2010 examined the effects of sugar reduction salsalate (salicylate prodrug) and reported that it was effective in

reducing HbA1c and fasting plasma glucose in T2DM patients (105). Clinical trials have been performed with agents to reduce oxidative stress. Oxidative stress is a major inflammatory event as it stimulates the formation of monocytes and macrophages that promote inflammatory responses involved in insulin resistance and DM (103,106,107). It also regulates the expression of pro cytokines and thus enhances inflammatory mediators (88, 108). Thus, free radical-induced inflammation is one of the possible links between oxidative stress and insulin resistance (99).

e) *Mitochondrial Dysfunction*

Mitochondria are cellular organelles that play a key role in energy production, reactive oxygen species (ROS), mediator signaling (130-133), apoptosis (9,10) calcium storage, heat production, and cell survival and act as part of the signal signal pathways (109,110). Mitochondria are major sources of ROS (134) production that cause mitochondrial dysfunction, insulin resistance and DM (111). Oxidative stress is an important factor contributing to mitochondrial dysfunction (112), which impairs mitochondrial function by altering normal MRC activity, reducing mitochondrial respiratory capacity, increasing proton leakage to MRC,

which alters potential fluid differentiation internal mitochondrial., and reduced the integrity of the mitochondrial layers (113-115). These processes can occur in pancreatic islands and / or systematically in adipocytes and muscle tissue (116). The normal process of glucose uptake depends largely on the body's function of healthy mitochondria that produce the energy needed to receive glucose from borderline tissues (117). Thus, mitochondrial dysfunction significantly reduces ATP cell production and interferes with cellular glucose uptake (88). As a result of these cells they fail to take up circulating glucose in response to insulin leading to insulin resistance (88, 16,118). In addition, oxidative stress can disrupt normal mitochondrial function by increasing the production of mitochondrial fatty acid oxidation and DAG (diacyl glycerol), which also stimulates many serine / threonine kinases leading to IST impairment (88). Thus, stress-dependent mitochondrial dysfunction is another molecular mechanism by which free radicals induce insulin resistance (88, 116,118). Oxidative stress and mitochondrial dysfunction have two interactions where both can produce energy (119).

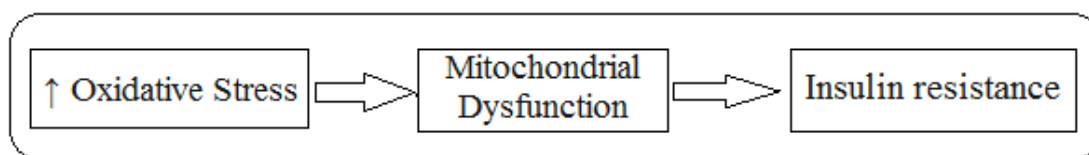


Figure 3: Oxidative stress induces insulin resistance.

VI. CONCLUSION

Diabetes is a metabolic disorder due to hyperglycaemia that can be completely cured but not normally controlled. There are many factors that contribute to the increase and progression of diabetes. Factors can be genetics, stress, obesity, and unhealthy lifestyle etc. But above all oxidative stress plays an important role in the development and progression of diabetes. Oxidation is a chemical process within the human body that leads to the production of free radicals. These oxidative reactions produce free radicals that slowly damage cells and organs by removing inflammatory mediators (such as TNF- α , cytokines, adipokines and chemicals etc.) and causing damage to cell organelles such as mitochondrial damage, damage of ribosomal, nucleus damage etc. leading to insulin resistance. Among other things, oxidative stress increases the rate of disease progression by interfering with insulin signaling pathways resulting in a decrease in insulin sensitivity. Oxidative stress increases apoptosis necrosis leading to beta cell dysfunction leading to insulin secretion. Antioxidants play a very important role in eliminating free radicals. They bind with free oxidative

radicals and remove them from the body by making it harder. Combining antioxidant treatment with standard hypoglycaemic medications will increase recovery rate and antioxidant therapy will help address diabetes problems such as nephrotoxicity, neuropathy and retinopathy, which may be caused by oxidative stress.

REFERENCES RÉFÉRENCES REFERENCIAS

1. Kharroubi, A.T.; Darwish, H.M. Diabetes mellitus: The epidemic of the century. *World J. Diabetes* 2015, 6, 850–867. [CrossRef] [PubMed]
2. H. W. Baynest, "Classification, pathophysiology, diagnosis and management of diabetes mellitus," *Journal of Diabetes & Metabolism*, vol. 6, no. 5, pp. 1–9, 2015.
3. J. M. Forbes and M. E. Cooper, "Mechanisms of diabetic complications," *Physiological Reviews*, vol. 93, no. 1, pp. 137–188, 2013.
4. Schroder, D.; Zuhlke, H. Gene technology, characterization of insulin gene and the relationship to diabetes research. *Endokrinologie* 1982, 79, 197–209.

5. Liu, M.; Weiss, M.A.; Arunagiri, A.; Yong, J.; Rege, N.; Sun, J.; Haataja, L.; Kaufman, R.J.; Arvan, P. Biosynthesis, structure, and folding of the insulin precursor protein. *Diabetes Obes. Metab.* 2018, 20, 28–50. [CrossRef]
6. Halliwell B (2006) Oxidative stress and neurodegeneration. Where are we now? *J. Neurochem*; 97: 1634-1658.
7. Grunewald T, Beal MF (1999) Bioenergetics in Huntington's disease. *Ann N Y AcadSci*; 893: 203-213.
8. H. Yari beygi, F. R. Farrokhi, A. E. Butler, and A. Sahebkar, "Insulin resistance: review of the underlying molecular mechanisms," *Journal of Cellular Physiology*, vol. 234, no. 6, pp. 8152–8161, 2019.
9. Redza-Dutordoir, M.; Averill-Bates, D.A. Activation of apoptosis signalling pathways by reactive oxygen species. *Biochim. Biophys. Acta* 2016, 1863, 2977–2992. [CrossRef]
10. Estaquier, J.; Vallette, F.; Vayssiere, J.-L.; Mignotte, B. The Mitochondrial Pathways of Apoptosis. In *Advances in Mitochondrial Medicine*; Scatena, R., Bottoni, P., Giardina, B., Eds.; Springer: Dordrecht, The Netherlands; pp. 157–183.
11. H. Yari beygi, S. L. Atkin, A. E. Butler, and A. Sahebkar, "Sodium–glucose co transporter inhibitors and oxidative stress: an update," *Journal of Cellular Physiology*, vol. 234, no. 4, pp. 3231–3237, 2019.
12. H. Yari beygi, A. E. Butler, G. E. Barreto, and A. Sahebkar, "Antioxidative potential of antidiabetic agents: a possible protective mechanism against vascular complications in diabetic patients," *Journal of Cellular Physiology*, vol. 234, no. 3, pp. 2436–2446, 2019.
13. H. Yari beygi, M. T. Mohammadi, and A. Sahebkar, "Crocins potentiates antioxidant defense system and improves oxidative damage in liver tissue in diabetic rats," *Biomedicine & Pharmacotherapy*, vol. 98, pp. 333–337, 2018.
14. H. Yari beygi, M. T. Mohammadi, and A. Sahebkar, "PPAR- α agonist improves hyperglycemia-induced oxidative stress in pancreatic cells by potentiating antioxidant defense system," *Drug Research*, vol. 68, no. 6, pp. 355–360, 2018.
15. S. Hurrell and W. H. Hsu, "The etiology of oxidative stress in insulin resistance," *Biomedical Journal*, vol. 40, no. 5, pp. 257–262, 2017.
16. American Diabetes Association, "Diagnosis and classification of diabetes mellitus," *Diabetes Care*, vol. 37, Supplement 1, pp. S81–S90, 2014.
17. J. de Faria Maraschin, "Classification of diabetes," in *Diabetes. Advances in Experimental Medicine and Biology*, vol. 771, S. I. Ahmad, Ed., pp. 12–19, Springer, New York, NY, USA, 2013.
18. K. S. O'Neal, J. L. Johnson, and R. L. Panak, "Recognizing and appropriately treating latent autoimmune diabetes in adults," *Diabetes Spectrum*, vol. 29, no. 4, pp. 249–252, 2016.
19. American Diabetes Association, "Diagnosis and classification of diabetes mellitus," *Diabetes Care*, vol. 33, Supplement 1, pp. S62–S69, 2010.
20. Browne SE, Bowling AC, MacGarvey U, Baik MJ, Berger SC, Muqit MM, Bird ED, Beal MF (1997) Oxidative damage and metabolic dysfunction in Huntington's disease. *Brain Pathol*; 9: 147-163.
21. Donath MY, Boni-Schnetzler M, Ellingsgaard H, Ehses JA. Islet inflammation impairs the pancreatic beta-cell in type 2 diabetes. *Physiology (Bethesda)*. 2009; 24: 325–31.
22. Akash MSH, Shen Q, Rehman K, Chen S. Interleukin-1 receptor antagonist: a new therapy for type 2 diabetes mellitus. *J Pharm Sci*. 2012; 101(5):1647–58.
23. B. Halliwell and J. M. Gutteridge, *Free Radicals in Biology and Medicine*, Oxford University Press, USA, 2015.
24. Festa A, D'Agostino Jr R, Tracy RP, Haffner SM. Elevated levels of acute phase proteins and plasminogen activator inhibitor-1 predict the development of type 2 diabetes: the insulin resistance atherosclerosis study. *Diabetes*. 2002; 51(4): 1131–7.
25. A.C. Maritim, R. A. Sanders, and J. B. Watkins, "Diabetes, oxidative stress, and antioxidants: a review," *Journal of Biochemical and Molecular Toxicology*, vol. 17, no. 1, pp. 24–38, 2003.
26. S. Tangvarasittichai, "Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetes mellitus," *World Journal of Diabetes*, vol. 6, no. 3, pp. 456–480, 2015.
27. R. Radi, A. Denicola, B. Morgan, and J. Zielonka, "Foreword to the free radical biology and medicine special issue on current fluorescence and chemiluminescence approaches in free radical and redox biology," *Free Radical Biology & Medicine*, vol. 128, pp. 1-2, 2018.
28. H. Sies, C. Berndt, and D. P. Jones, "Oxidative stress," *Annual Review of Biochemistry*, vol. 86, pp. 715–748, 2017.
29. J. L. Evans, I. D. Goldfine, B. A. Maddux, and G. M. Grodsky, "Are oxidative stress-activated signaling pathways mediators of insulin resistance and beta-cell dysfunction?," *Diabetes*, vol. 52, no. 1, pp. 1–8, 2003.
30. P. Rösen, P. P. Nawroth, G. King, W. Möller, H. J. Tritschler, and L. Packer, "The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a congress series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes

- Society," *Diabetes/Metabolism Research and Reviews*, vol. 17, no. 3, pp. 189–212, 2001.
31. H. Yaribeygi, F. Lhaf, T. Sathyapalan, and A. Sahebkar, "Effects of novel antidiabetes agents on apoptotic processes in diabetes and malignancy: implications for lowering tissue damage," *Life Sciences*, vol. 231, article 116538, 2019.
 32. H. Yaribeygi, N. Faghihi, M. T. Mohammadi, and A. Sahebkar, "Effects of atorvastatin on myocardial oxidative and nitrosative stress in diabetic rats," *Comparative Clinical Pathology*, vol. 27, no. 3, pp. 691–697, 2018.
 33. Liguori, G. Russo, F. Curcio et al., "Oxidative stress, aging, and diseases," *Clinical Interventions in Aging*, vol. 13, pp. 757–772, 2018.
 34. V. T. Samuel and G. I. Shulman, "The pathogenesis of insulin resistance: integrating signaling pathways and substrate flux," *The Journal of Clinical Investigation*, vol. 126, no. 1, pp. 12–22, 2016.
 35. M. S. Hosseini et al., "The effects of plasma levels of vitamin D3 on insulin resistance and biochemical factors of plasma in patients with type 2 diabetes," *Tehran University Medical Journal*, vol. 75, no. 11, pp. 797–804, 2018.
 36. Færch, D. Vistisen, G. Pacini et al., "Insulin resistance is accompanied by increased fasting glucagon and delayed glucagon suppression in individuals with normal and impaired glucose regulation," *Diabetes*, vol. 65, no. 11, pp. 3473–3481, 2016.
 37. E. Hall, *Guyton and Hall Textbook of Medical Physiology e-Book*, Elsevier Health Sciences, 2015.
 38. V. V. Kiselyov, S. Versteyhe, L. Gauguin, and P. de Meyts, "Harmonic oscillator model of the insulin and IGF1 receptors' allosteric binding and activation," *Molecular Systems Biology*, vol. 5, no. 1, p. 243, 2009.
 39. D. Copps and M. F. White, "Regulation of insulin sensitivity by serine/threonine phosphorylation of insulin receptor substrate proteins IRS1 and IRS2," *Diabetologia*, vol. 55, no. 10, pp. 2565–2582, 2012.
 40. C. K. Ho, G. Sriram, and K. M. Dipple, "Insulin sensitivity predictions in individuals with obesity and type II diabetes mellitus using mathematical model of the insulin signal transduction pathway," *Molecular Genetics and Metabolism*, vol. 119, no. 3, pp. 288–292, 2016.
 41. B. M. Koeppen and B. A. Stanton, *Berne and Levy Physiology e-book*, Elsevier Health Sciences, 2017.
 42. Pickup JC, Crook MA. Is type II diabetes mellitus a disease of the innate immune system? *Diabetologia*. 1998; 41(10): 1241–8.
 43. Jager J, Gremeaux T, Cormont M, Le Marchand-Brustel Y, Tanti JF. Interleukin-1 β -induced insulin resistance in adipocytes through downregulation of insulin receptor substrate-1 expression. *Endocrinology*. 2007; 148(1): 241–51.
 44. L. Mizgier, S. Rutti, M. Pinget, and K. Bouzakri, "Beta-cell function and survival are modulated differentially by type I or type II muscle through specific myokines," *Diabetes*, vol. 67, Supplement 1, pp. 266–2LB, 2018.
 45. E. Seelig, B. Trinh, H. Hanssen et al., "Exercise and the dipeptidyl-peptidase IV inhibitor sitagliptin do not improve beta-cell function and glucose homeostasis in long-lasting type 1 diabetes—a randomised open-label study," *Endocrinology, Diabetes & Metabolism*, vol. 2, no. 3, article e00075, 2019.
 46. D. Porte and S. E. Kahn, "Beta-cell dysfunction and failure in type 2 diabetes: potential mechanisms," *Diabetes*, vol. 50, Supplement 1, pp. S160–S163, 2001.
 47. G. White, J. A. Shaw, and R. Taylor, "Type 2 diabetes: the pathologic basis of reversible β -cell dysfunction," *Diabetes Care*, vol. 39, no. 11, pp. 2080–2088, 2016.
 48. G. Drews, P. Krippeit-Drews, and M. Düfer, "Oxidative stress and beta-cell dysfunction," *Pflügers Archiv-European Journal of Physiology*, vol. 460, no. 4, pp. 703–718, 2010.
 49. J. F. Turrens, "Mitochondrial formation of reactive oxygen species," *The Journal of Physiology*, vol. 552, no. 2, pp. 335–344, 2003.
 50. Newsholme, D. Morgan, E. Rebelato et al., "Insights into the critical role of NADPH oxidase (s) in the normal and dysregulated pancreatic beta cell," *Diabetologia*, vol. 52, no. 12, pp. 2489–2498, 2009.
 51. Y. Uchizono, R. Takeya, M. Iwase et al., "Expression of isoforms of NADPH oxidase components in rat pancreatic islets," *Life Sciences*, vol. 80, no. 2, pp. 133–139, 2006.
 52. G. Lenaz, "The mitochondrial production of reactive oxygen species: mechanisms and implications in human pathology," *IUBMB Life*, vol. 52, no. 3-5, pp. 159–164, 2001.
 53. D. L. Eizirik, M. L. Colli, and F. Ortis, "The role of inflammation in insulinitis and β -cell loss in type 1 diabetes," *Nature Reviews Endocrinology*, vol. 5, no. 4, pp. 219–226, 2009.
 54. A. A. Starkov, B. M. Polster, and G. Fiskum, "Regulation of hydrogen peroxide production by brain mitochondria by calcium and Bax," *Journal of Neurochemistry*, vol. 83, no. 1, pp. 220–228, 2002.
 55. A. Gerber and G. A. Rutter, "The role of oxidative stress and hypoxia in pancreatic beta-cell dysfunction in diabetes mellitus," *Antioxidants & Redox Signaling*, vol. 26, no. 10, pp. 501–518, 2017.
 56. P. Robertson and J. S. Harmon, "Pancreatic islet β -cell and oxidative stress: the importance of glutathione peroxidase," *FEBS Letters*, vol. 581, no. 19, pp. 3743–3748, 2007.
 57. G. Drews, C. Krämer, M. Düfer, and P. Krippeit-Drews, "Contrasting effects of alloxan on islets and

- single mouse pancreatic β -cells," *Biochemical Journal*, vol. 352, no. 2, pp. 389–397, 2000.
58. P. Krippeitdrews, S. Britsch, F. Lang, and G. Drews, "Effects of SH-group reagents on Ca^{2+} and K^{+} channel currents of pancreatic B-cells," *Biochemical and Biophysical Research Communications*, vol. 200, no. 2, pp. 860–866, 1994.
 59. M. S. Islam, P.O. Berggren, and O. Larsson, "Sulfhydryl oxidation induces rapid and reversible closure of the ATP regulated K^{+} channel in the pancreatic β -cell," *FEBS Letters*, vol. 319, no. 1-2, pp. 128–132, 1993.
 60. P. Robertson, "Oxidative stress and impaired insulin secretion in type 2 diabetes," *Current Opinion in Pharmacology*, vol. 6, no. 6, pp. 615–619, 2006.
 61. B. Gier, P. Krippeit-Drews, T. Sheiko et al., "Suppression of KATP channel activity protects murine pancreatic β cells against oxidative stress," *The Journal of Clinical Investigation*, vol. 119, no. 11, pp. 3246–3256, 2009.
 62. J. Wang and H. Wang, "Oxidative stress in pancreatic beta cell regeneration," *Oxidative Medicine and Cellular Longevity*, vol. 2017, Article ID 1930261, 9 pages, 2017.
 63. E. Hussey, S. L. McGee, A. Garnham, G. K. McConell, and M. Hargreaves, "Exercise increases skeletal muscle GLUT4 gene expression in patients with type 2 diabetes," *Diabetes, Obesity and Metabolism*, vol. 14, no. 8, pp. 768–771, 2012.
 64. E. A. Richter and M. Hargreaves, "Exercise, GLUT4, and skeletal muscle glucose uptake," *Physiological Reviews*, vol. 93, no. 3, pp. 993–1017, 2013.
 65. C. M. Reno, E. C. Puente, Z. Sheng et al., "Brain GLUT4 knockout mice have impaired glucose tolerance, decreased insulin sensitivity, and impaired hypoglycemic counter regulation," *Diabetes*, vol. 66, no. 3, pp. 587–597, 2017.
 66. M. Gaster, P. Staehr, H. Beck-Nielsen, H. D. Schröder, and A. Handberg, "GLUT4 is reduced in slow muscle fibers of type 2 diabetic patients: is insulin resistance in type 2 diabetes a slow, type 1 fiber disease?," *Diabetes*, vol. 50, no. 6, pp. 1324–1329, 2001.
 67. D. J. O'Gorman, H. K. R. Karlsson, S. McQuaid et al., "Exercise training increases insulin-stimulated glucose disposal and GLUT4 (SLC2A4) protein content in patients with type 2 diabetes," *Diabetologia*, vol. 49, no. 12, pp. 2983–2992, 2006.
 68. G. Boden, C. Homko, C. A. Barrero et al., "Excessive caloric intake acutely causes oxidative stress, GLUT4 carbonylation, and insulin resistance in healthy men," *Science Translational Medicine*, vol. 7, no. 304, article 304re7, 2015.
 69. P. Manna, A. E. Achari, and S. K. Jain, "Vitamin D supplementation inhibits oxidative stress and upregulate SIRT1/ AMPK/GLUT4 cascade in high glucose-treated 3T3L1 adipocytes and in adipose tissue of high fat diet-fed diabetic mice," *Archives of Biochemistry and Biophysics*, vol. 615, pp. 22–34, 2017.
 70. D. Pessler, A. Rudich, and N. Bashan, "Oxidative stress impairs nuclear proteins binding to the insulin responsive element in the GLUT4 promoter," *Diabetologia*, vol. 44, no. 12, pp. 2156–2164, 2001.
 71. D. J. Fazakerley, A. Y. Minard, J. R. Krycer et al., "Mitochondrial oxidative stress causes insulin resistance without disrupting oxidative phosphorylation," *The Journal of Biological Chemistry*, vol. 293, no. 19, pp. 7315–7328, 2018.
 72. A. Rudich, A. Tirosh, R. Potashnik, R. Hemi, H. Kanety, and N. Bashan, "Prolonged oxidative stress impairs insulin induced GLUT4 translocation in 3T3-L1 adipocytes," *Diabetes*, vol. 47, no. 10, pp. 1562–1569, 1998.
 73. D. W. Cooke and M. D. Lane, "The transcription factor nuclear factor I mediates repression of the GLUT4 promoter by insulin," *The Journal of Biological Chemistry*, vol. 274, no. 18, pp. 12917–12924, 1999.
 74. H. She and Z. Mao, "Regulation of myocyte enhancer factor-2 transcription factors by neurotoxins," *Neurotoxicology*, vol. 32, no. 5, pp. 563–566, 2011.
 75. J. V. Esteves, F. J. Enguita, and U. F. Machado, "Micro RNAs mediated regulation of skeletal muscle GLUT4 expression and translocation in insulin resistance," *Journal of Diabetes Research*, vol. 2017, Article ID 7267910, 11 pages, 2017.
 76. J. He and B.-H. Jiang, "Interplay between reactive oxygen species and micro RNAs in cancer," *Current Pharmacology Reports*, vol. 2, no. 2, pp. 82–90, 2016.
 77. J. Matsuzaki and T. Ochiya, "Extracellular microRNAs and oxidative stress in liver injury: a systematic mini review," *Journal of Clinical Biochemistry and Nutrition*, vol. 63, no. 1, pp. 6–11, 2018.
 78. Z. Wang, Y. Liu, N. Han et al., "Profiles of oxidative stress related micro RNA and mRNA expression in auditory cells," *Brain Research*, vol. 1346, pp. 14–25, 2010.
 79. C. Jolival, C. A. Lee, K. K. Beiswenger et al., "Defective insulin signaling pathway and increased glycogen synthase kinase-3 activity in the brain of diabetic mice: parallels with Alzheimer's disease and correction by insulin," *Journal of Neuroscience Research*, vol. 86, no. 15, pp. 3265–3274, 2008.
 80. R. Mackenzie and B. Elliott, "Akt/PKB activation and insulin signaling: a novel insulin signaling pathway in the treatment of type 2 diabetes," *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 7, p. 55, 2014.

81. M. J. Birnbaum, "Turning down insulin signaling," *The Journal of Clinical Investigation*, vol. 108, no. 5, pp. 655–659, 2001.
82. K. Paz, R. Hemi, D. LeRoith et al., "A molecular basis for insulin resistance. Elevated serine/threonine phosphorylation of IRS-1 and IRS-2 inhibits their binding to the juxta membrane region of the insulin receptor and impairs their ability to undergo insulin-induced tyrosine phosphorylation," *The Journal of Biological Chemistry*, vol. 272, no. 47, pp. 29911–29918, 1997.
83. B. A. Maddux, W. See, Lawrence JC Jr, A. L. Goldfine, I. D. Goldfine, and J. L. Evans, "Protection against oxidative stress—induced insulin resistance in rat L6 muscle cells by micromolar concentrations of α -lipoic acid," *Diabetes*, vol. 50, no. 2, pp. 404–410, 2001.
84. A. S. Blair, E. Hajduch, G. J. Litherland, and H. S. Hundal, "Regulation of glucose transport and glycogen synthesis in L6 muscle cells during oxidative stress evidence for crosstalk between the insulin and SAPK2/p38 mitogenactivated protein kinase signaling pathways," *The Journal of Biological Chemistry*, vol. 274, no. 51, pp. 36293–36299, 1999.
85. V. Aguirre, T. Uchida, L. Yenush, R. Davis, and M. F. White, "The c-Jun NH (2)-terminal kinase promotes insulin resistance during association with insulin receptor substrate-1 and phosphorylation of Ser (307)," *The Journal of Biological Chemistry*, vol. 275, no. 12, pp. 9047–9054, 2000.
86. J. W. Eriksson, "Metabolic stress in insulin's target cells leads to ROS accumulation—a hypothetical common pathway causing insulin resistance," *FEBS Letters*, vol. 581, no. 19, pp. 3734–3742, 2007.
87. M. Balbaa, S. A. Abdulmalek, and S. Khalil, "Oxidative stress and expression of insulin signaling proteins in the brain of diabetic rats: role of Nigella sativa oil and antidiabetic drugs," *PLoS One*, vol. 12, no. 5, article e0172429, 2017.
88. J. L. Rains and S. K. Jain, "Oxidative stress, insulin signaling, and diabetes," *Free Radical Biology & Medicine*, vol. 50, no. 5, pp. 567–575, 2011.
89. A. Bloch-Damti and N. Bashan, "Proposed mechanisms for the induction of insulin resistance by oxidative stress," *Antioxidants & Redox Signaling*, vol. 7, no. 11–12, pp. 1553–1567, 2005.
90. Talior, M. Yarkoni, N. Bashan, and H. Eldar-Finkelman, "Increased glucose uptake promotes oxidative stress and PKC- δ activation in adipocytes of obese, insulin resistant mice," *American Journal of Physiology- Endocrinology and Metabolism*, vol. 285, no. 2, pp. E295–E302, 2003.
91. R. B. Goldberg, "Cytokine and cytokine-like inflammation markers, endothelial dysfunction, and imbalanced coagulation in development of diabetes and its complications," *The Journal of Clinical Endocrinology & Metabolism*, vol. 94, no. 9, pp. 3171–3182, 2009.
92. H. Yaribeygi, N. Katsiki, A. E. Butler, and A. Sahebkar, "Effects of antidiabetic drugs on NLRP3 inflammasome activity, with a focus on diabetic kidneys," *Drug Discovery Today*, vol. 24, no. 1, pp. 256–262, 2019.
93. R. J. Perry, J. G. Camporez, R. Kursawe et al., "Hepatic acetyl CoA links adipose tissue inflammation to hepatic insulin resistance and type 2 diabetes," *Cell*, vol. 160, no. 4, pp. 745–758, 2015.
94. Sindhu, N. Akhter, H. Arefanian et al., "Increased circulatory levels of fractalkine (CX3CL1) are associated with inflammatory chemokines and cytokines in individuals with type-2 diabetes," *Journal of Diabetes & Metabolic Disorders*, vol. 16, no. 1, p. 15, 2017.
95. S. Gupta, A. Maratha, J. Siednienko et al., "Analysis of inflammatory cytokine and TLR expression levels in type 2 diabetes with complications," *Scientific Reports*, vol. 7, no. 1, article 7633, 2017.
96. C. B. Guest, M. J. Park, D. R. Johnson, and G. G. Freund, "The implication of Proinflammatory cytokines in type 2 diabetes," *Frontiers in Bioscience*, vol. 13, no. 1, pp. 5187–5194, 2008.
97. S. Basu, A. Larsson, J. Vessby, B. Vessby, and C. Berne, "Type 1 diabetes is associated with increased cyclooxygenase and cytokine-mediated inflammation," *Diabetes Care*, vol. 28, no. 6, pp. 1371–1375, 2005.
98. Y. Benomar, A. Gertler, P. de Lacy et al., "Central resistin overexposure induces insulin resistance through toll-like receptor 4," *Diabetes*, vol. 62, no. 1, pp. 102–114, 2013.
99. N. Keane, V. F. Cruzat, R. Carlessi, P. I. H. de Bittencourt, and P. Newsholme, "Molecular events linking oxidative stress and inflammation to insulin resistance and β -cell dysfunction," *Oxidative Medicine and Cellular Longevity*, vol. 2015, Article ID 181643, 15 pages, 2015.
100. S. J. Richardson, A. Willcox, A. J. Bone, A. K. Foulis, and N. G. Morgan, "Islet-associated macrophages in type 2 diabetes," *Diabetologia*, vol. 52, no. 8, pp. 1686–1688, 2009.
101. Kawazoe Y, Naka T, Fujimoto M, Kohzaki H, Morita Y, Narazaki M, Okumura K, Saitoh H, Nakagawa R, Uchiyama Y, Akira S, Kishimoto T. Signal transducer and activator of transcription (STAT)-induced STAT inhibitor 1 (SSI-1)/ suppressor of cytokine signaling 1 (SOCS1) inhibits insulin signal transduction pathway through modulating insulin receptor substrate 1 (IRS-1) phosphorylation. *J Exp Med*. 2001; 193(2): 263–9.
102. M. Krause, J. Rodrigues-Krause, C. O'Hagan et al., "Differential nitric oxide levels in the blood and skeletal muscle of type 2 diabetic subjects may be

- consequence of adiposity: a preliminary study," *Metabolism*, vol. 61, no. 11, pp. 1528–1537, 2012.
103. H. Bae, C. H. Jeong, W. N. Cheng, K. Hong, H. G. Seo, and S. G. Han, "Oxidative stress-induced inflammatory responses and effects of N-acetylcysteine in bovine mammary alveolar cells," *Journal of Dairy Research*, vol. 84, no. 4, pp. 418–425, 2017.
 104. S. P. Weisberg, R. Leibel, and D. V. Tortoriello, "Dietary curcumin significantly improves obesity-associated inflammation and diabetes in mouse models of diabetes," *Endocrinology*, vol. 149, no. 7, pp. 3549–3558, 2008.
 105. A. B. Goldfine, V. Fonseca, K. A. Jablonski et al., "The effects of salsalate on glycemic control in patients with type 2 diabetes: a randomized trial," *Annals of Internal Medicine*, vol. 152, no. 6, pp. 346–357, 2010.
 106. A. Zhang, Q. Shen, Y. Chen et al., "Myricitrin alleviates oxidative stress-induced inflammation and apoptosis and protects mice against diabetic cardiomyopathy," *Scientific Reports*, vol. 7, no. 1, article 44239, 2017.
 107. H. Ma, S. Y. Li, P. Xu et al., "Advanced glycation endproduct (AGE) accumulation and AGE receptor (RAGE) upregulation contribute to the onset of diabetic cardiomyopathy," *Journal of Cellular and Molecular Medicine*, vol. 13, no. 8b, pp. 1751–1764, 2009.
 108. Emanuelli B, Peraldi P, Filloux C, Sawka-Verhelle D, Hilton D, Van Obberghen E. SOCS-3 is an insulin-induced negative regulator of insulin signaling. *J Biol Chem*. 2000;275(21):15985–91. 26. Krebs DL, Hilton DJ. SOCS: physiological suppressors of cytokine signaling. *J Cell Sci*. 2000; 113(Pt 16): 2813–9.
 109. R. S. Balaban, S. Nemoto, and T. Finkel, "Mitochondria, oxidants, and aging," *Cell*, vol. 120, no. 4, pp. 483–495, 2005.
 110. W. Elrod and Å. B. Gustafsson, Editorial Overview: Mitochondria Biology, Elsevier, 2018.
 111. K. Montgomery and N. Turner, "Mitochondrial dysfunction and insulin resistance: an update," *Endocrine Connections*, vol. 4, no. 1, pp. R1–R15, 2015.
 112. S. Rose, R. E. Frye, J. Slattery et al., "Oxidative stress induces mitochondrial dysfunction in a subset of autistic lymphoblastoid cell lines," *Translational Psychiatry*, vol. 4, no. 4, article e377, 2014.
 113. J. Wada and A. Nakatsuka, "Mitochondrial dynamics and mitochondrial dysfunction in diabetes," *Acta Medica Okayama*, vol. 70, no. 3, pp. 151–158, 2016.
 114. S. Rose, R. E. Frye, J. Slattery et al., "Erratum: Oxidative stress induces mitochondrial dysfunction in a subset of autistic lymphoblastoid cell lines," *Translational Psychiatry*, vol. 5, no. 3, article e526, 2015.
 115. A. Agil, M. el-Hammadi, A. Jiménez-Aranda et al., "Melatonin reduces hepatic mitochondrial dysfunction in diabetic obese rats," *Journal of Pineal Research*, vol. 59, no. 1, pp. 70–79, 2015.
 116. D. Brand and D. G. Nicholls, "Assessing mitochondrial dysfunction in cells," *Biochemical Journal*, vol. 435, no. 2, pp. 297–312, 2011.
 117. L. Tokarz, P. E. MacDonald, and A. Klip, "The cell biology of systemic insulin function," *The Journal of Cell Biology*, vol. 217, no. 7, pp. 2273–2289, 2018.
 118. S. Supale, N. Li, T. Brun, and P. Maechler, "Mitochondrial dysfunction in pancreatic β cells," *Trends in Endocrinology & Metabolism*, vol. 23, no. 9, pp. 477–487, 2012.
 119. S. Sifuentes-Franco, F. P. Pacheco-Moisés, A. D. Rodríguez-Carrizalez, and A. G. Miranda-Díaz, "The role of oxidative stress, mitochondrial function, and autophagy in diabetic polyneuropathy," *Journal of Diabetes Research*, vol. 2017, Article ID 1673081, 15 pages, 2017.
 120. Kern PA, Ranganathan S, Li C, Wood L, Ranganathan G. Adipose tissue tumor necrosis factor and interleukin-6 expression in human obesity and insulin resistance. *Am J Physiol Endocrinol Metab*. 2001; 280(5): E745–751.
 121. Kirwan JP, Jing M. Modulation of insulin signaling in human skeletal muscle in response to exercise. *Exerc Sport Sci Rev*. 2002; 30(2): 85–90.
 122. Nieto-Vazquez I, Fernandez-Veledo S, Kramer DK, Vila-Bedmar R, GarciaGuerra L, Lorenzo M. Insulin resistance associated to obesity: the link TNF α . *Arch Physiol Biochem*. 2008;114(3):183–94.
 123. Swaroop JJ, Rajarajeswari D, Naidu JN. Association of TNF- α with insulin resistance in type 2 diabetes mellitus. *Indian J Med Res*. 2012;135:127–30.
 124. Choquet, H.; Meyre, D. Genetics of Obesity: What have we Learned? *Curr. Genom*. 2011, 12, 169–179. [CrossRef]
 125. Dodson, G.; Steiner, D. The role of assembly in insulin's biosynthesis. *Curr. Opin. Struct. Biol*. 1998, 8, 189–194. [CrossRef]
 126. Matschinsky, F.M. Regulation of pancreatic beta-cell glucokinase: From basics to therapeutics. *Diabetes* 2002, 51, S394–S404. [CrossRef] [PubMed]
 127. Matschinsky, F.M. Glucokinase as glucose sensor and metabolic signal generator in pancreatic beta-cells and hepatocytes. *Diabetes* 1990, 39, 647–652. [CrossRef]
 128. Meglasson, M.D.; Matschinsky, F.M. Pancreatic islet glucose metabolism and regulation of insulin secretion. *Diabetes Metab. Rev*. 1986, 2, 163–214. [CrossRef]

129. Newgard, C.B.; McGarry, J.D. Metabolic coupling factors in pancreatic beta-cell signal transduction. *Annu. Rev. Biochem.* 1995, 64, 689–719. [CrossRef]
130. Daiber, A.; Di Lisa, F.; Oelze, M.; Kröller-Schön, S.; Steven, S.; Schulz, E.; Münzel, T. Crosstalk of mitochondria with NADPH oxidase via reactive oxygen and nitrogen species signalling and its role for vascular function. *Br. J. Pharmacol.* 2017, 174, 1670–1689. [CrossRef] [PubMed]
131. Angelova, P.R.; Abramov, A.Y. Functional role of mitochondrial reactive oxygen species in physiology. *Free Radic. Biol. Med.* 2016, 100, 81–85. [CrossRef] [PubMed]
132. Owusu-Ansah, E.; Banerjee, U. Reactive oxygen species prime *Drosophila* haematopoietic progenitors for differentiation. *Nature* 2009, 461, 537–541. [CrossRef]
133. Wang, W.; Fang, H.; Groom, L.; Cheng, A.; Zhang, W.; Liu, J.; Wang, X.; Li, K.; Han, P.; Zheng, M.; et al. Superoxide flashes in single mitochondria. *Cell* 2008, 134, 279–290. [CrossRef]
134. Shigenaga MK, Hagen TM, Ames BN. Oxidative damage and mitochondrial decay in aging. *Proc Natl Acad Sci U S A.* 1994; 91: 10771–10778. Crossref. PubMed.
135. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor- α : direct role in obesity-linked insulin resistance. *Science.* 1993; 259(5091): 87–91.
136. Sharma R, Anker SD. Cytokines, apoptosis and cachexia: the potential for TNF antagonism. *Int J Cardiol.* 2002; 85(1): 161–71.

