

1      Role of Detoxifying Enzyme Nicotinamide Adenine Dinucleotide  
2      (Phosphate) H: Quinone Oxidoreductase-1 C609T Gene  
3      Polymorphism in Bronchogenic Carcinoma

Nermine Hossam Zakaria<sup>1</sup>

<sup>1</sup> Faculty of Medicine Alexandria University

Received: 9 December 2013 Accepted: 31 December 2013 Published: 15 January 2014

## 8 Abstract

9 Lung cancer is currently one of the most common cancers and a major cause of cancer-related  
10 death in the world. Eighty-five percent of lung cancers are non-small cell lung cancers  
11 (NSCLCs), and 15

*Index terms*—

## 1 Introduction

lung cancer is currently one of the most common cancers and a major cause of cancer-related death in the world. Among males, the highest lung cancer incidence rates are in Central and Eastern Europe 53.5 (per 100,000) and 50.4 in Eastern Asia. Among females, the highest lung cancer incidence rates are 33.8 in Northern America and 23.7 Northern Europe. ??1) Incidence data (ASR) for the Arab countries; for males, lung cancer incidence estimated as; 31.1 (per 100,000) in Tunisia followed by Lebanon, Libya, Jordan and in Egypt 11.21 (per 100,000). For females, estimated as; 11.0 in Lebanon followed by Bahrain, Syrian Ar -ab Repubic, United Arab Emirates, and in Egypt it reaches 3.76 (per 100,000). ??1) Eighty-five percent of lung cancers are nonsmall cell lung cancers (NSCLCs), and 15% are small cell lung cancers (SCLCs). (2) The most important risk factor for lung cancer is tobacco smoking. International variations in lung cancer rates and trends largely reflect differences in the stage and degree of the tobacco epidemic because smoking accounts for about 80% of global lung cancer deaths in men and 50% of the deaths in women. (3) Many of the compounds in tobacco smoke are oxidized by phase I enzymes into reactive metabolites, which are detoxified by phase II enzymes. Polycyclic aromatic hydrocarbons (PAH) are abundant in tobacco smoke and constitute a major etiological factor in lung cancer. (4) NQO1 [NAD (P): H-(quinone acceptor) oxidore ductase; EC 1.6.99.2] (5) enzyme is a homodimeric flavin adenine dinucleotide (FAD) containing cytosolic protein catalyzing the two-electron reduction of quinone substrates. (6) Quinine compounds are mainly derived from endogenous quinones, such as vitamin E quinine, and exogenous quinones, such as exhaust gas, tobacco smoke. (7) NQO1 prevents the generation of free radicals and reactive oxygen, thus protecting the cells from oxidative damage. This pathway is thought to be the major mechanism responsible for modifying the toxicity of quinones, including those arising from the formation of DNA adducts induced by benzo(a)pyrene 3,6-quinone, one of the most potent polycyclic aromatic hydrocarbons present in tobacco smoke. (8) The NQO1 gene is located on chromosome 16q22.1, spanning 17.2 kb and consisting of 6 exons and 5 introns. (9) The polymorphic variant is a C?T transition at nucleotide position 609 (amino acid codon 187) that results in a proline-to-serine amino acid substitution in the protein. The reference number of this SNP in the database of the National Center for Biotechnology Information (NCBI) is rs1800566. ??10) Three genotypes of NQO1 are known to be associated with different enzymatic activities: C/C is the homozygous wild-type with normal activity, C/T is heterozygous with reduced activity, and T/T is the homozygous variant with only 2-4% of the enzyme activity of the wild-type. (11) Several studies have examined the relationship between the NQO1 genetic polymorphism and lung cancer risk, but the conclusions have been inconsistent. (9,12,13) The study of the role of SNPs and in particular the NQO1 609C?T polymorphism as a risk factor for the development of bronchogenic carcinoma among Egyptian male patients is of utmost importance for the unmasking of the risk factors underlying bronchogenic carcinoma in Egypt.

## 6 RESULTS

---

### 46 2 II.

### 47 3 Patients and Methods

48 The current study was conducted as a hospitalbased case-control study of 200 subjects; 100 male patients  
49 presenting to the Chest diseases unit in the Alexandria Main University Hospital with bronchogenic carcinoma  
50 in different stages of the disease between 2011 and 2013. Lung cancer cases were newly diagnosed cases aged  
51 above 40 years old with absence of previous precancerous conditions and any known primary cancerous lesion  
52 elsewhere. Diagnosis of primary lung cancer was confirmed through a review of each patient's pathology report by  
53 the Alexandria University Hospital pathology department. One hundred age-matched male adults from the same  
54 socioeconomic class were recruited. All patients and controls provided informed consent. a) DNA extraction and  
55 genotyping analysis DNA was extracted from EDTA whole blood using DNA purification kit (QIAamp DNA  
56 blood Mini Kit, Qiagen, Hilden, Germany). The extraction was done according to the manufacturer's instructions  
57 using the Spin Protocol. The quantity and purity of DNA for each sample was assayed using Nanodrop 2000  
58 spectrophotometer (Thermo scientific, USA). The final concentration of target DNA was adjusted to 50-100 ng  
59 in the following amplification reaction to exclude variability in DNA concentration.

60 DNA amplification was conducted by polymerase chain reaction with confronting two-pair primers  
61 (PCRCTPP) (14) , using four primers; F1: 5'-CCT TAT CAGAGT GTC TTA CTG AGA-3' (54.4°C) and  
62 R1: 5'-CAA TGCTAT ATG TCA GTT GAG G-3' (54.7°C), for C allele amplifying a 165-bp band, as well as  
63 F2: 5'-GTG GCT TCC AAG TCTTAG AAT-3' (54.9°C) and R2: 5'-TTT CTA GCT TTG ATC TGGTTG-5'  
64 (54.5°C) for T allele amplifying a 283-bp band. A common 406-bp band was designed to be amplified between  
65 primers F1 and R2.

66 The results of PCR-CTPP genotyping were confirmed by PCR-RFLP with HinF1 enzyme, which produces  
67 188-bp and 85-bp bands for C allele and 151bp and 85-bp bands for T allele using primers F: 5'-AGT GGCATT  
68 CTG CAT TTC TGT G-3', and R: 5'-GAT GGA CTT GCCCAA GTG ATG-3'. (14) Two hundred samples  
69 were successfully genotyped; 100 lung cancer patients (62 were CC, 31 were CT, and 7 were TT) and 100 healthy  
70 individuals as a control group (73 were CC, 26 were CT, and 1 was TT), The results of PCR-CTPP were  
71 consistent with those of the gold standard method PCR-RFLP.

### 72 4 b) Statistical analysis

73 All statistical analyses were performed using the SPSS20.0 program. An effect was considered statistically  
74 significant at P<0.05. X<sup>2</sup> test was cond -ucted for examining the Hardy-Weinberg equilibrium and independence  
75 of genotype frequency between cases and controls. Logistic regression analysis was then performed to compute  
76 odds ratios and 95% confidence intervals, after adjustment for age, family history and smoking index pack-years  
77 (number of years smo -ked×number of packs smoked per day).

### 78 5 III.

### 79 6 Results

80 The study population was Egyptians, they consisted of 100 male lung cancer patients and 100 male of age-matched  
81 healthy controls. The mean age was  $54.2 \pm 8.2$  years for cases and  $52.7 \pm 8.4$  years for controls. There was  
82 a statistically significant difference between the cases and controls concerning the smoking status and smoking  
83 index ( $p<0.001$ ).Cases were more likely than controls to be current smokers. The mean Pack-Years were  $43.9 \pm 20.8$  among cases and  $19.5 \pm 14.3$  among controls. Twenty three (23%) of the patients had positive family  
85 history of cancer compared to 4% of the control group with a statistically significant difference ( $p<0.001$ ).

86 In the group of lung cancer patients, the most prevalent histological subtype of lung cancer was Adenocarcinoma  
87 with a frequency of 37%, followed by 31% squamous cell carcinoma, 19% SCLC (small cell lung cancer) and 13%  
88 with other histological subtypes (7 cases were large cell anaplastic carcinoma, 4 cases were undifferentiated  
89 NSCLC and 2 cases were carcinoid tumor). As regards the tumour staging, 5 out of 100 patients were stage I,  
90 27 patients were stage II, 32 patients were stage III and 36 patients were stage IV.

91 All included subjects were analyzed for the NQO1 609C?T (rs1800566) SNP genotype. The distributions of the  
92 NQO1 genotypes among lung cancer patients and controls were in agreement with Hardy- Weinberg equilibrium  
93 ( $p= 0.267, 0.425$ , respectively). The frequency of NQO1 CC, CT, TT genotypes was 62, 31 and 7%, respectively  
94 in patients and 73, 26 and 1% in controls. The relative frequencies of the wild (C) allele and the variant (T) allele  
95 of NQO1 609C?T (rs1800566) SNP were 77.5% and 22.5%, among the patients respectively, and were 86% and  
96 14% among the controls, respectively. There was a significant difference among the two groups as regards their  
97 allele frequencies ( $p =0.028$ ). Patients with one or two copies of the T variant allele had 2.2 -fold increased lung  
98 cancer risk than those with CC genotype (adjusted OR=2.2; 95%CI: 0.63-7.9). To assess whether the NQO1  
99 variant T allele may impart different risks for the various lung cancer cell types, we computed the adjusted ORs  
100 for lung cancer for each histological type. Patients with one or two copies of the T variant allele had 4.3-fold  
101 increased SCLC risk than those with CC genotype (adjusted OR=4.3; 95%CI: 0.57-33.4), while, they had 3.8-fold  
102 increased risk for lung cancer with other histological type (adjusted OR=3.8; 95%CI:0.49-30.7), 2.1-fold increased  
103 adenocarcinoma risk (adjusted OR=2.1; 95%CI: 0.43-10.3), and 1.2-fold increased squamous cell carcinoma risk  
104 more than those with CC genotype (adjusted OR=1.2; 95%CI: 0.21-7.0). The study subjects were stratified by

105 age and smoking status. As regards study subject's age, elevated lung cancer risk associated with TT genotype  
106 was evident in younger individuals (age  $\leq$ 60) (adjusted OR=13.6; 95%CI: 0.55-58.7). On the contrary, elevated  
107 lung cancer risk associated with CT genotype individually and with one or two copies of the T variant allele (CT  
108 and TT combined) were evident in older individuals (age  $\geq$ 60) ((adjusted OR=9.1; 95%CI: 0.90-91.7), (adjusted  
109 OR=10.6; 95%CI: 1.1-58.7), respectively). The only significant OR was for the combined CT and TT genotype  
110 group above 60 years.

111 When adjustment was made for smoking index pack-years (light smokers and heavy smokers), among light  
112 smokers ( $<21$  p-y), the patients with one or two copies of the T variant allele had 1.9-fold increased lung Volume  
113 XIV Issue III Version I Year ( ) K cancer risk compared to those with CC genotype (adjusted OR=1.9; 95%CI:  
114 0.29-13.2), while, the heavy smoker ( $>21$  p-y) patients with one or two copies of the T variant allele had 3.6-  
115 fold increased lung cancer risk compared to those with CC genotype (adjusted OR=3.6; 95%CI: 0.41-30.7).  
116 The patients with one or two copies of the T variant allele had 3.6-fold increased lung cancer risk compared  
117 to those with CC genotype (adjusted OR=3.6; 95%CI: 0.41-30.7), while, the risk for squamous cell carcinoma  
118 was 2.4-fold (adjusted OR=2.4; 95%CI: 0.21-27.9), 17.9-fold for SCLC (adjusted OR=17.9; 95%CI: 1.5-40.6),  
119 2.4-fold for adenocarcinoma (adjusted OR=2.4; 95%CI: 0.25-23.8), and 2.5-fold for lung cancer other histological  
120 type (adjusted OR=2.5; 95%CI: 0.18-34.4). The odds ratio of the combined SCLC heavy smokers group was  
121 statistically significant. # OR adjusted for age and family history \* OR significant at 0.05

122 Assessing the efficiency of CTPP-PCR considering allelic discrimination through genotyping analysis of NQO1  
123 609C?T (rs1800566) SNP, comparing CTPP-PCR results with those of RFLP-PCR as a goldstandard method.  
124 The efficiency of CTPP-PCR was assessed in 200 samples; 100 lung cancer patients and 100 healthy individuals  
125 as a control group. There was no discrepancy between the CTPP-PCR and RFLP-PCR results in all our study  
126 subject samples. The previous performance reflects a sensitivity of 100% and specificity of 100%. The positive  
127 predictive value (PPV) and the negative predictive value (NPV) were 100%.

## 128 7 IV.

## 129 8 Discussion

130 Cancer lung is one of the leading causes of cancer related mortalities worldwide. The rapidly evolving field of  
131 cancer genetics has opened up new possibilities for the discovery of susceptibility genes for numerous cancers  
132 including lung cancer.

133 Lung cancer has been the most common cancer worldwide since 1985, both in terms of incidence and mortality.  
134 The 5-year survival rate in the United States for lung cancer is 15.6 %. (15) Cigarette smoking is the main risk  
135 factor for lung cancer, accounting for about 90% of the cases in men and 70% of the cases in women. (16) The  
136 pathogenesis of lung cancer had a genetic component, whether it relates to host susceptibility to lung cancer,  
137 with or without exposure to cigarette smoke to the development of certain types of lung cancer. (15) NAD (P)  
138 H: quinone oxidoreductase 1 (NQO1) is a two-electron reductase, which reduces reactive quinones to less reactive  
139 and less toxic hydroquinones, resulting in protection of the cells. (13) The quinones are mainly derived from  
140 endogenous quinones, such as vitamin E quinone and exogenous quinones, such as tobacco smoke. (17) The  
141 T/T (Ser/Ser) variable allele of NQO1 lacks enzymatic activity and fails to detoxify quinone metabolites into  
142 the reduced form. (13) It was thus hypothesized that individuals lacking NQO1activity would be at high risk  
143 of malignancies (lung cancer) because of the exposure to procarcinogens, that are included in cigarette smoke,  
144 which are oxidized to quinone metabolites. (6) In this study, we analyzed the relationship between NQO1genetic  
145 polymorphisms and lung cancer, comparing lung cancer patients and healthy individuals in Egypt. Our results  
146 showed, the frequencies of the wild (C) allele and the variant (T) allele of NQO1 609C?T (rs1800566) SNP for  
147 the patients were 77.5% and 22.5%, respectively, and for the controls were 86% and 14%, respectively. The allele  
148 frequencies in both cases and controls were in Hardy-Weinberg equilibrium. The frequency of the variant allele  
149 in Egyptians was similar to the frequencies reported in another study on Arab population. (18) The NQO1  
150 609C?T polymorphism exhibits ethnic variation (4-22%) with the highest prevalence of the T allele occurring  
151 in Asian populations and the lowest in Caucasians (4%), while, in Arabs TT genotype frequency was 6.4%  
152 (Middle Eastern Arab origin (95% Saudi Arabians and 5% from other Arab countries such as Jordan, Syria,  
153 Lebanon, Yemen) and data in this study are consistent with this frequency. (18,19) We found that, Egyptian  
154 Patients with one or two copies of the T variant allele had 2.2 -fold increased lung cancer risk than those with  
155 CC genotype. However, the patient with one or two copies of the T variant allele had a 4.3-fold increased risk  
156 for developing small cell lung cancer, 3.8-fold increased risk to lung cancer other than squamous cell carcinoma  
157 or adenocarcinoma and 2.1-fold increased risk of adenocarcinoma more than those with CC genotype. Table  
158 ?? : Log-rank test comparing survival distributions among the three groups of the patients as regards NQO1  
159 609C?T genotype Our results are supported by Lewis et al., (13) who reported the increased risk associated with  
160 genotypes containing at least one variant T allele seems to be restricted to SCLC only among Caucasians. In  
161 studies performed in Asians, the NQO1 CC genotype was found to be associated with lung cancer, particularly  
162 adenocarcinoma, (20) whereas the variant NQO1 variant T allele has been suggested to be a risk factor for lung  
163 cancer in Caucasians. (9,13,21) The only previous study on Caucasians with sufficient number of cases that  
164 performed an analysis of histologic subtypes was in agreement with our study that the variant NQO1 genotypes  
165 were overrepresented in squamous cell carcinoma. (21) On the other side, in Asian population, the wildtype C

## 8 DISCUSSION

---

166 allele was found with higher incidence among subjects with adenocarcinoma. (12,20) Furthermore, the variant  
167 allele was found protective against adenocarcinoma in this population, while no such effect was observed in case of  
168 SCLC. (22) To explore the connection between NQO1 609C?T (rs1800566) polymorphism and lung cancer risks,  
169 we stratified the data by age, smoking and family history among our study subjects. We observed a statistically  
170 significant 10.6-fold increased risk among older individuals (age ?60) with even one variant T allele compared  
171 to 1.2-fold among younger individuals (age?60). However, the association between NQO1poly -morphism and  
172 lung cancer risk might differ depending on subject's age. Studies demonstrating the procarcinogenic effect of  
173 NQO1variant T allele in young Caucasians but rather protective effect in older ones can be found (age <50 years:  
174 OR = 1.28; age ?50 years: OR = 0.46). (23) In our study, the patients who were heavy smokers (?21 pack-years)  
175 and with one or two copies of the T variant allele had 17.9-fold increased risk for SCLC lung cancer than light  
176 smoker patients with 2.1fold increased risk. However, the overall lung cancer risk among heavy smokers (?21  
177 pack-years) 3.6-fold increased risk compared to 1.9-fold among light smokers (?21 pack-years).

178 Xu et al. (21) found that both the C/T and T/T genotype produced a higher risk of lung cancer compared with  
179 the wild-type genotype in those who smoked Polymerase chain reaction with confronting twopair primers (CTPP-  
180 PCR) is an effective genotyping method for single nucleotide polymorphisms (SNPs) in aspects of reducing time  
181 and costs for analysis. In the present study, the study subject's genotypes of NQO1 by CTPP-PCR method were  
182 the same as those genotyped with a RFLP-PCR.

183 Triplex PCR-RFLP for CYP1A1, GSTM1and GSTT1 polymorphisms has been reported by Bailey et al., (24)  
184 compared with PCR-RFLP, PCR-CTPP has the advantage of low cost and rapidity, because it allows genotyping  
185 of SNPs without incubation with a restriction enzyme for PCR product digestion. Multiplex PCR-CTPP is  
186 applicable; there is no doubt that it is superior to multiplex PCR-RFLP. PCR-CTPP needs less material input  
187 and time than PCR-RFLP, even for single polymorphism genotyping. (25) However, technical problems should  
188 be noted for PCR-CTPP. The strength of bands is dependent on the balance in melting temperature of each  
189 primer. The balance is also sensitive to annealing temperature of PCR. General speaking, a similar melting  
190 temperature for all primers provides the best chance to find an optimal primer set, so primers with a similar  
191 melting temperature have to be used. If a suitable primer set cannot be found, this method may not be applicable.  
192 This is a common problem to usual PCR with one pair of primers. ( ??4) Kawase et al., (25) reported these  
193 conditions after several unsuccessful combinations were tried.

Volume XIV Issue III Version I Year ( ) K <sup>1</sup>



Figure 1:

Overall Comparisons					
		Chi-Square	df	Sig.	
Log Rank (Mantel-Cox)		5.010	2	.057	
Test of equality of survival distributions for the different levels of Genotype.					

1

Figure 2: Figure I :

1

Genotype	patients (n=100)		Controls (n=100)		? 2	p
	No.	%	No.	%		
CC	62	62.0	73	73.0		
CT	31	31.0	26	26.0	5.835 *	MC p = 0.047 *
TT	7	7.0	1	1.0		
Allele						
C T	155 45	77.5 22.5	172 28	86.0 14.0	4.843 *	0.028 *
?						

[Note: 2 : value for Chi square \*: Statistically significant at p ? 0.05]

Figure 3: Table 1 :

2

	controls										a	
	Genotype					OR (95% CI)						
	No	CC	%	No	CT	%	No	TT	%	Crude		
Controls	73	54.1%		26	45.6%		1	12.5%		1	1	
All cases	62	45.9%		31	54.4%		7	87.5%		1.7 (0.91-3.0)	2.2 (0.63-7.9)	
SCC	19	20.7%		8	23.5%		4	80.0%		1.7 (0.73-3.9)	1.2 (0.21-7.0)	
SCLC	9	11.0%		10	27.8%		0			0.0% 0 (1.1-8.2)*	4.3 (0.533.4)	
Adeno-carcinoma	25	25.5%		10	27.8%		2	66.7%		1.3 (0.57-2.9)	2.1 (0.410.3)	
Others	9	11.0%		3	10.3%		1	50.0%		1.2 (0.34-4.2)	3.8 (0.430.7)	

a Odds ratios are for CT and TT versus CC.

# OR adjusted for age, family history, smoking index\* OR significant at 0.05

Figure 4: Table 2 :

## 8 DISCUSSION

---

3

	Genotype						Test	p of sig.
	CC (n = 62)		CT (n = 31)		TT (n = 7)			
	No	%	No	%	No	%		
Smoking								
Non smoker	0	62.0	2	27.2	6.5	25.0	28.6	18.737
Smoker			100.0		87.1		71.4	*
EX smoker		0.0			6.5		0.0	=
								0.001
								*
p 1				0.012 *			FE	p = 0.009 *
p 2							MC	p = 0.173
Smoking intensity								
Min. -Max.	0.50	-4.0	0.50	-3.0	1.50	-2.50		KW
Mean ± SD.	1.58	± 0.63	1.39	± 0.51	1.90	± 0.42	5.436	0.066
Median	1.50		1.50		2.0			
MW p 1			0.187		0.091			
MW p 2				0.025			*	
Duration of smoking								
(years)								
Min. -Max.	15.0	-50.0	5.0	-55.0	20.0	-40.0		KW
Mean ± SD.	28.60	± 9.99	27.93	± 13.0	31.0	± 7.42	0.832	0.367
Median	30.0		30.0		30.0			
MW p 1			0.935		0.499			
MW p 2				0.711				
Smoking Index (p/year)								
Min. -Max.	7.0	-100.0	2.50	-110.0	30.0	-100.0		KW
Mean ± SD.	43.37	± 17.28	41.91	± 26.20	60.50	± 25.27	0.238	2.873
Median	42.50		40.0		60.0			
MW p 1			0.680		0.105			
MW p 2				0.117				

p: p value for comparing between different genotype

p 1 : p value for comparing between CC with CT and TT

p 2 : p value for comparing between CT and TT

? 2 : value of Chi square

MC: Monte Carlo test

FE: Fisher Exact test

KW ? ? : Kruskal Wallis test

MW: Mann Whitney test

\*: Statistically significant at p ? 0.05

Figure 5: Table 3 :

---

4

	Smoking index <21				OR (95% CI)		Smoking index >21				OR (CI) Crude
	No	CC	% CT+TT	No %	Crude	Adjusted #	No	CC	% CT+TT	No %	
Controls	13	76.5%	5	45.5%	1	1	8	12.1%	1	3.7%	1
All cases	4	23.5%	6	54.5%	3.9 (0.76-19.9)	1.9 (0.29-13.2)	58	87.9%	26	96.3%	3.6 (30.1)
SCC	3	18.8%	2	28.6%	1.7 (0.22-13.7)	1.2 (0.05-18.6)	16	66.7%	6	85.7%	3.0 (29.4)
SCLC	1	7.1%	0	0.0%	2.4 (0.25-14.7)	2.1 (0.21-25.9)	8	50.0%	9	90.0%	9.0 (88.5)
Adenocarcinoma	0	0.0%	2	28.6%	3.7 (0.16-10.6)	1.3 (0.07-16.8)	25	75.8%	9	90.0%	2.9 (26.4)
Others	0	0.0%	2	28.6%	3.7 (0.16-10.6)	1.4 (0.05-17.5)	9	52.9%	2	66.7%	1.8 (23.5)

Figure 6: Table 4 :



195 [Xu et al.] , L L Xu , J C Wain , D P Miller , S W Thurston , L Su , T J Lynch . NAD (P) H: quinone  
196 Oxidoreductase 1.

197 [ Gene Polymorphism and Lung Cancer Differential Susceptibility Based on Smoking Behavior. *Cancer Epidemiology Biomarkers*  
198 , *Gene Polymorphism and Lung Cancer Differential Susceptibility Based on Smoking Behavior. Cancer*  
199 *Epidemiology Biomarkers & Prevention* 2001. 10 (4) p. .

200 [Jemal et al. ()] , A Jemal , F Bray , M M Center , J Ferlay , E Ward , D Forman . *Global cancer statistics. CA: a cancer journal for clinicians* 2011. 61 (2) p. .

202 [Yu et al. ()] 'A functional NQO1 609C> T polymorphism and risk of gastrointestinal cancers: a meta-analysis'.  
203 H Yu , H Liu , L-E Wang , Q Wei . *PLOS ONE* 2012. 7 (1) p. e30566.

204 [Lin et al. ()] 'Analysis of NQO1, GSTP1, and MnSOD genetic polymorphisms on lung cancer risk in Taiwan'.  
205 P Lin , Y-M Hsueh , J-L Ko , Y-F Liang , K-J Tsai , C-Y Chen . *Lung Cancer* 2003. 40 (2) p. .

206 [Bailey et al. ()] 'Breast cancer and CYP1A1, GSTM1, and GSTT1 polymorphisms: evidence of a lack of  
207 association in Caucasians and African Americans'. L R Bailey , N Roodi , C S Verrier , C J Yee , W D  
208 Dupont , F F Parl . *Cancer Research* 1998. 58 (1) p. .

209 [Gasdaska et al. ()] 'Cigarette smoking is a determinant of DTdiaphorase gene expression in human non-small  
210 cell lung carcinoma'. P Y Gasdaska , G Powis , P Hyman , H Fisher . *Cancer Research* 1993. 53 (22) p. .

211 [Hamajima et al. ()] 'Competitive amplification and unspecific amplification in polymerase chain reaction with  
212 confronting two-pair primers'. N Hamajima , T Saito , K Matsuo , K Tajima . *The Journal of molecular*  
213 *diagnostics* 2002. 4 (2) p. .

214 [Sunaga et al. ()] 'Contribution of the NQO1 and GSTT1 polymorphisms to lung adenocarcinoma susceptibility'.  
215 N Sunaga , T Kohno , N Yanagitani , H Sugimura , H Kunitoh , T Tamura . *Cancer Epidemiology Biomarkers*  
216 *& Prevention* 2002. 11 (8) p. .

217 [Jemal et al. ()] 'Global patterns of cancer incidence and mortality rates and trends'. A Jemal , M M Center , C  
218 Desantis , E M Ward . *Cancer Epidemiology Biomarkers & Prevention* 2010. 19 (8) p. .

219 [Jaiswal ()] 'Human NAD (P) H: quinone oxidoreductase (NQO1) gene structure and induction by dioxin'. A K  
220 Jaiswal . *Biochemistry* 1991. 30 (44) p. .

221 [Rosvold et al. ()] 'Identification of an NAD (P) H: quinone oxidoreductase polymorphism and its association  
222 with lung cancer and smoking'. E A Rosvold , K A Mcglynn , E D Lustbader , K H Buetow . *Pharmacogenetics*  
223 1995. 5 (4) p. 199.

224 [Dela et al. ()] 'Lung cancer: epidemiology, etiology, and prevention'. Ccs Dela , L T Tanoue , R A Matthay .  
225 *Clinics in chest medicine* 2011. 32 (4) p. 605.

226 [Kawai et al. ()] 'Multiplex PCR with confronting twopair primers for CYP1A1 Ile462Val, GSTM1, GSTT1, and  
227 NQO1 C609T'. S Kawai , K Nishio , S Nakamura , Y Sekido , T Niwa , N Hamajima . *Asian Pacific Journal*  
228 *of Cancer Prevention* 2005. 6 (3) p. 346.

229 [Nebert et al. ()] 'NAD (P) H: quinone oxidoreductase (NQO1) polymorphism, exposure to benzene, and  
230 predisposition to disease: a HuGE review'. D W Nebert , A L Roe , S E Vandale , E Bingham , G G  
231 Oakley . *Genetics in Medicine* 2002. 4 (2) p. .

232 [Chao et al. ()] 'NAD (P) H: quinone oxidoreductase 1 (NQO1) Pro187Ser polymorphism and the risk of lung,  
233 bladder, and colorectal cancers: a meta-analysis'. C Chao , Z-F Zhang , J Berthiller , P Boffetta , M Hashibe  
234 . *Cancer Epidemiology Biomarkers & Prevention* 2006. 15 (5) p. .

235 [Lafuente et al. ()] 'NAD (P) H: quinone oxidoreductase-dependent risk for colorectal cancer and its association  
236 with the presence of K-ras mutations in tumors'. M J Lafuente , X Casterad , M Trias , C Ascaso , R Molina  
237 , A Ballesta . *Carcinogenesis* 2000. 21 (10) p. .

238 [Joseph and Jaiswal ()] 'NAD (P) H: quinone oxidoreductase1 (DT diaphorase) specifically prevents the formation  
239 of benzo [a] pyrene quinone-DNA adducts generated by cytochrome P4501A1 and P450 reductase'. P  
240 Joseph , A K Jaiswal . *Proceedings of the National Academy of Sciences* 1994. 91 (18) p. .

241 [Bock et al. ()] 'NQO1 T allele associated with decreased risk of later age at diagnosis lung cancer among never  
242 smokers: results from a population-based study'. C H Bock , A S Wenzlaff , M L Cote , S J Land , A G  
243 Schwartz . *Carcinogenesis* 2005. 26 (2) p. .

244 [Spivack et al. ()] 'Phase I and II carcinogen metabolism gene expression in human lung tissue and tumors'. S  
245 D Spivack , G J Hurteau , M J Fasco , L S Kaminsky . *Clinical Cancer Research* 2003. 9 (16) p. .

246 [Lewis et al. ()] 'Polymorphisms in theNAD( P)H: quinone oxidoreductase gene and small cell lung cancer risk  
247 in a UK population'. S J Lewis , N M Cherry , R M Niven , P V Barber , A C Povey . *Lung Cancer* 2001.  
248 34 (2) p. .

249 [Shopland ()] *Tobacco use and its contribution to early cancer mortality with a special emphasis on cigarette*  
250 *smoking. Environmental health perspectives*, D R Shopland . 1995. 103 p. 131. (Suppl)

## 8 DISCUSSION

---

251 [Kawase et al. ()] 'Triplex Polymerase Chain Reactions with Confronting Two-Pair Primers (PCR-CTPP) for  
252 NQO1C609T, GSTM1and GSTT1Polymorphisms: a Convenient Genotyping Method'. H Kawase , Nh , At ,  
253 T S Kw , KT . *Asian Pacific J Cancer Prev* 2003. 4 p. .

254 [Bu et al. ()] 'Variable drug metabolism genes in Arab population'. R Bu , M I Gutierrez , M Al-Rasheed , A  
255 Belgaumi , K Bhatia . *The pharmacogenomics journal* 2004. 4 (4) p. .