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Volume XIV Issue VII Version I Bidur Arval¹ ¹ ST. Xaviers College, Maitighar, Kathmandu Received: 7 December 2013 Accepted: 4 January 2014 Published: 15 January 2014 Abstract 6 Background: The present study analyzes the clinical profile, identifies the pathogenic distribution and their antimicrobial susceptibility pattern in childhood urinary tract infections 8 in order to provide standard reference for the optimal use of antibiotics in Nepal.Methods: A hospital based cross section study was conducted among children suspected of urinary tract

10 infection in Kanti Children's Hospital over a period of six months from August 2012 to

11 November 2012. A total of 1890both sexes, ranging from post natal period to 14 years of age 12

were studied. The modes of presentation, laboratory investigation reports, which included 13

urine routine microscopy, bacterial isolation with colony count from urine culture, antibiotic 14

sensitivity pattern and multidrug resistant profile, were documented. Data were analyzed by 15

the Chi Square Test.Results: Among 1890 urine samples, 300(15.88 16

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Index terms-18

1 Introduction 19

rinary tract infection (UTI) is common in pediatric practice and an important cause of morbidity and mortality in 20 children. However, UTI is a common problem throughout the world, the microbial isolates and their sensitivity 21 22 pattern need to be analyzed at regular interval to monitor the changing pattern of microbial flora and the 23 development of resistance to drugs, which may help the physician to treat UTI in better way and to prevent 24 further complications.

$\mathbf{2}$ II. 25

3 Materials and Methods 26

We conducted the prospective analysis of the cases attending pediatric OPD and those admitted in the ward of 27 Kanti Children's Hospital, Kathmandu, Nepal. Study period was six months from August 2012 to November 28 2012. Children of both sexes up to the age of 14 years were included in the study. Their clinical presentation with 29 associated condition and risk factors were noted. Approximately 1890 urine samples were screened and 300 urine 30 31 samples showed positive culture result. Parents were explained about the study and professional care was taken 32 to collect the urine sample for routine culture and sensitivity by sterile technique. Urine was sampled for culture 33 by aseptic supra pubic bladder aspiration in infants. Sterile plastic receptacles were used for collection of urine in younger patients to avoid contamination with stool. Clean catch mid-stream urine was sampled in older children 34 and adolescents after proper cleansing of urethra and under supervision. The samples were than processed for 35 routine microscopy. Only samples with more than 5 WBC per high power field (hpf) were subjected for culture 36 and antimicrobial susceptibility testing in the bacteriology laboratory of Kanti Children Hospital. Receptacle 37 sample and mid-stream urine sample with culture with >10 5 colony forming units of bacteria/ml of urine in 38 young infants and adolescents. Any colony count with supra pubic count in infants. 39

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41 5 Observation and Result

Among the 1890 urine samples included in the study, 300(15.88%) showed positive culture result (fig. 1). Since, the study includes newly born babies up to 14 years of age. The high frequency of UTI was found in 0-2 years of age followed by 8-10 years of age. Among the 300 culture positive cases, 114(38%) were males and 186(62%) were females (Table 1). Among the antibiotics used, Nitrofurantoin was found to have the highest sensitivity (71.67%) amongst most bacteria. Amikacin, Norfloxacin and Gentamicin had sensitivity of 69%, 61.71% and 61.67% respectively.

Pseudomonas aeruginosa was 100% sensitive to Tobramycin, Piperacillin and Imipenem. Though sensitivity
to Vancomycin was tested to 44 cases and it showed no resistance and it was 100% sensitive to Staphylococcus
aureus and Streptococcus fecalis.

Highest degree of resistance was noted with Ceftazidime(64%), Ofloxacin(61.33%), Ampicillin(60%)

52 ciprofloxacin (55.67%), Cotrimoxazole (52%), Gentamicin (38.33%), Amikacin (28%) and Nitrofurantoin

(23.67%). The sensitivity pattern of various organisms was also studied. E.coli responded better with Ni trofurantoin, Aminoglycosides and Fluroquinolones but displayed a highresistance with most of thebeta lactams.

55 Resitance was also noted with Ofloxacin, Nalidixic acid and Ciprofloxacin. IV.

56 6 Discussion

Urinary tract infection (UTI) is common cause of febrile illness in young children. In the first year of life. Urinary 57 tract infection(UTI) is one of the most important causes of morbidity in the general population and the second 58 most common cause of hospital visits ??Das et al., 2006). Urinary tract infection(UTI) is not uncommon cause 59 of bacterial illness in children, 4-8% of children have had an UTI from a population-based study (Suresh kumaret 60 61 al., 2009). The prevalence and incidence of is higher in female than in male children, which are likely the result of 62 several clinical factors including anatomical differences, hormonal effects and behavior pattern (Griebling, 2009). The prevalence of UTIs is quite different between two gender and age with high incidence in girls (1% in male 63 and 3% in female), except the male infants with an incidence of 0.7% compared to the 0.1-0.4% of female infants 64 (Foxman, 2002), which is due to bacteria harboring in prepuce of young infants. 65 Among the growth positive samples, 144(48%) were male patients and 156(52%) were female patients. Among

66 1890 urine samples, 1094(57.88%) were symptomatic, in which 166(15.18%) was culture positive. Urinary 67 symptoms like dysuria, burning urine, increased frequency, haematuria, oliguria, bed wetting, chills and rigors, 68 abdominal pain, vomiting, loose stool, etc. The first and the most critical step in establishing the diagnosis 69 70 of UTI in infants and young children is the method by which the urine is collected. In the young infants 71 care must be taken to prepare carefully the perineum and periurethral area for placement of sterile plastic 72 receptacle for collection of urine. In the infants, the purest way to obtain urine for culture aseptically is by precutaneoussuprapubic aspiration. Older children and adolescents can be instructed to collect the midstream 73 74 urine specimen after proper cleansing of the urethral area. These steps were strictly followed for collection of urine in our study. The presence of 105 organisms or more per ml of urine is diagnostic of UTI. If 103 to 105 75 colony forming units of a single genus and species per ml are recovered from two successive urine culture of a 76 child, a diagnosis of UTI should be made6. 77

In our context, such cases were not included in our study as it was difficult to call the patient for repeated
urine culture though they were empirically (2012); Beyene and Tsegaye (2011); ??aoud 1%.Yet in another study,
the findings were consistent with ours where the pathogens were E.coli (47%), Klebsiellaspp (18%), S.aureus
(13.4%), Proteus spp (9%), E.fecalis (5.3%), P.aeruginosa (5%), and others 2.3%.

In our study,Nitrofurantoin was found to have the highest sensitivity (71.67%) amongst most bacteriawhereas Proteus, Salmonella paratyphi B and P. aeruginosa was resistant to the same.Amikacin, Norfloxacin and Gentamicin had sensitivity of 69%, 6

1.71% and 61.67% respectively. Pseudomonas aeruginosa was 100% sensitive to Tobramycin, Piperacillin and
Imipenem. Though sensitivity to Vancomycin was tested to 44 cases it was 100% sensitive to Staphylococcus
aureus and Streptococcus fecalis. Highest degree of resistance was noted with Ceftazidime (64%), Ofloxacin
(61.33%), Ampicillin (60%), Ciprofloxacin (55.67%), Cotrimoxazole (52%), Gentamicin (38.33%), Amikacin(28%)
and Nitrofurantoin (23.67%).

In this study, 69% (207/300) were found to be Multidrug resistant (MDR) i.e. they were resistant to treated as
suspected UTI. The suprapubic bladder aspiration or by catheterization contain fewer than 105 organisms because
the organisms have not had sufficient time to multiply before the removal of urine from the bladder(Griebling
TL.,2009).

In this study, 300(15.88%) resulted a positive culture in urine with significant colony count of ?105rest 1590(84.12%) were culture negative or they had colony count <105 (fig. 1). E.coli(52.33%) was found to be predominant organism in this study which resembles with the study done by Raiet al., (2008); Maliangoet al., more than two drugs which is similar to the result of Pokhrelet al., 2006 in which 60.40% were MDR. The MDR in E.coli was found to be 66.87% (105/300). Although multidrug resistance was shown 100% by P. aeruginosa,Enterococcus fecalis and S. parathypi B, these were low in number and considered insignificant. In a study done by Tuladharet al., 2003 at TUTH, MDR bacterial strains were detected in 35.2% cases in which the
 most predominant was E.coli(22.2%) followed by Klebsiellaspp (6.1%) and Staphylococcus aureus (2.2%).

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103 8 Conclusion

As UTI is the significant problem in the children and still continues to be a major threat for morbidity and mortality in subtropical parts of the world, larger scale studies must be carried out at a regular intervals in order to identify the changing trend in the pathogenic organisms and update on its changing antibiotic susceptibility. Based on the sensitivity patterns we recommend empirical use of Nitrofurantoin, Amikacin, Norfloxacin and

Gentamicin for patients with UTI.Vancomycin showed 100% sensitivity to grampositive bacteria. Gram-negative

- bacteria like Proteus spp, P. aeruginosa and S.paratyphi B was 100% resistant to Nitrofurantoin. P.aeruginosa
- 110 was 100% resistant to Imipenem, Piperacilin and Tobramycin. So, Vancomycin should be kept as reserve drug for gram positive organisms and Tobramycin, Imipenem and Piperacilin for P. aeruginosa.



Figure 1: Figure 1 :

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 \mathbf{Sex} 0-2-4-Age(years) $\mathbf{2}$ 4 6-8 6 8-10 Male 62 71710 Female90 312513Total 152 48 3520(14%) followed by Streptococcus fecalis (0.66%)Among the gram negative isolates, the most E. organisms isolatedvas common coli(52.33%) followed by Klebsiella pneumonia (16%), Pseudomonas aeruginosa (4.33%), Proteus mirabilis (2.66%), Citrobacter freundii (2%), Acinetobacter spp.(1.66%), Klebsiella oxytoca, Enterobacter spp., Hafniaalvei (1.33%), Proteus vulgaris, Citrobacter koseri (1%)and Salmonella paratyphi B (0.33%). Staphylococcus aureus (14%) followed by Streptococcus fecalis (0.66%) of cases. (fig 2).

Figure 2: Table 1 :

Culture Result 90.00%84.12% 80.00%70.00%60.00%50.00%40.00%Culture Result 30.00%20.00%15.88%10.00% 0.00%Growth No growth

Figure 3: Table 2 :

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